

# Supplemental data

## Table of contents

Abbreviations	Page 1
<b>Methods</b>	
Search strategy	Page 2-3
PubMed search	Page 2
Embase search	Page 3
Explanation of the method used for classification of biomarkers based on the biomarker-specific figures	Page 4
<b>Results</b>	
Table S1: Baseline characteristics of included studies	Page 5-12
Table S2: Biomarker classification	Page 13-34
Biomarker-specific figures and meta-analyses of biomarkers studied by more than one study:	
<u>Clinical DCI</u>	Page 35-45
Figure S1    ADAM Metallopeptidase with Thrombospondin Type 1 Motif 13 (blood)	Page 35
Figure S1.1    Meta-analyses of two of three studies	Page 35
Figure S2    Albumin (blood)	Page 35
Figure S3    C-C Motif Chemokine Ligand 5 (blood)	Page 35
Figure S4    Soluble CD40 ligand (blood)	Page 35
Figure S5    Cortisol (blood)	Page 36
Figure S6    C-reactive protein (blood)	Page 36
Figure S7 <i>high-sensitivity</i> C-reactive protein (blood)	Page 36
Figure S8    D-dimer (blood)	Page 36

Figure S8.1	Meta-analysis of two of seven studies	Page 36
Figure S9	Endothelin-1 (blood)	Page 36
Figure S10	Endothelin-1 (cerebrospinal fluid)	Page 37
Figure S11	Endothelin-1 SNP rs1800541	Page 37
Figure S12	Endothelin-1 SNP rs2070699	Page 37
Figure S13	E-selectin (blood)	Page 37
Figure S14	F2-isoprostane (urine)	Page 37
Figure S15	Fibrinogen (blood)	Page 37
Figure S16	Ficolin-1 (blood)	Page 37
Figure S17	Ficolin-2 (blood)	Page 37
Figure S18	Ficolin-3 (blood)	Page 38
Figure S19	Glucose (blood)	Page 38
Figure S20	Haptoglobin 1-1 polymorphism (meta-analysis)	Page 38
Figure S21	Haptoglobin 2-1 polymorphism (meta-analysis)	Page 38
Figure S22	Haptoglobin 2-2 polymorphism (meta-analysis)	Page 39
Figure S23	Hematocrit (blood)	Page 39
Figure S24	Hemoglobin (blood)	Page 39
Figure S25	<i>glycated</i> Hemoglobin (blood)	Page 39
Figure S26	Intercellular Adhesion Molecule 1 (blood)	Page 39
Figure S27	Interferon gamma (blood)	Page 40
Figure S28	Interleukin-1B (blood)	Page 40
Figure S29	Interleukin-6 (blood)	Page 40
Figure S30	Interleukin-6 (cerebrospinal fluid)	Page 40
Figure S31	Interleukin-8 (blood)	Page 40
Figure S32	Lactate (microdialysate)	Page 40
Figure S33	Lactate/pyruvate ratio (microdialysate)	Page 40
Figure S34	Leukocytes (blood)	Page 41

Figure S35	L-selectin (blood)	Page 41
Figure S36	Lymphocytes (blood)	Page 41
Figure S37	Magnesium (blood)	Page 41
Figure S38	Mannose-binding lectin (blood)	Page 42
Figure S39	Mean platelet volume (blood)	Page 42
Figure S40	Mean platelet volume/platelet count ratio (blood)	Page 42
Figure S41	Neutrophils (blood)	Page 42
Figure S42	Neutrophil/lymphocyte ratio (blood)	Page 42
Figure S43	<i>endothelial</i> Nitric oxide synthase SNP rs2070744 T-786C	Page 42
Figure S44	Platelet count (blood)	Page 43
Figure S45	Prothrombin time (blood)	Page 43
Figure S46	P-selectin (blood)	Page 43
Figure S47	s-100B (blood)	Page 43
Figure S48	Sodium (urine)	Page 43
Figure S49	Sodium (blood)	Page 44
Figure S50	Tau protein (cerebrospinal fluid)	Page 44
Figure S51	Thrombin-antithrombin III complex (blood)	Page 44
Figure S52	Thrombospondin-1 (blood)	Page 44
Figure S53	Tumor necrosis factor a (blood)	Page 44
Figure S54	Tumor necrosis factor a (cerebrospinal fluid)	Page 44
Figure S55	Transferrin (cerebrospinal fluid)	Page 44
Figure S56	Vascular cell adhesion protein 1 (blood)	Page 45
Figure S57	Von Willebrand factor (blood)	Page 45
	Figure S57.1 Meta-analysis of two of four studies	Page 45
Figure S58	Von Willebrand factor propeptide (blood)	Page 45

Figure S59	Big endothelin-1 (blood)	Page 46
Figure S60	Brain natriuretic peptide (blood)	Page 46
Figure S61	Carbon dioxide partial pressure (blood)	Page 46
Figure S62	<i>total</i> Cholesterol (blood)	Page 46
Figure S63	C-reactive protein (blood)	Page 46
Figure S64	D-dimer (blood)	Page 46
Figure S65	<i>soluble</i> Endoglin (blood)	Page 47
Figure S66	Endothelin-1 (blood)	Page 47
Figure S67	Glucose (blood)	Page 47
Figure S68	Hemoglobin (blood)	Page 47
Figure S69	Lactate (blood)	Page 47
Figure S70	Lactate/pyruvate ratio (microdialysate)	Page 47
Figure S71	Leukocytes (blood)	Page 48
Figure S72	Neutrophil/lymphocyte ratio (blood)	Page 48
Figure S73	Platelet count (blood)	Page 48
Figure S74	Potassium (blood)	Page 48
Figure S75	Procalcitonin (blood)	Page 48
Figure S76	Sodium (blood)	Page 48
Figure S77	Triglycerides (blood)	Page 48
References		Page 49-65

## Abbreviations

CSF: cerebrospinal fluid

DCI: delayed cerebral ischemia

h: hours

MD: microdialysate

POD: post-operative day

SAH: subarachnoid hemorrhage

## Search strategy

PubMed:

#	Searches	Results
1	Validat\$.tw. or Predict\$.ti. or Rule\$.tw. or (Predict\$ and (Outcome\$ or Risk\$ or Model\$)).tw. or ((History or Variable\$ or Criteria or Scor\$ or Characteristic\$ or Finding\$ or Factor\$) and (Predict\$ or Model\$ or Decision\$ or Identif\$ or Prognos\$)).tw. or (Decision\$.tw. and ((Model\$ or Clinical\$).tw. or logistic models/)) or (Prognostic and (History or Variable\$ or Criteria or Scor\$ or Characteristic\$ or Finding\$ or Factor\$ or Model\$)).tw. or ("Stratification" or "Discrimination" or "Discriminate" or "c-statistic" or "c statistic" or "Area under the curve" or "AUC" or "Calibration" or "Indices" or "Algorithm" or "Multivariable").tw.	5050668
2	exp VASOSPASM, INTRACRANIAL/ or ((Delayed or Secondary or symptomatic) adj2 (Ischemia or infarc*)).ti,ab,kf. or ((intracranial or cerebral or cerebrovascular) adj (vasospasm* or angiospasm* or vascular spasm*)).ti,ab,kf. or ((Delayed or Secondary) adj3 (Ischemic or neurological) adj Deficit*).ti,ab,kf. or (DCI or Vasospasm).ti,ab,kf.	18111
3	1 or 2	5064922
4	exp Biomarkers/ or exp Peptides/ or exp "Genetic Phenomena"/ or exp Genes/ or exp Proteins/ or exp "Polymorphism, Single Nucleotide"/ or exp "Genome-Wide Association Study"/ or exp MicroRNAs/ or exp "Sequence Analysis, DNA"/ or exp DNA/ or (biomarker* or Marker* or Surrogate Endpoint* or Polypeptides or Neuropeptide or (Genetic adj (Concept* or Phenomenon* or Process* or material*)) or Gene or Genes or Cistron* or SNP or SNPs or "Single Nucleotide Polymorphism" or MicroRNA or miRNA* or stRNA or "Small Temporal RNA" or Genome-Wide Association or (Sequence adj (Analys?s or determinaion*)) or Sequencing or Sequence or "Deoxyribonucleic Acid" or ds-DNA).ti,ab,kf.	9819019
5	exp Cerebrospinal Fluid/ or exp CYTOKINES/ or exp C-Reactive Protein/ or exp SEROLOGY/ or exp Feces/ or exp urine/ or (micro-particle* or microparticle* or spinal fluid or cerebrospinal fluid or serology or serum or plasma or urine or stool or crp or cytokine* or interleukin* or adhesion).ti,ab,kf. or (level* or levels or concentration or value).ti,ab.	7835561
6	4 or 5	13711807
7	exp Subarachnoid Hemorrhage/ or (SAH or (subarachnoid adj (h?emorrhage or h?ematoma or bleeding))).ti,ab,kf.	34848
8	6 and 7	11317
9	3 and 8	5607
10	(exp animals/ not humans/) or (mice or mouse or rat or rats or rodent or canine or dog or rabbit or dogs or murine or cat or cats or rabbits or rodents or canines).ti.	5204407
11	9 not 10	4209

Embase:

#	Searches	Results
1	Validat\$.mp. or Predict\$.ti. or Rule\$.mp. or (Predict\$ and (Outcome\$ or Risk\$ or Model\$)).tw. or ((History or Variable\$ or Criteria or Scor\$ or Characteristic\$ or Finding\$ or Factor\$) and (Predict\$ or Model\$ or Decision\$ or Identif\$ or Prognos\$)).tw. or (Decision\$.tw. and ((Model\$ or Clinical\$).tw. or statistical model/)) or (Prognostic and (History or Variable\$ or Criteria or Scor\$ or Characteristic\$ or Finding\$ or Factor\$ or Model\$)).tw. or ("Stratification" or "Discrimination" or "Discriminate" or "c-statistic" or "c statistic" or "Area under the curve" or "AUC" or "Calibration" or "Indices" or "Algorithm" or "Multivariable").tw. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword heading word, floating subheading word, candidate term word]	6992976
2	exp brain vasospasm/ or ((Delayed or Secondary or symptomatic) adj2 (Ischemia or infarc*)).ti,ab,kw. or ((intracranial or cerebral or cerebrovascular) adj (vasospasm* or angiospasm* or vascular spasm*)).ti,ab,kw. or ((Delayed or Secondary) adj3 (Ischemic or neurological) adj Deficit*).ti,ab,kw. or (DCI or Vasospasm).ti,ab,kw.	28604
3	1 or 2	7014925
4	exp biological marker/ or exp peptide/ or exp heredity/ or exp gene/ or exp protein/ or exp single nucleotide polymorphism/ or exp genome-wide association study/ or exp genetic association/ or exp microRNA/ or exp circulating microRNA/ or exp sequence analysis/ or exp DNA/ or (biomarker* or Marker* or Surrogate Endpoint* or Polypeptides or Neuropeptide or (Genetic adj (Concept* or Phenomenon* or Process* or material*)) or Gene or Genes or Cistron* or SNP or SNPs or "Single Nucleotide Polymorphism" or MicroRNA or miRNA* or stRNA or "Small Temporal RNA" or Genome-Wide Association or (Sequence adj (Analys?s or determinaion*)) or Sequencing or Sequence or "Deoxyribonucleic Acid" or ds-DNA).ti,ab,kw.	7506594
5	exp cerebrospinal fluid/ or exp cytokine/ or exp C reactive protein/ or exp serology/ or exp feces/ or exp feces analysis/ or exp urine/ or (micro-particle* or microparticle* or spinal fluid or cerebrospinal fluid or serology or serum or plasma or urine or stool or crp or cytokine* or interleukin* or adhesion).ti,ab,kw. or (level* or levels or concentration).ti,ab.	10030196
6	4 or 5	14546244
7	exp Subarachnoid Hemorrhage/ or (SAH or (subarachnoid adj (h?emorrhage or h?ematoma or bleeding))).ti,ab,kw.	57415
8	6 and 7	16360
9	3 and 8	7769
10	((exp experimental organism/ or animal tissue/ or animal cell/ or exp animal disease/ or exp carnivore disease/ or exp bird/ or exp experimental animal welfare/ or exp animal husbandry/ or animal behavior/ or exp animal cell culture/ or exp mammalian disease/ or exp mammal/ or exp marine species/ or nonhuman/ or animal.hw.) not human/) or (mice or mouse or rat or rats or rodent or canine or dog or rabbit or dogs or murine or cat or cats or rabbits or rodents or canines).ti.	7923711
11	9 not 10	6182
12	limit 11 to embase	4135

## Methods used for classification of biomarkers which are measured by more than one study.

If a biomarker was investigated by more than one study, the classification into significant and non-significant was either based on a biomarker-specific meta-analysis, if methodologically possible (similar type of analyses and timing of measurement), or by descriptive comparison.

Meta-analyses were performed using Review Manager 5.4. For continuous data we used the inverse variance method with random effects model to calculate weighted mean differences with 95% confidence intervals. For dichotomous data we used the Mantel-Haenszel method with random effects model to calculate estimated odds ratio with 95% confidence intervals.

Descriptive comparison of the studies was done by summarizing the results of all relevant studies in a biomarker-specific figure, which was then used to classify the biomarkers into significant, inconclusive and non-significant.

The biomarker-specific figures display the author, year, number of patients with and without DCI, the type of analyses and the results. Results were divided into categories of sampling timing (day 0-3, 4-8, 9-13 and >14 after SAH). Results could also cover more than one time period (e.g. repeated measures).

Classification into significant, inconclusive and non-significant was done as follows:

- 1) If all results within the biomarker-specific figure were **significant**, the biomarker was considered significant. Similarly, biomarkers with exclusively non-significant results were considered **non-significant**.
- 2) In case the biomarker-figure contained both significant and non-significant results, we evaluated the results within each time intervals (day 0-3, 4-8, 9-13 and >14 after SAH) to determine whether a biomarker was deemed significant.
  - a. A time interval was considered non-significant if the results of all studies within that time interval showed non-significant results
  - b. A time interval was considered significant if the results of all studies within that time interval showed significant results, pointing in the same direction (e.g. all studies show higher biomarker levels in DCI compared to no DCI patients).
  - c. In case of both significant and non-significant results, the time interval was considered inconclusive.
  - d. In case of multiple significant results, however the results point in opposite directions (study A shows higher biomarker levels, whereas study B shows lower levels in DCI), the time interval was considered inconclusive.
  - e. As long as at least one time interval was considered significant, the biomarker was deemed **significant**. Otherwise, the biomarker was classified as **inconclusive**.



Table S1 - Baseline characteristics of included studies

1 <sup>st</sup> author, year	Method <sup>1</sup>	Study Design	SAH n	Age <sup>2</sup> mean (SD)	Female n (%)	Poor grade <sup>3</sup> n (%)	Fisher 3-4 <sup>4</sup> n(%)	DCI n (%)	DCI definition <sup>5</sup>	Risk of bias
Cohen, 2021 <sup>1</sup>	Pro	Cohort	356	57 (13)	241 (68)	WFNS 134 (38)	-	142* (40)	C	4/8
Albanna, 2021 <sup>2</sup>	Pro	Cohort	66	55 (1)	51 (77)	WFNS: 18 (27)	mF: 34 (52)	33 (50) 9 (14)	C R	6/9
Helbok, 2021 <sup>3</sup>	Pro	Cohort	36	57 (48-67)	26 (72)	HH: 26 (72)	33 (91)	9* (25)	R	4/8
Veldeman, 2021 <sup>4</sup>	Pro	Cohort	30	54 (14)	17 (57)	HH: 7 (23)	mF: 16 (53)	13 (43)	C	4/8
Krenzlin, 2021 <sup>5</sup>	Pro	Cohort	29	55 (12)	19 (66)	mean HH: 3	mean: 4	13 (45)	C	2/9
Darkwah, 2021 <sup>6</sup>	Retro	Cohort	633	55 (14)	422 (67)	WFNS: 300 (47)	mF: 538 (91)	153 (24)	R	7/8
Park, 2021 <sup>7</sup>	Retro	Cohort	510	56	348 (68)	HH: 83 (16)	-	47* (9)	C	4/8
Zhang, 2021 <sup>8</sup>	Retro	Cohort	439	-	304 (69)	WFNS: 93 (21)	mF: 123 (28)	84 (24)	C	7/8
Veldeman, 2021 <sup>9</sup>	Retro	Cohort	24	59 (12)	19 (79)	HH: 10 (42)	mF: 18 (75)	14 (58)	C	2/8
Neumaier, 2021 <sup>10</sup>	-	Cohort	96	54 (12)	71 (74)	HH: 21 (22)	mF: 55 (57)	48 (50)	C	5/8
Heinsberg, 2020 <sup>11</sup>	Pro	Cohort	591	53 (11)	423 (72)	WFNS: 142 (24)	350 (59)	103* (34)	C	5/8
Heinsberg, 2020 <sup>12</sup>	Pro	Cohort	474	54	343 (72)	WFNS: 111 (23)	272 (57)	179* (39)	C	6/8
Luo, 2020 <sup>13</sup>	Pro	Cohort	418	58 (49-66)	242 (58)	HH: 127 (30)	mF: 217 (52)	89 (21)	C	6/9
Veldeman, 2020 <sup>14</sup>	Pro	Cohort	132	56 (12)	91 (96)	HH: 31 (24)	mF: 80 (61)	64 (46)	C	5/9
Chaudhry, 2020 <sup>15</sup>	Pro	Cohort	80	58 (12)	49 (61)	HH: 27 (34)	77 (96)	28* (35)	C	5/9
Chaudhry, 2020 <sup>16</sup>	Pro	Cohort	76	59 (12)	46 (61)	HH: 24 (31)	73 (96)	16* (21)	R	3/9
Kim, 2020 <sup>17</sup>	Pro	Cohort	76	56	50 (66)	HH: 27(36)	59 (78)	25 (33)	C	3/8
Bache, 2020 <sup>18</sup>	Pro	Cohort	63	59 (11)	49 (78)	WFNS: 40 (63)	61 (97)	17* (49)	C	5/9
Wang, 2020 <sup>19</sup>	Pro	Cohort	61	55 (6)	36 (59)	WFNS: 15 (25)	19 (31)	19 (31)	C	7/9
Park, 2020 <sup>20</sup>	Pro	Cohort	52	61	26 (50)	HH: 23 (44)	26 (50)	18 (52)	C	4/8
Kofler, 2020 <sup>21</sup>	Pro	Cohort	51	59 (50-67)	36 (71)	HH: 35 (69)	mF: 46 (90)	16 (31)	R	4/8
Wisniewski, 2020 <sup>22</sup>	Pro	Cohort	38	59	24 (63)	HH: 0	26 (68)	19 (50)	C	6/9
Rzeplinski, 2020 <sup>23</sup>	Pro	Cohort	37	56 (40-67)	22 (60)	WFNS: 14 (38)	mF: 24 (65)	8 (22)	C	6/8
Youn, 2020 <sup>24</sup>	Pro	Cohort	33	61	15 (45)	HH3-5: 17 (52)	-	12 (36)	C	2/8
Manner, 2020 <sup>25</sup>	Pro	Case-control	18	57 (13)	9 (50)	WFNS: 12 (67)	16 (89)	7 (44)	R	4/9
Torne, 2020 <sup>26</sup>	Pro	Cohort	16	58 (7)	-	WFNS: 16 (100)	15 (94)	5 (31)	R	2/8
Roa, 2020 <sup>27</sup>	Pro	Cohort	13	56	8 (62)	WFNS: 7 (54)	mF: 9(69)	5 (39)	C	3/8

Liu, 2020 <sup>28</sup>	Retro	Cohort	887	-	562 (63)	HH: 123 (14)	mF: 158 (18)	224 (25)	C	6/8
Arleth, 2020 <sup>29</sup>	Retro	Cohort	384	56 (48-64)	275 (72)	WFNS3-5: 163 (44)	-	102* (35)	C	5/8
Chen, 2020 <sup>30</sup>	Retro	Cohort	197	60 (51-67)	114 (58)	HH: 49 (25)	mF: 135 (69)	58 (29)	C	6/8
Hurth, 2020 <sup>31</sup>	Retro	Cohort	138	53	104 (75)	WFNS: 62 (45)	119 (86)	27 (20)	R	5/8
Li, 2020 <sup>32</sup>	Retro	Cohort	75	59 (2)	50 (67)	HH: 21 (28)	mean: 4	37 (40)	R	5/8
Xie, 2020 <sup>33</sup>	Retro	Cohort	66	58 (10)	47 (71)	HH: 66 (100)	-	26 (39)	R	6/8
Matzen, 2020 <sup>34</sup>	Retro	Cohort	63	59 (11)	49 (78)	WFNS: 39 (62)	62 (98)	18* (29)	C	6/9
Wang, 2020 <sup>35</sup>	Retro	Cohort	54	59 (51-63)	31 (57)	HH: 11 (20)	28 (52)	19* (33)	C	4/9
Jayamanoharan, 2020 <sup>36</sup>	Retro	Cohort	30	59	16 (53)	WFNS: 11 (37)	-	13 (43)	C	5/8
Guresir, 2020 <sup>37</sup>	-	Cohort	231	54 (14)	-	WFNS: 0 (0)	F3: 207 (90)	22 (10)	R	5/8
Al-Mufti, 2019 <sup>38</sup>	Pro	Cohort	1067	-	731 (69)	HH3-5: 645 (61)	560 (52)	202* (19) 170* (16)	C R	6/8
Zhang, 2019 <sup>39</sup>	Pro	Cohort	535	54 (11)	325 (60)	HH: 65 (12)	-	161 (30)	C	6/9
Giede-Jeppe, 2019 <sup>40</sup>	Pro	Cohort	319	-	221 (60)	-	-	112 (35)	R	6/8
Bevers, 2019 <sup>41</sup>	Pro	Cohort	240	58	162 (68)	-	-	86* (45)	C	5/9
Zhu, 2019 <sup>42</sup>	Pro	Cohort	196	58 (10)	103 (53)	WFNS: 55 (28)	mF: 102 (52)	41 (21) 57 (29)	C R	6/8
Kanamaru, 2019 <sup>43</sup>	Pro	Cohort	109	65 (13)	77 (71)	WFNS: 39 (36)	mF: 96 (77)	16 (15)	C	7/9
Chen, 2019 <sup>44</sup>	Pro	Cohort	104	52 (36-61)	61 (69)	median WFNS: 3	median mF: 2	35 (34)	C	5/9
Tanioka, 2019 <sup>45</sup>	Pro	Cohort	95	66	66 (69)	WFNS: 30 (32)	88 (93)	13 (14) 24 (25)	C R	7/8
Rasmussen. 2019 <sup>46</sup>	Pro	Cohort	90	53 (22-77)	74 (82)	WFNS: 13 (15)	90 (100)	24 (27)	C	5/8
Ahn, 2019 <sup>47</sup>	Pro	Cohort	60	52 (13)	46 (77)	median WFNS: 2	median mF: 3	14 (23)	C	6/8
Hviid, 2019 <sup>48</sup>	Pro	Cohort	44	58 (10)	13 (30)	WFNS: 20 (45)	33 (75)	9 (20)	C	5/9
Kim, 2019 <sup>49</sup>	Pro	Cohort	42	58	30 (71)	HH: 17 (40)	33 (79)	14 (33)	C	3/8
Kwan, 2019 <sup>50</sup>	Pro	Cohort	25	56 (10)	15 (60)	mean WFNS: 3	mean mF: 4	9 (36)	C	4/9
Qi, 2019 <sup>51</sup>	Retro	Cohort	252	53 (13)	108 (43)	median HH: 1	-	53 (21)	C	6/8
Wu, 2019 <sup>52</sup>	Retro	Cohort	122	55 (11)	74 (61)	HH: 24 (20)	109 (89)	43 (35)	C	7/8
Lin, 2019 <sup>53</sup>	-	Cohort	146	-	-	-	-	34 (23) 31 (21)	C R	5/9
Aida, 2019 <sup>54</sup>	-	Cohort	27	58	20 (74)	HK: 2 (7)	23 (85)	7 (26)	C	3/8
Hostettler, 2018 <sup>55</sup>	Pro	Cohort	548	55 (13)	67 (67)	WFNS: 145 (26)	485 (89)	127 (23)	R	5/8
Ayling, 2018 <sup>56</sup>	Pro	Cohort	413	51 (11)	289 (70)	WFNS: 100 (24)	-	72* (19)	C	5/8

Griessenauer, 2018 <sup>57</sup>	Pro	Cohort	149	55 (13)	114 (77)	HH: 34 (23)	126 (85)	34 (23) 31* (21)	C R	7/9
Hendrix, 2018 <sup>58</sup>	Pro	Cohort	149	55 (13)	114 (77)	HH: 34 (23)	126 (85)	34 (23) 31* (21)	C R	7/9
Nakatsuka, 2018 <sup>59</sup>	Pro	Cohort	109	65 (12)	76 (70)	WFNS: 32 (29)	mF4: 37 (34)	17 (16) 14 (13)	C R	8/9
Yang, 2018 <sup>60</sup>	Pro	Cohort	108	53 (38-61)	63 (58)	median WFNS 3	median mF: 2	36 (33)	C	5/9
Cai, 2018 <sup>61</sup>	Pro	Cohort	60	56 (12)	38 (63)	HH: 13 (21)	mF: 31 (52)	17 (28)	R	5/8
Thomas, 2018 <sup>62</sup>	Pro	Cohort	21	-	9 (43)	-	-	4 (19)	R	6/9
Halawa, 2018 <sup>63</sup>	Pro	Cohort	19	61 (43-79)	14 (74)	HH: 4 (21)	17 (89)	7 (37)	C	3/8
Kim, 2018 <sup>64</sup>	Pro	Cohort	87	60	56 (64)	HH: 46 (53)	60 (69)	38 (44)	C	3/8
Matano, 2018 <sup>65</sup>	Retro	Cohort	333	60	210 (63)	HK: 104 (31)	-	41 (12)	R	5/8
Ray, 2018 <sup>66</sup>	Retro	Cohort	169	53 (13)	114 (68)	WFNS: 67 (40)	mF: 106 (63)	38 (25)	C	4/8
Suzuki, 2018 <sup>67</sup>	Retro	Cohort	156	64	116 (74)	WFNS: 67 (43)	122 (78)	21 (13)	C	8/9
Chaudhry, 2018 <sup>68</sup>	Retro	Cohort	53	58 (12)	35 (66)	-	F3: 43 (81)	20* (38)	R	5/9
Tao, 2017 <sup>69</sup>	Pro	Cohort	247	56 (12)	159 (64)	median WFNS 2	median mF: 3	47 (19)	C	5/8
Griessenauer, 2017 <sup>70</sup>	Pro	Cohort	149	55 (13)	144 (77)	HH: 34 (23)	126 (85)	34 (23) 31* (21)	C R	6/8
Hendrix, 2017 <sup>71</sup>	Pro	Cohort	149	55 (13)	144 (77)	HH: 34 (23)	126 (85)	34 (23) 31* (21)	C R	6/8
Hendrix, 2017 <sup>72</sup>	Pro	Cohort	149	55 (13)	144 (77)	HH: 34 (23)	126 (85)	34 (23) 31* (21)	C R	7/9
Hendrix, 2017 <sup>73</sup>	Pro	Cohort	149	55 (13)	144 (77)	HH: 34 (23)	126 (85)	34* (23) 31* (21)	C R	6/9
Hendrix, 2017 <sup>74</sup>	Pro	Cohort	149	55 (13)	144 (77)	HH: 34 (23)	126 (85)	34 (23) 31* (21)	C R	6/8
Frontera, 2017 <sup>75</sup>	Pro	Cohort	106	55	63 (59)	HH: 24 (23)	-	-	C	5/9
Chamling, 2017 <sup>76</sup>	Pro	Cohort	89	54	58 (65)	WFNS: 38 (43)	75 (84)	24 (27)	C	6/8
Beseoglu, 2017 <sup>77</sup>	Pro	Cohort	87	57 (11)	57 (66)	39 (45)	61 (69)	36 (41) 14 (16)	C R	4/8
Hollig, 2017 <sup>78</sup>	Pro	Cohort	81	54 (13)	51 (63)	WFNS: 37 (45)	67 (82)	18 (22)	C	5/8
Chaudhry, 2017 <sup>79</sup>	Pro	Cohort	80	58 (12)	49 (61)	HH: 27 (34)	77 (96)	28 (35) 17 (21)	C R	5/9
Chaudhry, 2017 <sup>80</sup>	Pro	Cohort	80	57 (12)	50 (63)	HH: 28 (35)	77 (96)	28 (35) 16 (20)	C R	5/9
Bache, 2017 <sup>81</sup>	Pro	Cohort	27	57 (13)	19 (70)	WFNS:29 (70)	26 (96)	8* (47)	C	5/9

Griessenauer, 2017 <sup>82</sup>	Pro	Cohort	27	55 (22-85)	14 (52)	HH: 11 (41)	25 (93)	9 (33)	R	6/8
Wisniewski, 2017 <sup>83</sup>	Pro	Cohort	20	60	11 (55)	HH: 2 (10)	13 (65)	9 (45)	C	6/9
Spitzer, 2017 <sup>84</sup>	Pro	Cohort	15	51 (9)	9 (60)	WFNS: 4 (27)	11 (73)	8 (53)	C	4/8
Xu, 2017 <sup>85</sup>	Retro	Cohort	386	56	243 (63)	HH: 115 (30)	139 (36)	23 (6)	C	6/8
Fukuda, 2017 <sup>86</sup>	Retro	Cohort	187	64	150 (80)	WFNS: 65 (35)	-	24 (13) 12 (6)	C R	6/8
Li, 2017 <sup>87</sup>	-	Cohort	83	60 (5)	38 (46)	HH: 18 (22)	-	31 (37)	C	3/8
Lu, 2017 <sup>88</sup>	-	Case-control	40	59	22 (55)	WFNS3-5: 13 (33)	40 (100)	20 (50)	R	3/9
Ewelina, 2017 <sup>89</sup>	-	Cohort	24	59	11 (46)	WFNS median 2	median: 3	6 (25)	C	5/9
Srinivasan, 2016 <sup>90</sup>	Pro	Cohort	246	50	129 (52)	WFNS: 44 (18)	211 (86)	94 (38) 62 (25)	C R	6/8
Ding, 2016 <sup>91</sup>	Pro	Cohort	125	52 (13)	76 (61)	WFNS median 3	median: 2	38 (30)	C	5/9
Jiang, 2016 <sup>92</sup>	Pro	Cohort	118	42 (12)	48 (41)	WFNS median 3	median: 3	33* (30)	C	5/9
Rasmussen, 2016 <sup>93</sup>	Pro	Cohort	90	53	74 (82)	WFNS: 13 (15)	90 (100)	24 (27)	C	5/8
Mijiti, 2016 <sup>94</sup>	Retro	Cohort	343	-	210 (61)	HH:117 (34)	mF: 149 (43)	99 (29) 87 (25)	C R	7/8
van Donkelaar, 2016 <sup>95</sup>	Retro	Cohort	285	55 (47-65)	189 (66)	WFNS: 141 (49)	-	84 (29)	R	7/8
Behrouz, 2016 <sup>96</sup>	Retro	Cohort	142	55 (13)	85 (60)	-	142 (100)	54 (38)	C	5/8
Wu, 2016 <sup>97</sup>	-	Cohort	57	58 (10)	24 (42)	HH: 8 (14)	-	27 (47)	C	5/9
Donnelly, 2015 <sup>98</sup>	Pro	Cohort	332	53	239 (72)	HH3-5:141 (42)	224 (67)	183* (55)	C	6/8
Donnelly, 2015 <sup>99</sup>	Pro	Cohort	332	53	239 (72)	HH3-5: 141 (42)	224 (67)	183* (55)	C	5/8
Chen, 2015 <sup>100</sup>	Pro	Cohort	120	41 (12)	49 (41)	WFNS mean 2.6	mean 2.8	36 (30) 15 (13)	C R	5/9
Shen, 2015 <sup>101</sup>	Pro	Cohort	118	42 (12)	48 (41)	WFNS mean 3	mean: 3	33 *(30) 14 (12)	C R	4/9
Crago, 2015 <sup>102</sup>	Pro	Cohort	99	50 (10)	62 (63)	HH: 24 (24)	63 (64)	45* (47)	C	6/8
Helbok, 2015 <sup>103</sup>	Pro	Cohort	26	55 (47-67)	15 (58)	HH: 18 (69)	mF: 20 (77)	6 (23)	R	4/8
Kofler, 2015 <sup>104</sup>	Pro	Cohort	25	55 (47-68)	14 (56)	HH: 18 (72)	mF: 19 (76)	5 (20)	R	4/8
Leclerc, 2015 <sup>105</sup>	Both	Cohort	74	55 (15)	54 (73)	WFNS mean 2.5	43 (58)	22 (30)	C	5/8
Sun, 2015 <sup>106</sup>	Retro	Cohort	218	-	139 (64)	HH3-5: 31 (14)	91 (42)	78 (36)	C	8/8
Tang, 2015 <sup>107</sup>	-	Cohort	58	53 (11)	34 (59)	HH: 8 (14)	mF: 22 (38)	12 (21)	C	4/9
Suzuki, 2015 <sup>108</sup>	-	Cohort	30	63 (2)	23 (77)	WFNS: 13 (43)	30 (100)	14* (47)	C	6/9
Helbok, 2015 <sup>109</sup>	-	Cohort	22	56 (47-68)	15 (68)	HH: 17 (77)	mF: 16 (73)	4 (18)	R	4/8
Cai, 2014 <sup>110</sup>	Pro	Cohort	120	41 (12)	49 (41)	WFNS mean 2.6	mean 2.8	36 (30)	C	5/8

Martini, 2014 <sup>111</sup>	Pro	Cohort	95	55 (-)	66 (69)	HH: 36 (38)	87 (92)	28 (29)	R	5/8
Bergstrom, 2014 <sup>112</sup>	Pro	Cohort	48	-	39 (81)	-	43 (90)	15* (32)	C	4/9
Zanier, 2014 <sup>113</sup>	Pro	Cohort	39	56 (13)	25 (64)	WFNS: 21 (54)	38 (97)	14 (34) 6 (15)	C R	4/9
Radolf, 2014 <sup>114</sup>	Pro	Cohort	18	52	10 (55)	WFNS: 8 (44)	median 3	8 (45)	C	5/8
Gomes, 2014 <sup>115</sup>	Pro	Cohort	12	58 (6)	10 (83)	WFNS median 3.5	mF median: 4	7 (58)	C	3/8
Beseoglu, 2014 <sup>116</sup>	Retro	Cohort	274	54	187 (68)	WFNS: 132 (48)	-	- -	C R	6/8
Uekusa, 2014 <sup>117</sup>	-	Cohort	20	-	15 (75)	WFNS: 16 (80)	-	7* (41)	R	2/8
De Rooij, 2013 <sup>118</sup>	Pro	Cohort	371	44 (22-67)	261 (70)	WFNS: 104 (28)	mF: 324 (87)	110 (30)	R	6/8
Bian, 2013 <sup>119</sup>	Pro	Cohort	239	53 (12)	123 (51)	WFNS: 25 (11)	127 (53)	34* (15)	C	7/8
Gallek, 2013 <sup>120</sup>	Pro	Cohort	235	53	165 (70)	HH: 44 (19)	161 (69)	113* (48)	C	3/8
Nakagawa, 2013 <sup>121</sup>	Pro	Cohort	103	66	68 (66)	HK: 41 (40)	57 (55)	13 (13)	C	4/8
Ohnishi, 2013 <sup>122</sup>	Pro	Cohort	95	62	53 (56)	HK: 39 (41)	86 (91)	22 (23) 18 (19)	C R	6/8
July, 2013 <sup>123</sup>	Pro	Cohort	44	52 (11)	24 (55)	WFNS: 6 (14)	F4: 11 (25)	29 (66)	C	4/8
Barges-Coll, 2013 <sup>124</sup>	Pro	Cohort	40	47 (2)	22 (55)	WFNS: 0 (0)	33 (83)	17* (43)	C	6/9
Lanterna, 2013 <sup>125</sup>	Pro	Cohort	26	53 (13)	19 (73)	HH: 0 (0)	9 (35)	12* (46)	C	3/8
Fischer, 2013 <sup>126</sup>	Pro	Cohort	20	52	16 (80)	WFNS: 7 (35)	18 (90)	7* (35)	R	5/9
Kim, 2013 <sup>127</sup>	-	Cohort	245	54	172 (70)	-	-	109* (45)	C	2/8
Aggarwal, 2013 <sup>128</sup>	-	Cohort	74	50	-	WFNS: 16 (22)	61 (82)	39 (53)	C	3/8
Maimaitili, 2013 <sup>129</sup>	-	Cohort	49	53 (13)	27 (57)	HH: 12 (24)	27 (55)	19 (39)	C	3/9
Zanier, 2013 <sup>130</sup>	-	Cohort	38	54	31 (82)	WFNS: 24 (63)	37 (97)	11* (31) 10 (26)	C R	4/9
Nyquist, 2013 <sup>131</sup>	-	Case-control	28	-	-	-	-	14 (50)	C	4/9
Juvela, 2012 <sup>132</sup>	Pro	Cohort	178	50 (13)	90 (51)	WFNS: 37 (21)	129 (72)	48 (26) 50 (28)	C R	6/8
Papanikolaou, 2012 <sup>133</sup>	Pro	Cohort	37	43 (2)	23 (62)	HH: 23 (62)	23 (62)	12* (33)	R	6/8
Yang, 2012 <sup>134</sup>	Pro	Cohort	20	52	10 (50)	HH mean: 3	Mean mF: 3	10 (50)	R	8/9
Zhu, 2012 <sup>135</sup>	Retro	Cohort	303	44 (12)	172 (57)	WFNS mean 2.3	mean mF: 2.7	131 (43)	C	6/9
Jeon, 2012 <sup>136</sup>	Retro	Cohort	93	57	60 (65)	HH: 31 (33)	F3: 37 (40)	26 (29)	C	5/8
Vrsajkov, 2012 <sup>137</sup>	Retro	Cohort	82	53	60 (73)	WFNS mean 2.4	-	39 (48)	C	5/8
Watanabe, 2012 <sup>138</sup>	Retro	Cohort	34	64	21 (62)	27 979)	34 (100)	6 (18)	C	4/8

Jung, 2012 <sup>139</sup>	Retro	Cohort	24	51 (11)	19 (79)	WFNS: 7 (29)	24 (100)	4 (17)	C	4/9
Sarrafzadeh, 2012 <sup>140</sup>	Retro	Cohort	21	55 (46-62)	11 (52)	WFNS: 12 (57)	median: 4	12 (57)	C	5/8
Beeftink, 2011 <sup>141</sup>	Pro	Cohort	67	57	47 (70)	WFNS: 13 (19)	-	10 (15)	C	8/8
Zanier, 2011 <sup>142</sup>	Pro	Cohort	38	54 (2)	31 (82)	WFNS: 24 (63)	37 (97)	14* (47)	C	3/9
Dietmann, 2011 <sup>143</sup>	Pro	Cohort	20	52	16 (80)	WFNS: 7 (35)	-	7 (35)	R	4/9
Fischer, 2011 <sup>144</sup>	Pro	Cohort	20	52	16 (80)	WFNS: 7 (35)	18 (90)	7 (35)	R	6/9
Zheng, 2011 <sup>145</sup>	Retro	Cohort	383	56 (-)	66	WFNS: 124 (32)	-	54 (24)	R	5/8
Zhu, 2011 <sup>146</sup>	Retro	Cohort	303	44 (12)	172 (57)	WFNS mean 2.3	mean mF: 2.7	131 (43)	C	7/9
Nakae, 2011 <sup>147</sup>	Retro	Cohort	142	61 (12)	85 (60)	HH3-5: 113 (80)	71 (50)	28 (20)	R	3/8
Taub, 2011 <sup>148</sup>	Retro	Cohort	119	57 (-)	90 (76)	HH: 19 (16)	49 (41)	46 (39)	R	6/8
Zanier, 2011 <sup>149</sup>	-	Cohort	35	-	-	-	35 (100)	10 (29)	R	4/9
Hirashima, 2011 <sup>150</sup>	-	Cohort	28	62	19 (68)	WFNS3-5: 15 (54)	F3: 19 (68)	14 (50)	C	3/8
Wang, 2011 <sup>151</sup>	-	Cohort	21	46	11 (52)	-	-	7 (33)	C	7/9
Naidech, 2010 <sup>152</sup>	Pro	RCT	44	57	33 (75)	WFNS: 13 (30)	43 (98)	10 (23) 20 (45)	C R	3/7
Lackner, 2010 <sup>153</sup>	Pro	Cohort	20	52	16 (80)	WFNS: 7 (35)	18 (90)	7 (35)	R	6/9
Kasius, 2010 <sup>154</sup>	Retro	Cohort	106	54	71 (67)	WFNS: 25 (24)	-	14* (15)	C	6/8
Ruigrok, 2010 <sup>155</sup>	-	Cohort	208	60 (15)	149 (72)	WFNS: 41 (20)	-	49 (24)	C	6/9
Vergouwen, 2010 <sup>156</sup>	-	Cohort	31	53 (11)	19 (61)	WFNS median 2	-	11 (35)	C	5/8
Juvela, 2009 <sup>157</sup>	Pro	Cohort	105	48 (11)	52 (50)	WFNS: 20 (19)	73 (70)	24 (23) 29* (28)	C R	7/8
Vergouwen, 2009 <sup>158</sup>	Pro	Cohort	31	53 (11)	19 (61)	WFNS median 2	-	11 (35)	C	5/8
Oddo, 2009 <sup>159</sup>	Pro	Cohort	20	51	13 (65)	WFNS: 14 (70)	19 (95)	8 (40)	C	5/8
Kruyt, 2008 <sup>160</sup>	Pro	Cohort	265	56	175 (66)	49 (18)	-	86* (33)	C	5/8
van den Bergh, 2008 <sup>161</sup>	Pro	Cohort	167	56 (15)	110 (66)	WFNS: 47 (28)	-	49 (29)	C	7/8
Starke, 2008 <sup>162</sup>	Pro	Cohort	77	54	49 (64)	HH: 13 (17)	42 (55)	24 (30) 8 (10)	C R	7/8
Lewis, 2008 <sup>163</sup>	-	Cohort	30	56	22 (73)	WFNS: 15 (50)	30 (100)	17 (57)	C	4/9
Isman, 2008 <sup>164</sup>	Pro	Cohort	20	46 (13)	9 (45)	HH: 1 (5)	9 (45)	14 (70)	C	4/9
Dorhout Mees, 2007 <sup>165</sup>	Pro	Cohort	155	56	102 (66)	WFNS: 33 (21)	-	27 (17)	C	6/8
Tseng, 2007 <sup>166</sup>	Pro	Cohort	80	53 (12)	44 (55)	WFNS: 26 (19)	69 (86)	14 (18)	C	5/8
Igarashi, 2007 <sup>167</sup>	Pro	Cohort	67	55	44 (66)	HK: 7 (10)	58 (87)	12 (18)	C	4/8
Oh, 2007 <sup>168</sup>	Retro	Cohort	158	51	98 (62)	HH: 0 (0)	F3: 73 (46)	46 (31)	C	5/8
Schebesch, 2007 <sup>169</sup>	Retro	Cohort	88	53	54 (61)	HH: 22 (25)	-	37 (42)	C	3/8

Schoch, 2007 <sup>170</sup>	Retro	Cohort	64	55 (12)	49 (77)	WFNS: 39 (61)	60 (94)	17 (27)	R	3/8
Siironen, 2007 <sup>171</sup>	-	Cohort	105	48	52 (50)	WFNS: 20 (19)	F3: 73 (70)	24 (23)	C	6/8
Lewis, 2007 <sup>172</sup>	-	Cohort	20	54	15 (75)	WFNS: 13 (65)	20 (100)	13 (65)	C	4/9
Juvela, 2006 <sup>173</sup>	Pro	Cohort	136	50	67 (49)	WFNS: 27 (20)	99 (73)	37 (27) 37* (30)	C R	5/8
Rothoerl, 2006 <sup>174</sup>	Pro	Cohort	15	47	10 (66)	HH: 2 (13)	-	5 (33)	C	3/8
Rothoerl, 2006 <sup>175</sup>	Retro	Cohort	88	53	54 (61)	HH: 22 (25)	-	37 (42)	C	3/8
Frijns, 2006 <sup>176</sup>	-	Cohort	106	54	72 (68)	WFNS: 25 (24)	-	12* (13)	C	6/8
Frijns, 2006 <sup>177</sup>	-	Cohort	86	55	62 (72)	WFNS: 19 (22)	-	9 (14)	C	5/8
Oertel, 2006 <sup>178</sup>	-	Cohort	51	52 (11)	32 (63)	WFNS: 28 (45)	34 (67)	26 (51)	C	5/8
Lanterna, 2005 <sup>179</sup>	Pro	Cohort	101	52 (8)	53 (53)	HH: 0 (0)	50 (50)	37 (37)	C	5/8
Kastner, 2005 <sup>180</sup>	Pro	Cohort	20	58 (10)	11 (55)	WFNS: 5 (25)	13 (65)	7 (35) 7 (35)	C R	4/9
Asaeda, 2005 <sup>181</sup>	-	Cohort	34	63	22 (65)	-	30 (88)	15 (44)	R	3/8
Dohi, 2005 <sup>182</sup>	-	Cohort	15	62 (15)	4 (27)	HH: 6 (40)	-	7 (47)	C	4/8
Tanriverdi, 2005 <sup>183</sup>	-	Cohort	12	47	8 (67)	HH: 3 (25)	7 (58)	4 (33)	C	5/9
Juvela, 2005 <sup>184</sup>	Pro	Cohort	175	51	88 (50)	WFNS: 36 (21)	127 (73)	46* (26) 46* (30)	C R	6/8
Khurana, 2004 <sup>185</sup>	Pro	Case-control	51	54 (12)	35 (69)	WFNS: 5 (10)	41 (80)	12 (24)	C	7/9
McGirt, 2004 <sup>186</sup>	Pro	Cohort	40	53 (12)	32 (80)	HH median: 2	26 (65)	16 (40)	C	8/8
Sarrafzadeh, 2004 <sup>187</sup>	Pro	Cohort	13	46 (14)	9 (69)	WFNS: 4 (31)	10 (77)	10 (77)	C	3/9
Collignon, 2004 <sup>188</sup>	-	Cohort	128	54	83 (65)	HH: 40 (31)	-	55* (43)	C	3/8
Hendryk, 2004 <sup>189</sup>	-	Cohort	30	-	-	-	-	14 (47)	C	3/8
van den Bergh, 2003 <sup>190</sup>	Pro	Cohort	107	-	66 (62)	WFNS: 45 (42)	-	21 (20)	C	8/8
Qureshi, 2002 <sup>191</sup>	Pro	Cohort	298	52	218 (73)	-	-	-	C	6/8
Juvela, 2002 <sup>192</sup>	Pro	Cohort	51	51	23 (45)	WFNS: 12 (24)	35 (69)	13* (26) 9 (18)	C R	6/8
Nissen, 2001 <sup>193</sup>	Pro	Cohort	36	-	23 (64)	WFNS: 0 (0)	-	13 (36)	C	2/8
Mascia, 2001 <sup>194</sup>	Pro	Cohort	20	55 (16)	13 (65)	WFNS median 3	19 (95)	4 (20)	C	4/8
Hirashima, 2001 <sup>195</sup>	-	Cohort	28	61	17 (61)	-	-	10 (36)	R	4/9
Nam, 2001 <sup>196</sup>	-	Cohort	24	55	15 (63)	HH: 6 (25)	-	11 (46)	C	3/8
Juvela, 2000 <sup>197</sup>	Pro	Cohort	70	46	34 (49)	HH: 12 (17)	44 (63)	20* (31)	R	2/9
Gruber, 2000 <sup>198</sup>	Pro	Cohort	44	51 (14)	26 (60)	HH: 23 (52)	35 (80)	17 (39)	R	4/8
Takenaka, 2000 <sup>199</sup>	-	Cohort	20	56	13 (65)	HH: 2 (10)	-	7 (35)	C	3/9

Mayer, 1999 <sup>200</sup>	Retro	Cohort	72	51 (13)	47 (65)	HH: 14 (20)	36 (50)	27 (36)	C	6/8
Fuji, 1997 <sup>201</sup>	-	Cohort	117	58	67 (57)	HH: 37 (32)	-	18 (15)	C	4/8
Hirashima, 1997 <sup>202</sup>	-	Cohort	21	58	8 (38)	HH: 8 (38)	14 (67)	9 (43)	C	3/9
Hirashima, 1997 <sup>203</sup>	-	Cohort	19	61	11 (58)	HH: 5 (26)	15 (83)	6* (33)	R	3/9
Okuchi, 1996 <sup>204</sup>	Pro	Cohort	17	60	-	HH: 0	-	4 (24)	C	3/8
Hirashima, 1995 <sup>205</sup>	Pro	Cohort	32	59	18 (56)	WFNS: 4 (13)	23 (72)	18 (56) 7 (22)	C R	3/8
Hirashima, 1994 <sup>206</sup>	-	Cohort	32	58	16 (50)	WFNS: 11 (34)	19 (59)	9 (28)	R	3/8
Hirashima, 1994 <sup>207</sup>	-	Cohort	21	59	13 (62)	HH: 5 (24)	14 (67)	6 (29)	R	2/8
Suzuki, 1989 <sup>208</sup>	-	Cohort	14	64	11 (79)	-	-	11 (79)	C	3/9
Rodriguez, 1988 <sup>209</sup>	-	Cohort	40	-	-	HH: 0 (0)	-	16 (40)	C	4/9

<sup>1</sup> Retro: retrospective study, pro: prospective study.

<sup>2</sup> Age is given in mean with SD or median with quartile 1 – quartile 3, if available.

<sup>3</sup> Poor outcome is defined as WFNS 4-5, HH 4-5 or HK 4-5.

<sup>4</sup> F3: fisher 3 score, F4: fisher 4 score, mF: modified Fisher scale.

<sup>5</sup> C: clinical DCI, R: radiological DCI

\* data on DCI development was not available of the entire SAH cohort



Table S2 - Biomarker classification table of all 724 biomarkers studied for clinical or radiological DCI.

Biomarker	Abbreviation	Biomarker (sub-)group	Medium	Clinical DCI		Radiological DCI	
				Studies	Conclusion	Studies	Conclusion
<b>Brain injury</b>							
A-II Spectrin Breakdown Product 120	SBPD120	Brain injury	CSF	172	Significant		
A-II Spectrin Breakdown Product 145	SBPD145	Brain injury	CSF	172	Significant		
A-II Spectrin Breakdown Product 150	SBPD150	Brain injury	CSF	172	Significant		
CD45-positive mitochondrial particles		Other	CSF	24	Non-significant		
Glutamate-aspartate transporter-allophycocyanin positive mitochondrial particles	GLAST-positive	Other	CSF	24	Significant		
Glycerol		Brain injury	MD	187	Non-significant		
extracellular JC-1 red/green ratio	JC-1	Brain injury	CSF	24	Significant		
Lactate dehydrogenase	LDH	Brain injury	Blood	90	Non-significant	90	Non-significant
Myelin basic protein	MBP	Brain injury	CSF			195	Significant
phosphorylated axonal Neurofilament subunit H	pNF-H	Brain injury	Blood	163	Non-significant		
phosphorylated axonal Neurofilament subunit H	pNF-H	Brain injury	CSF	163	Significant		
Neurofilament light chain	NFL	Brain injury	Blood	48	Non-significant		
Neurofilament light chain	NFL	Brain injury	CSF	63	Non-significant	149	Non-significant
Neuron specific enolase	NSE	Brain injury	Blood	178	Non-significant		
S-100B		Brain injury	Blood	112, 178	Significant		
Tau protein	Tau	Brain injury	CSF	63, 130	Significant	130	Non-significant
Tau protein	Tau	Brain injury	MD			109	Non-significant
Taurine		Brain injury	Blood	124	Non-significant		
Taurine		Brain injury	MD			104	Significant
<b>Coagulation cascade</b>							
α <sub>2</sub> -antiplasmin	A2-PI	Coagulation cascade	Blood	201	Non-significant		
Activated partial thromboplastin time	APTT	Coagulation cascade	Blood	201	Non-significant	134	Non-significant
ADAM Metalloproteinase with thrombospondin type 1 motif 13	ADAMTS13	Coagulation cascade	Blood	87,107,158	Significant		
Antiphospholipid antibodies	aPLs	Coagulation cascade	Blood	205	Non-significant	205	Non-significant
Antithrombin III	AT III	Coagulation cascade	Blood	201	Non-significant		
D-dimer		Coagulation cascade	Blood	28, 35, 86, 146, 166, 173, 201	Significant	31, 86, 173	Significant
D-dimer		Coagulation cascade	CSF	36	Significant		
D-dimer/plasminogen ratio	D-dimer/PLG	Coagulation cascade	CSF	36	Significant		
Enhancement of platelet sensitivity	EPS	Coagulation cascade	Blood	201	Non-significant		

Fibrinogen		Coagulation cascade	Blood	8, 35, 166, 201	Significant	33	Non-significant
International normalized ratio	INR	Coagulation cascade	Blood			134	Non-significant
Mean platelet volume	MPV	Coagulation cascade	Blood	8, 23, 30	Significant		
Mean platelet volume/platelet count ratio	MPV-PLT ratio	Coagulation cascade	Blood	23, 30, 66	Significant		
Plasmin- $\alpha_2$ -antiplasmin complex	PAP	Coagulation cascade	Blood	201	Non-significant		
Plasminogen	PLG	Coagulation cascade	Blood	201	Non-significant		
Plasminogen	PLG	Coagulation cascade	CSF	36	Non-significant		
Platelet activating factor	PAF	Coagulation cascade	Blood	202	Significant	206	Significant
Platelet activating factor	PAF	Coagulation cascade	CSF			207	Significant
Platelet activating factor - acetylhydrolase	PAF-AH	Coagulation cascade	Blood			206	Non-significant
Platelet activating factor - acetylhydrolase	PAF-AH	Coagulation cascade	CSF	150	Non-significant	207	Significant
Platelet aggregation		Coagulation cascade	Blood			206	Non-significant
Platelet count	PC	Coagulation cascade	Blood	23, 28, 30, 35, 69, 128, 136, 151, 154, 169, 193, 201	Inconclusive	134, 206	Significant
Platelet factor 4	PF4	Coagulation cascade	Blood	202	Non-significant	206	Significant
Platelet membrane glycoprotein-140	GMP-140	Coagulation cascade	Blood	87	Significant		
CD41+/A+ platelet microparticle	PMP	Coagulation cascade	Blood			153	Significant
CD41/CD61-positive mitochondrial particles		Other	CSF	24	Non-significant		
Platelet/large cell ratio	P-LCR	Coagulation cascade	Blood	23	Non-significant		
Protein C		Coagulation cascade	Blood	201	Non-significant		
Prothrombin fragment F1+2	PF 1+2	Coagulation cascade	Blood	201	Non-significant		
Prothrombin time	PT	Coagulation cascade	Blood	28, 35, 201	Non-significant	134	Non-significant
Signal peptide-Cub-Egf domain-containing protein-1	SCUBE1	Coagulation cascade	Blood	91	Significant		
Thrombin-antithrombin III complex	TAT	Coagulation cascade	Blood	201, 202	Significant		
Thrombin-antithrombin III complex	TAT	Coagulation cascade	CSF			203	Significant
Thromboelastography maximal amplitude	TEG-MA	Coagulation cascade	Blood	75	Significant		
Tissue factor	TF	Coagulation cascade	Blood	202	Non-significant		
Membrane-bound tissue factor	mTF	Coagulation cascade	CSF			195	Significant
von Willebrand factor	vWF	Coagulation cascade, Endothelial injury	Blood	87, 107, 176, 202	Significant		
von Willebrand factor antigen	vWF antigen	Coagulation cascade, Endothelial injury	Blood	158	Significant		
von Willebrand factor propeptide	vWF propeptide	Coagulation cascade, Endothelial injury	Blood	158, 176	Significant		
von Willebrand factor ristocetin cofactor activity	vWFRiCof	Coagulation cascade, Endothelial injury	Blood	158	Non-significant		

<b>Cortical spreading depressions</b>							
Glutamate		Cortical spreading depressions	CSF	19	Significant		
Glutamate		Cortical spreading depressions	MD	187	Non-significant		
<b>Endothelial injury</b>							
Angiopoetin 1	Ang-1	Endothelial injury	Blood			144	Significant
Angiopoetin 2	Ang-2	Endothelial injury	Blood			144	Non-significant
CD105+/A+ endothelial microparticle	EMP	Endothelial injury, Vascular tone	Blood			153	Significant
<i>Extra domain 1-Fibronectin</i>	ED1-FN	Endothelial injury	Blood	176	Non-significant		
Homocysteine	Hcy	Endothelial injury	Blood	28	Significant		
Sphingosine 1-phosphate	S1P	Endothelial injury	CSF			25	Non-significant
von Willebrand Factor positive mitochondrial particles	vWF-positive	Endothelial injury	CSF	24	Significant		
<b>Genetic</b>							
Apolipoprotein E ε2 allele	ApoE ε2	Genetic - Genomic	Buccal swabs	157	Non-significant	157	Non-significant
Apolipoprotein E ε3 allele	ApoE ε3	Genetic - Genomic	Buccal swabs	157	Non-significant	157	Non-significant
Apolipoprotein E ε4 allele	ApoE ε4	Genetic - Genomic	Buccal swabs	157	Non-significant	157	Non-significant
Apolipoprotein E ε4 allele	ApoE ε4	Genetic - Genomic	Blood	179	Significant		
Platelet activating factor - acetylhydrolase genotype	PAF-AH	Genetic - Genomic	Blood	150	Non-significant		
microRNA	miRNA	Genetic - microRNA	Blood			88	
1. miR-15a							1. Non-significant
2. miR-27a							2. Non-significant
3. miR-27b							3. Non-significant
4. miR-93-5p							4. Significant
5. miR-125a							5. Non-significant
6. miR-132							6. Significant
7. miR-297							7. Significant
8. miR-324							8. Non-significant
9. miR-339-5p							9. Non-significant
10. miR-421							10. Significant
11. miR-574							11. Significant
12. miR-1268							12. Significant
13. miR-1290							13. Significant
14. miR-3178							14. Significant

15. miR-3651						15. Significant
16. miR-4433						16. Non-significant
17. miR-4440						17. Non-significant
18. miR-4449						18. Significant
19. miR-4454						19. Non-significant
20. miR-4459						20. Significant
21. miR-4463						21. Significant
22. miR-4492						22. Significant
23. miR-4497						23. Significant
24. miR-4532						24. Significant
25. miR-4674						25. Significant
26. miR-4689						26. Significant
27. miR-4690						27. Significant
28. miR-4793						28. Significant
microRNA	miRNA	Genetic – MicroRNA	CSF	18, 81		
1. miR-9-3p					1. Non-significant	
2. miR-9-5p					2. Non-significant	
3. miR-10b-3p					3. Non-significant	
4. miR-15a-5p					4. Non-significant	
5. miR-15b-5p					5. Non-significant	
6. miR-16-5p					6. Non-significant	
7. miR-19a-3p					7. Non-significant	
8. miR-19b-3p					8. Non-significant	
9. miR-20a-5p					9. Non-significant	
10. miR-21-5p					10. Non-significant	
11. miR-22-3p					11. Non-significant	
12. miR-23a-3p					12. Non-significant	
13. miR-24-3p					13. Non-significant	
14. miR-25-3p					14. Non-significant	
15. miR-26a-5p					15. Non-significant	
16. miR-27a-3p					16. Non-significant	
17. miR-27b-3p					17. Non-significant	
18. miR-29a-3p					18. Non-significant	
19. miR-29b-3p					19. Non-significant	
20. miR-29c-3p					20. Non-significant	
21. miR-30a-5p					21. Non-significant	
22. miR-30d-5p					22. Non-significant	
23. miR-32-5p					23. Non-significant	
24. miR-34a-5p					24. Non-significant	
25. miR-34b-3p					25. Non-significant	
26. miR-92a-3p					26. Non-significant	

27. miR-93-5p 28. miR-125b-5p 29. miR-130a-3p 30. miR-130b-3p 31. miR-132-3p 32. miR-143-3p 33. miR-146a-5p 34. miR-152-3p 35. miR-185-5p 36. miR-193a-5p 37. miR-204-5p 38. miR-208a-3p 39. miR-221-3p 40. miR-222-3p 41. miR-223-3p 42. miR-328-3p 43. miR-361-5p 44. miR-451a 45. miR-483-5p 46. miR-486-5p 47. miR-490-3p 48. miR-520h 49. miR-553 50. miR-643 51. miR-let-7b-5p					27. Non-significant 28. Non-significant 29. Non-significant 30. Non-significant 31. Non-significant 32. Non-significant 33. Non-significant 34. Non-significant 35. Non-significant 36. Non-significant 37. Non-significant 38. Non-significant 39. Non-significant 40. Non-significant 41. Non-significant 42. Non-significant 43. Non-significant 44. Non-significant 45. Non-significant 46. Non-significant 47. Non-significant 48. Non-significant 49. Non-significant 50. Non-significant 51. Non-significant		
Cadherin related family member 5 – DNA methylation CpG-site cg11464053	CDHR5	Genetic - Methylation	Blood	<sup>17</sup>	Significant		
<i>DNA methylation intensity of distal intergenic region upstream of Inositol 1-,4-,5-trisphosphate receptor</i>	ITPR3	Genetic - Methylation	Blood	<sup>49</sup>	Significant		
DNA methylation CpG-sites 1. Cg0268232 2. Cg00074576 3. Cg00789789 4. Cg01141087 5. Cg01620248 6. Cg01834661 7. Cg02180699 8. Cg02750582 9. Cg03297297 10. Cg06479032	CpG	Genetic - Methylation	Blood	<sup>17</sup>	1. Significant 2. Significant 3. Significant 4. Significant 5. Significant 6. Significant 7. Significant 8. Significant 9. Significant 10. Significant		

<ol style="list-style-type: none"> <li>11. Cg06502011</li> <li>12. Cg07259358</li> <li>13. Cg08789159</li> <li>14. Cg09217327</li> <li>15. Cg11337626</li> <li>16. Cg12601633</li> <li>17. Cg14592104</li> <li>18. Cg14916658</li> <li>19. Cg15090337</li> <li>20. Cg16837008</li> <li>21. Cg18101414</li> <li>22. Cg19132447</li> <li>23. Cg19751670</li> <li>24. Cg21222185</li> <li>25. Cg21486195</li> <li>26. Cg21808832</li> <li>27. Cg23222472</li> <li>28. Cg23407997</li> <li>29. Cg25058463</li> <li>30. Cg25934800</li> <li>31. Cg26306080</li> <li>32. Cg26826512</li> <li>33. Cg089669578</li> </ol>					<ol style="list-style-type: none"> <li>11. Significant</li> <li>12. Significant</li> <li>13. Significant</li> <li>14. Significant</li> <li>15. Significant</li> <li>16. Significant</li> <li>17. Significant</li> <li>18. Significant</li> <li>19. Significant</li> <li>20. Significant</li> <li>21. Significant</li> <li>22. Significant</li> <li>23. Significant</li> <li>24. Significant</li> <li>25. Significant</li> <li>26. Significant</li> <li>27. Significant</li> <li>28. Significant</li> <li>29. Significant</li> <li>30. Significant</li> <li>31. Significant</li> <li>32. Significant</li> <li>33. Significant</li> </ol>		
<p>Hepcidin antimicrobial peptide gene DNA methylation CpG-site</p> <ol style="list-style-type: none"> <li>1. cg02131995</li> <li>2. cg04085447</li> <li>3. cg04668516</li> <li>4. cg17907567</li> <li>5. cg18149657</li> <li>6. cg23677000</li> <li>7. cg26283059</li> <li>8. cg27273033</li> </ol>	HAMP	Genetic - Methylation	CSF	<sup>11</sup>	<ol style="list-style-type: none"> <li>1. Non-significant</li> <li>2. Non-significant</li> <li>3. Non-significant</li> <li>4. Non-significant</li> <li>5. Non-significant</li> <li>6. Non-significant</li> <li>7. Non-significant</li> <li>8. Non-significant</li> </ol>		
<p>Insulin Receptor DNA methylation CpG-site cg00441765</p>	INSR	Genetic - Methylation	Blood	<sup>17</sup>	Significant		
<p><i>Soluble</i> Epoxide Hydrolase 2 polymorphism</p> <ol style="list-style-type: none"> <li>1. K55R</li> <li>2. R287Q</li> </ol>	EPHX2	Genetic - Polymorphism	Blood			<sup>111</sup>	<ol style="list-style-type: none"> <li>1. Significant</li> <li>2. Non-significant</li> </ol>
<p>Factor XIII subunit A polymorphism</p> <ol style="list-style-type: none"> <li>1. Pro564Leu</li> <li>2. Tyr204Phe</li> </ol>	FXIII A	Genetic - Polymorphism	Blood	<sup>155</sup>	<ol style="list-style-type: none"> <li>1. Non-significant</li> <li>2. Non-significant</li> </ol>		

3. Val34Leu					3. Non-significant	
Factor XIII subunit B polymorphism His95Arg	FXIII B	Genetic - Polymorphism	Blood	155	Non-significant	
Haptoglobin polymorphism	Hp	Genetic - Polymorphism	Blood	64, 105, 122		122
1. 1-1					1. Non-significant	1. Non-significant
2. 2-1					2. Significant	2. Non-significant
3. 2-2					3. Significant	3. Non-significant
Haptoglobin 2-1 subunit	Hp	Genetic – Polymorphism	Blood	64		
1. a1					1. Significant	
2. a2					2. Non-significant	
Single nucleotide polymorphism	SNP	Genetic - Polymorphism	Blood	127		
1. rs999662					1. Non-significant	
2. rs3849943					2. Non-significant	
3. rs6805782					3. Non-significant	
4. rs10967965					4. Non-significant	
Activating Signal Cointegrator 1 Complex subunit 2 mRNA	ASCC2	Genetic - RNA	Blood	17	Non-significant	
Cadherin related family member 5 - mRNA	CDHR5	Genetic - RNA	Blood	17	Significant	
DNA methyltransferase mRNA	DNMT1	Genetic - RNA	Blood	49	Significant	
Interleukin-1B RNA	IL-1B RNA	Genetic - RNA	Blood	196	Significant	
Immunoglobulin like domain containing receptor 1 mRNA	ILDR1	Genetic - RNA	Blood	17	Non-significant	
inositol 1-,4-,5trisphosphate receptor mRNA	ITPR3	Genetic - RNA	Blood	49	Significant	
Insulin receptor mRNA	INSR	Genetic - RNA	Blood	17	Significant	
Leucine rich single-pass membrane protein 1 mRNA	LSMEM1	Genetic - RNA	Blood	17	Non-significant	
Mitogen-activated protein kinase kinase kinase 13 mRNA	MAP3K13	Genetic - RNA	Blood	17	Non-significant	
Nucleolar protein 4 like mRNA	NOL4L	Genetic - RNA	Blood	17	Non-significant	
Ten-eleven translocation enzyme 1 mRNA	TET1	Genetic - RNA	Blood	49	Significant	
Ten-eleven translocation enzyme 2 mRNA	TET2	Genetic - RNA	Blood	49	Non-significant	
Ten-eleven translocation enzyme 3 mRNA	TET3	Genetic - RNA	Blood	49	Non-significant	
Tumor necrosis factor a RNA	TNF-α	Genetic - RNA	Blood	196	Non-significant	
Zinc finger protein 706 mRNA	ZNF706	Genetic - RNA	Blood	17	Non-significant	
20-Hydroxyeicosatetraenoic acid SNP CYP4F2 SNP rs3093089 g.4593T>C	20-HETE SNP CYP4F2	Genetic – SNP	CSF & Blood	99	Significant	
Aconitase 1 SNPs	ACO1	Genetic - SNP	Blood	12		
1. rs1028932					1. Non-significant	
2. rs2026739					2. Non-significant	
3. rs4879583					3. Non-significant	
4. rs7866419					4. Non-significant	

5. rs10435797 6. rs10738885 7. rs10813816 8. rs10970972 9. rs13292540 10. rs13302577					5. Non-significant 6. Non-significant 7. Non-significant 8. Non-significant 9. Non-significant 10. Non-significant	
Aconitase 2 SNPs 1. rs2076196 2. rs9611598	ACO2	Genetic - SNP	Blood	<sup>12</sup>	1. Non-significant 2. Non-significant	
Amyloid precursor protein SNPs 1. rs3991 2. rs128648 3. rs216763 4. rs216772 5. rs216780 6. rs373521 7. rs429410 8. rs436587 9. rs454017 10. rs465853 11. rs466609 12. rs1701000 13. rs1783016 14. rs1787438 15. rs2026225 16. rs2070652 17. rs2234988 18. rs2242683 19. rs2829970 20. rs2829979 21. rs2830012 22. rs2830036 23. rs2830042 24. rs2830045 25. rs2830048 26. rs2830050 27. rs2830051 28. rs2830053 29. rs2830066 30. rs2830104 31. rs9305268	APP	Genetic - SNP	Blood	<sup>12</sup>	1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant 7. Non-significant 8. Non-significant 9. Non-significant 10. Non-significant 11. Non-significant 12. Non-significant 13. Non-significant 14. Non-significant 15. Non-significant 16. Non-significant 17. Non-significant 18. Non-significant 19. Non-significant 20. Non-significant 21. Non-significant 22. Non-significant 23. Non-significant 24. Non-significant 25. Non-significant 26. Non-significant 27. Non-significant 28. Non-significant 29. Non-significant 30. Non-significant 31. Non-significant	



32. rs9980729 33. rs9981258 34. rs11087985					32. Non-significant 33. Non-significant 34. Non-significant		
Angiotensin 2 receptor type 1 SNP rs5186 A>C	AT1	Genetic - SNP	Blood	70	Non-significant	70	Non-significant
Angiotensin 2 receptor type 2 SNP rs11091046 A>C	AT2	Genetic - SNP	Blood	70	Non-significant	70	Non-significant
Angiotensin 2 receptor type 2 SNP rs1403543 G>A	AT2	Genetic - SNP	Blood	70	Non-significant	70	Non-significant
Angiotensin converting enzyme SNP rs4340 I/D	ACE	Genetic - SNP	Blood	70	Non-significant	70	Non-significant
Angiotensinogen SNP rs699 C>T	AGT	Genetic - SNP	Blood	70	Non-significant	70	Non-significant
Brain-derived neurotrophic factor SNP Val66Met	BNDP	Genetic - SNP	Blood	171	Non-significant		
Ceruloplasmin SNP 1. rs701755 2. rs772908 3. rs773050 4. rs3816893 5. rs9853335 6. rs13072552 7. rs13075921 8. rs16861579 9. rs16861634 10. rs17787768 11. rs17838831	CP	Genetic - SNP	Blood	12	1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant 7. Non-significant 8. Non-significant 9. Non-significant 10. Non-significant 11. Non-significant		
Cubilin SNP 1. rs703064 2. rs780810 3. rs780837 4. rs780839 5. rs780846 6. rs796667 7. rs1276711 8. rs1276722 9. rs1398431 10. rs1687702 11. rs1797081 12. rs1914172 13. rs2087631 14. rs2130806 15. rs2356823	CUBN	Genetic - SNP	Blood	12	1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant 7. Non-significant 8. Non-significant 9. Non-significant 10. Non-significant 11. Non-significant 12. Non-significant 13. Non-significant 14. Non-significant 15. Non-significant		

16.	rs2603796				16. Non-significant
17.	rs2883972				17. Non-significant
18.	rs3012477				18. Non-significant
19.	rs3012501				19. Non-significant
20.	rs3740169				20. Non-significant
21.	rs3847364				21. Non-significant
22.	rs3847369				22. Non-significant
23.	rs3914238				23. Non-significant
24.	rs4335444				24. Non-significant
25.	rs6602163				25. Non-significant
26.	rs7071422				26. Non-significant
27.	rs7072049				27. Non-significant
28.	rs7075040				28. Non-significant
29.	rs7082270				29. Non-significant
30.	rs7098659				30. Non-significant
31.	rs7100290				31. Non-significant
32.	rs7893395				32. Non-significant
33.	rs7893507				33. Non-significant
34.	rs7897704				34. Non-significant
35.	rs7898283				35. Non-significant
36.	rs7900486				36. Non-significant
37.	rs7904146				37. Non-significant
38.	rs7906242				38. Non-significant
39.	rs7917400				39. Non-significant
40.	rs10508517				40. Non-significant
41.	rs10508518				41. Non-significant
42.	rs10752063				42. Non-significant
43.	rs10795440				43. Non-significant
44.	rs10795445				44. Non-significant
45.	rs10904831				45. Non-significant
46.	rs10904850				46. Significant
47.	rs11254235				47. Non-significant
48.	rs11254238				48. Non-significant
49.	rs11254248				49. Non-significant
50.	rs11254308				50. Non-significant
51.	rs11591673				51. Non-significant
52.	rs12146414				52. Non-significant
53.	rs12244831				53. Non-significant
54.	rs12354787				54. Non-significant
55.	rs17139666				55. Non-significant
56.	rs17347211				56. Non-significant

57. rs41335345 58. rs41517646 59. rs41526049						57. Non-significant 58. Non-significant 59. Non-significant	
Cystathionine b-synthase SNP 1. rs1801181 1080C>T 2. rs234706 699G>A, c.699C>T 3. 844ins68	CBS	Genetic - SNP	Blood	58		58	1. Non-significant 2. Non-significant 3. Non-significant
Divalent metal transporter 1 SNP 1. rs2928672 2. rs2978475 3. rs7015818 4. rs7833754 5. rs7834536 6. rs11781222 7. rs12548753 8. rs13266950	SLC11A2	Genetic - SNP	Blood	12			1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant 7. Non-significant 8. Non-significant
Duodenal cytochrome b SNP 1. rs12476341 2. rs13009270 3. rs13392902	CYBRD1	Genetic - SNP	Blood	12			1. Non-significant 2. Non-significant 3. Non-significant
Endothelin receptor type A SNP 1. rs5333 2. rs5335 3. rs5342 4. rs1801708 5. rs6537484 6. rs6812093 7. rs6841799 8. rs10008744 9. rs10305860	ET <sub>A</sub>	Genetic - SNP	Blood	57, 120		57	1. Non-significant 2. Significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant 7. Non-significant 8. Non-significant 9. Non-significant
Endothelin receptor type B SNP 1. rs5351 2. rs3027111 3. rs3818416 4. rs4885491 5. rs4885493 6. rs9574124	ET <sub>B</sub>	Genetic - SNP	Blood	57, 120		57	1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant
Endothelin-1 SNP 1. rs5369 2. rs5370 3. rs5371	ET1	Genetic - SNP	Blood	57, 120		57	1. Non-significant 2. Non-significant 3. Non-significant

<ol style="list-style-type: none"> <li>4. rs1476046</li> <li>5. rs1626492</li> <li>6. rs1800541</li> <li>7. rs2070699</li> <li>8. rs2071943</li> <li>9. rs3087459</li> <li>10. rs6912834</li> </ol>					<ol style="list-style-type: none"> <li>4. Non-significant</li> <li>5. Non-significant</li> <li>6. Non-significant</li> <li>7. Non-significant</li> <li>8. Non-significant</li> <li>9. Non-significant</li> <li>10. Non-significant</li> </ol>	<ol style="list-style-type: none"> <li>6. Non-significant</li> <li>7. Non-significant</li> </ol>
Epoxyeicosatrienoic acid metabolic pathway SNP <ol style="list-style-type: none"> <li>1. CYP2C8 SNP rs1058930 g.16136C&gt;G lle264Met</li> <li>2. CYP2J2 SNP rs1155002 g.30354600T&gt;G</li> </ol>	EET SNP  CYP2C8  CYP2J2	Genetic - SNP	Blood & CSF	<sup>98</sup>	<ol style="list-style-type: none"> <li>1. Significant</li> <li>2. Significant</li> </ol>	
Factor V Leiden SNP G1691A	FV Leiden	Genetic - SNP	Blood	<sup>155</sup>	Non-significant	
Feline Leukemia Virus Subgroup C Receptor SNP <ol style="list-style-type: none"> <li>1. rs963328</li> <li>2. rs6674933</li> <li>3. rs7553536</li> <li>4. rs12404754</li> </ol>	FLVCR1	Genetic - SNP	Blood	<sup>12</sup>	<ol style="list-style-type: none"> <li>1. Non-significant</li> <li>2. Non-significant</li> <li>3. Non-significant</li> <li>4. Non-significant</li> </ol>	
Mitochondrial ferritin SNP rs1560550	FTMT	Genetic - SNP	Blood	<sup>12</sup>	Non-significant	
Ferrochelatase SNP <ol style="list-style-type: none"> <li>1. rs317806</li> <li>2. rs317807</li> <li>3. rs492274</li> <li>4. rs533952</li> <li>5. rs1041951</li> <li>6. rs1736439</li> <li>7. rs1788002</li> <li>8. rs2269219</li> <li>9. rs8095390</li> <li>10. rs12454808</li> <li>11. rs12969847</li> </ol>	FECH	Genetic - SNP	Blood	<sup>12</sup>	<ol style="list-style-type: none"> <li>1. Non-significant</li> <li>2. Non-significant</li> <li>3. Non-significant</li> <li>4. Non-significant</li> <li>5. Non-significant</li> <li>6. Non-significant</li> <li>7. Non-significant</li> <li>8. Non-significant</li> <li>9. Non-significant</li> <li>10. Non-significant</li> <li>11. Non-significant</li> </ol>	
Frataxin SNP <ol style="list-style-type: none"> <li>1. rs1411675</li> <li>2. rs2498432</li> <li>3. rs3793451</li> <li>4. rs4069737</li> <li>5. rs7847872</li> <li>6. rs7851458</li> <li>7. rs7870295</li> <li>8. rs7871596</li> </ol>	FXN	Genetic - SNP	Blood	<sup>12</sup>	<ol style="list-style-type: none"> <li>1. Non-significant</li> <li>2. Non-significant</li> <li>3. Non-significant</li> <li>4. Non-significant</li> <li>5. Non-significant</li> <li>6. Non-significant</li> <li>7. Non-significant</li> <li>8. Non-significant</li> </ol>	

9. rs10126006 10. rs10869867 11. rs11145194 12. rs11790543 13. rs12001326						9. Non-significant 10. Non-significant 11. Non-significant 12. Non-significant 13. Non-significant		
Glutaredoxin 5 SNP 1. rs716315 2. rs1007813 3. rs4905335 4. rs10131473	GLRX5	Genetic - SNP	Blood	<sup>12</sup>		1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant		
Heme-oxygenase 1 SNP 1. rs2269533 2. rs2285112 3. rs5995097	HMOX1	Genetic - SNP	Blood	<sup>12</sup>		1. Non-significant 2. Non-significant 3. Non-significant		
Heme-oxygenase 2 SNP rs2160567	HMOX2	Genetic - SNP	Blood	<sup>12</sup>		Non-significant		
Hemoglobin scavenger receptor SNP 1. rs3210140 2. rs6488336 3. rs6488338 4. rs7487755 5. rs7954492 6. rs10743939 7. rs10845265 8. rs11054074 9. rs12304718	CD163	Genetic - SNP	Blood	<sup>12</sup>		1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant 7. Non-significant 8. Non-significant 9. Non-significant		
Hemopexin SNP rs2035675	HPX	Genetic - SNP	Blood	<sup>12</sup>		Non-significant		
Hepcidin antimicrobial peptide gene SNP 1. rs7251432 2. rs10421768	HAMP	Genetic - SNP	Blood	<sup>11</sup>		1. Non-significant 2. Non-significant		
Hephaestin SNP 1. rs1264215 2. rs6624875 3. rs12689282	HEPH	Genetic - SNP	Blood	<sup>12</sup>		1. Non-significant 2. Non-significant 3. Non-significant		
High-mobility group box 1 SNP rs2249825 3814C>G	HMGB1	Genetic - SNP	Blood	<sup>72</sup>		Non-significant	<sup>72</sup>	Significant
Human hemochromatosis protein SNP 1. rs707889 2. rs1800562 3. rs2071303	HFE	Genetic - SNP	Blood	<sup>12</sup>		1. Non-significant 2. Non-significant 3. Non-significant		
Iron responsive element binding protein 2 SNP	IREB2	Genetic - SNP	Blood	<sup>12</sup>				

<ol style="list-style-type: none"> <li>rs924840</li> <li>rs8041628</li> <li>rs11636431</li> <li>rs16969899</li> <li>rs17483721</li> </ol>					<ol style="list-style-type: none"> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> </ol>		
Low-density lipoprotein receptor related protein SNP <ol style="list-style-type: none"> <li>rs1799986</li> <li>rs2306692</li> <li>rs4759277</li> <li>rs7975818</li> <li>rs10876966</li> </ol>	LRP1	Genetic - SNP	Blood	12	<ol style="list-style-type: none"> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> </ol>		
Methylenetetrahydrofolate reductase SNP C677T	MTHFR	Genetic - SNP	Blood	155	Non-significant		
Endothelial Nitric oxide synthase SNP <ol style="list-style-type: none"> <li>27VNTR</li> <li>G894T</li> </ol>	eNOS SNP	Genetic - SNP	Blood	185	<ol style="list-style-type: none"> <li>Non-significant</li> <li>Non-significant</li> </ol>		
Endothelial Nitric oxide synthase SNP rs2070744 T-786C	eNOS SNP	Genetic - SNP	Blood	74, 185	Inconclusive	74	Significant
Endothelial Nitric oxide synthase SNP rs2070744 T-786C	eNOS SNP	Genetic - SNP	Buccal swab	162	Significant	162	Non-significant
Progesterone receptor membrane SNP <ol style="list-style-type: none"> <li>rs2428744</li> <li>rs2428757</li> <li>rs5909608</li> </ol>	PGRMC1	Genetic - SNP	Blood	12	<ol style="list-style-type: none"> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> </ol>		
Prothrombin SNP rs1799963 G20210A		Genetic - SNP	Blood	155	Non-significant		
Ryanodine Receptor 1 SNP rs35364374 c.6178G>T	RYR1	Genetic - SNP	Blood	73	Non-significant	73	Non-significant
Serine proteinase inhibitor family E member 1 SNP <ol style="list-style-type: none"> <li>rs6090 G/A</li> <li>rs6092 G/A</li> <li>rs7242 T/G</li> <li>rs2227631 G/A</li> <li>rs2227684 G/A</li> <li>rs1799889 4G/5G</li> <li>rs2227631 + rs1799889</li> <li>rs6092+rs6090+rs2227684+rs7242</li> <li>rs2227631+rs1799889+rs6092+rs6090+rs2227684+rs7242</li> </ol>	SERPINE1	Genetic - SNP	Blood	53, 71	<ol style="list-style-type: none"> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Significant</li> </ol>	53, 71	<ol style="list-style-type: none"> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Non-significant</li> <li>Significant</li> <li>Non-significant</li> <li>Significant</li> </ol>

Solute carrier family 11 member 1 SNP 1. rs2276631 2. rs2695343	SLC11A1	Genetic - SNP	Blood	<sup>12</sup>	1. Non-significant 2. Non-significant		
Solute carrier family 25 member 37 SNP (Mitoferrin 1) 1. rs150909 2. rs224446 3. rs224572 4. rs224589 5. rs2269683	SLC25A37	Genetic - SNP	Blood	<sup>12</sup>	1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant		
Solute carrier family 40 member 1 SNP (Ferroportin) 1. rs930373 2. rs1123109 3. rs1123110 4. rs1439816 5. rs2304704 6. rs4667287	SLC40A1	Genetic - SNP	Blood	<sup>12</sup>	1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant		
Solute carrier family 48 member 1 SNP 1. rs3782908 2. rs4760636	SLC48A1	Genetic - SNP	Blood	<sup>12</sup>	1. Non-significant 2. Non-significant		
STEAP3 metalloreductase SNP 1. rs838079 2. rs838083 3. rs838090 4. rs838102 5. rs838103 6. rs865688 7. rs1867749 8. rs1867856 9. rs3769659 10. rs6720040 11. rs10188946 12. rs11694139 13. rs12990907	STEAP3	Genetic - SNP	Blood	<sup>12</sup>	1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant 6. Non-significant 7. Non-significant 8. Non-significant 9. Non-significant 10. Non-significant 11. Non-significant 12. Non-significant 13. Non-significant		
Transferrin SNP 1. rs1049296 2. rs1799852 3. rs4241357 4. rs7645538 5. rs8177191	TF	Genetic - SNP	Blood	<sup>12</sup>	1. Non-significant 2. Non-significant 3. Non-significant 4. Non-significant 5. Non-significant		

6. rs8177201 7. rs8177224 8. rs8177235 9. rs8177248 10. rs8177277 11. 1rs12493168					6. Non-significant 7. Non-significant 8. Non-significant 9. Non-significant 10. Non-significant 11. Non-significant		
Transferrin receptor 1 SNP 1. rs3933 2. rs3804141 3. rs17788379	TFRC	Genetic - SNP	Blood	12	1. Non-significant 2. Non-significant 3. Non-significant		
Tumor necrosis factor a SNP 863C>A	TNF-α	Genetic - SNP	Blood	141	Non-significant		
<b>Inflammation and Immune system</b>							
CD45+ Hematopoietic cells	CD45+ HSC	Inflam & Immune System	Blood	27	Non-significant		
Calprotectine	S100A8/A9	Inflam. & Immune system	Blood	35	Significant		
C-C Motif Chemokine Ligand 2	CCL2	Inflam. & Immune system	Blood	47	Non-significant		
C-C Motif Chemokine Ligand 5	CCL5	Inflam. & Immune system	Blood	15, 47	Inconclusive		
C-C Motif Chemokine Ligand 5	CCL5	Inflam. & Immune system	CSF	15	Significant		
C-C Motif Chemokine Ligand 7	CCL7	Inflam. & Immune system	Blood	47	Non-significant		
<i>Soluble</i> CD40L ligand	sCD40L	Inflam. & Immune system	Blood	47, 100	Inconclusive	100	Non-significant
C-reactive protein	CRP	Inflam. & Immune system	Blood	35, 43, 75, 76, 131, 132, 136, 141, 146, 154, 175	Inconclusive	31, 37, 55, 65, 132	Significant
<i>high-sensitivity</i> C-reactive protein	hs-CRP	Inflam. & Immune system	Blood	46, 90	Non-significant	90	Non-significant
Erythrocyte sedimentation rate	ESR	Inflam. & Immune system	Blood	154	Non-significant		
E-selectin		Inflam. & Immune system	Blood	76, 151, 176, 193	Inconclusive		
E-selectin		Inflam. & Immune system	CSF	183	Non-significant		
Ficolin-1	FCN1	Inflam. & Immune system	Blood	34, 113	Non-significant	113	Non-significant
Ficolin-1	FCN1	Inflam. & Immune system	CSF	34	Non-significant		
Ficolin-2	FCN2	Inflam. & Immune system	Blood	34, 113	Non-significant	113	Non-significant
Ficolin-2	FCN2	Inflam. & Immune system	CSF	34	Non-significant		
Ficolin-3	FCN3	Inflam. & Immune system	Blood	34, 113	Inconclusive	113	Significant
Ficolin-3	FCN3	Inflam. & Immune system	CSF	34	Non-significant		
Galectin-3	Gal3	Inflam. & Immune system	Blood	45	Non-significant	45	Non-significant
Granulocyte colony-stimulating factor	G-CSF or CSF3	Inflam. & Immune system	Blood	47	Non-significant		
Granulocyte-macrophage colony-stimulating factor	GM-CSF or CSF2	Inflam. & Immune system	Blood	47	Non-significant		
<i>Soluble</i> Growth Stimulation Expressed Gene 2	sST2	Inflam. & Immune system	Blood	41	Significant		
High-mobility group box 1	HMGB1	Inflam. & Immune system	Blood	135	Significant	68	Non-significant



Interleukin-1 receptor antagonist	IL-1Ra	Inflam. & Immune system	Blood	47	Non-significant		
Interleukin-1B	IL-1B	Inflam. & Immune system	Blood	47, 202	Non-significant		
Interleukin-1B	IL-1B	Inflam. & Immune system	CSF	189	Significant		
Interleukin-2	IL-2	Inflam. & Immune system	Blood	47	Non-significant		
Interleukin-5	IL-5	Inflam. & Immune system	Blood	47	Non-significant		
Interleukin-6	IL-6	Inflam. & Immune system	Blood	46, 47, 76, 80, 107, 202	Inconclusive	80	Significant
Interleukin-6	IL-6	Inflam. & Immune system	CSF	27, 97, 142, 189	Inconclusive	170	Significant
Interleukin-6	IL-6	Inflam. & Immune system	MD			103	Significant
Interleukin-8	IL-8	Inflam. & Immune system	Blood	46, 47	Non-significant		
Interleukin-9	IL-9	Inflam. & Immune system	Blood	47	Non-significant		
Interleukin-10	IL-10	Inflam. & Immune system	Blood	47	Non-significant	16	Non-significant
Interleukin-17	IL-17	Inflam. & Immune system	Blood	79	Significant	79	Significant
Interleukin-23	IL-23	Inflam. & Immune system	Blood	79	Non-significant	79	Non-significant
Interferon alpha-2	IFN-a2	Inflam. & Immune system	Blood	47	Non-significant		
Interferon gamma	IFN-y	Inflam. & Immune system	Blood	46, 47	Non-significant		
Interferon gamma-induced protein 10	IP-10 or CXCL10	Inflam. & Immune system	Blood	47	Significant		
Intercellular Adhesion Molecule 1	ICAM-1	Inflam. & Immune system	Blood	46, 76, 151, 174, 176, 193	Inconclusive		
Intercellular Adhesion Molecule 1	ICAM-1	Inflam. & Immune system	CSF	174	Non-significant		
Functional Lectin complement pathway activity	LCP activity	Inflam. & Immune system	Blood	113	Significant	113	Significant
Leukemia inhibitor factor	LIF	Inflam. & Immune system	Blood	76	Non-significant		
Leukocytes	WBC	Inflam. & Immune system	Blood	28, 51, 52, 69, 76, 90, 94, 136, 141, 151, 154, 168, 175, 193, 201	Inconclusive	31, 37, 65, 90, 94, 134, 147	Inconclusive
L-selectin		Inflam. & Immune system	Blood	151, 193	Significant		
Monocyte lymphocyte function-associated antigen-1	LFA-1	Inflam. & Immune system	Blood			134	Significant
Neutrophil lymphocyte function-associated antigen-1	LFA-1	Inflam. & Immune system	Blood			134	Significant
Lymphocytes		Inflam. & Immune system	Blood	28, 51, 52, 69, 151	Inconclusive		
CD8+CD161+ "Tc17" lymphocytes		Inflam. & Immune system	CSF	27	Significant		
Monocyte macrophage antigen-1	Mac-1	Inflam. & Immune system	Blood			134	Significant
Neutrophil macrophage antigen-1	Mac-1	Inflam. & Immune system	Blood			134	Significant
Macrophage CD163 expression	CD163	Inflam. & Immune system	CSF			62	Non-significant
Macrophage imigration inhibitory factor	MIF	Inflam. & Immune system	CSF	50	Significant		
Macrophage inflammatory protein 1a	MIP-1a	Inflam. & Immune system	Blood	47	Significant		

Macrophage inflammatory protein 1b	MIP-1b	Inflam. & Immune system	Blood	47	Non-significant		
Mannose-binding lectin	MBL	Inflam. & Immune system	Blood	34, 113	Non-significant	113	Non-significant
Mannose-binding lectin	MBL	Inflam. & Immune system	CSF	34	Non-significant		
Matrix metalloproteinase-3	MMP-3	Inflam. & Immune system	Blood			126	Non-significant
Matrix metalloproteinase-9	MMP-9	Inflam. & Immune system	Blood	78	Non-significant	126	Significant
Matrix metalloproteinase-9	MMP-9	Inflam. & Immune system	CSF	27	Non-significant		
Matrix metalloproteinase-9	MMP-9	Inflam. & Immune system	MD	140	Non-significant	103	Non-significant
Pro-matrix metalloproteinase-9	Pro-MMP9	Inflam. & Immune system	MD	140	Significant		
<i>CD45<sup>dim</sup>CD11b<sup>+</sup></i> Microglia		Inflam. & Immune system	CSF	27	Non-significant		
Monocytes		Inflam. & Immune system	Blood	151	Non-significant		
<i>Activated CD16<sup>bright</sup>CD56<sup>dim</sup></i> cytotoxic natural killer cells		Inflam. & Immune system	CSF	84	Significant		
<i>CD3-CD161<sup>+</sup></i> + natural killer cells		Inflam. & Immune system	CSF	27	Non-significant		
Netrin-1		Inflam. & Immune system	Blood	44	Non-significant		
Neutrophil/albumin ratio	NAR	Inflam. & Immune system	Blood	8	Significant		
Neutrophil/lymphocyte ratio	NLR	Inflam. & Immune system	Blood	28, 38, 52, 69	Significant	38, 40	Inconclusive
Neutrophil elastase	NE	Inflam. & Immune system	Blood	202	Significant		
Neutrophils		Inflam. & Immune system	Blood	28, 52, 69, 75, 151	Inconclusive		
(Neutrophil/lymphocyte) / (platelet/lymphocyte) ratio	NLR-PLR	Inflam. & Immune system	Blood	69	Significant		
Osteopontin	OPN	Inflam. & Immune system	Blood	59	Significant	59	Significant
Pentraxin 3	PTX3	Inflam. & Immune system	Blood	142	Non-significant		
Pentraxin 3	PTX3	Inflam. & Immune system	CSF	142	Significant		
Periostin	POSTN	Inflam. & Immune system	Blood	43	Significant	45	Non-significant
Platelet-endothelial adhesion molecule	PECAM	Inflam. & Immune system	Blood	193	Non-significant		
Platelet/lymphocyte ratio	PLR	Inflam. & Immune system	Blood	69	Significant		
Procalcitonin	PCT	Inflam. & Immune system	Blood	14	Significant	31, 37	Non-significant
P-selectin		Inflam. & Immune system	Blood	107, 151, 176, 177, 193	Significant		
<i>Monocyte</i> P-selectin glycoprotein ligand-1	PSGL-1	Inflam. & Immune system	Blood			134	Significant
<i>Neutrophil</i> P-selectin glycoprotein ligand-1	PSGL-1	Inflam. & Immune system	Blood			134	Significant
<i>cleaved</i> Receptor for advanced glycation end-products	cRAGE	Inflam. & Immune system	Blood	60	Significant		
<i>soluble</i> Receptor for advanced glycation end-products	sRAGE	Inflam. & Immune system	Blood	54	Significant		
<i>CD3<sup>+</sup></i> T cells		Inflam. & Immune system	CSF	27	Non-significant		
Tenascin-C	TNC	Inflam. & Immune system	Blood	67	Significant		
Tenascin-C	TNC	Inflam. & Immune system	CSF	108	Significant		
Tissue inhibitors of metalloproteinases-3	TIMP-3	Inflam. & Immune system	Blood			126	Non-significant

Tumor necrosis factor a	TNF- $\alpha$	Inflam. & Immune system	Blood	46, 47, 141, 202	Non-significant		
Tumor necrosis factor a	TNF- $\alpha$	Inflam. & Immune system	CSF	27, 97, 142	Inconclusive		
Vascular cell adhesion protein 1	VCAM-1	Inflam. & Immune system	Blood	46, 151, 174, 193	Inconclusive		
Vascular cell adhesion protein 1	VCAM-1	Inflam. & Immune system	CSF	174	Non-significant		
Alkaline phosphotase	AP	Inflam. & Immune system, Vascular tone	Blood	42	Significant	42	Non-significant
<b>Oxidative stress</b>							
8-iso-Prostaglandin F <sub>2a</sub> (free)	8-iso-PGF <sub>2a</sub>	Oxidative stress	CSF	150	Significant		
8-iso-Prostaglandin F <sub>2a</sub> (free + esterified)	8-iso-PGF <sub>2a</sub>	Oxidative stress	CSF			181	Significant
Bilirubin		Oxidative stress	Blood	35	Non-significant		
Ceruloplasmin	CP	Oxidative stress	CSF	115	Non-significant		
Diacron reactive oxygen metabolites	d-ROMs	Oxidative stress	Blood			117	Significant
Diacron reactive oxygen metabolites	d-ROMs	Oxidative stress	CSF			117	Non-significant
F2-isoprostane	F2-IsoPs	Oxidative stress	Urine	22, 83	Significant		
Ferritin		Oxidative stress	CSF	115	Non-significant		
Gamma-glutamyl transferase	GGT	Oxidative stress	Blood	85	Non-significant		
Pro-Hepcidin		Oxidative stress	CSF	115	Non-significant		
Free Hydroxyecosatetraenoic acids	HETE	Oxidative stress	CSF	150	Non-significant		
Free Hydroxyoctadecadienoic acids	HODE	Oxidative stress	CSF	150	Non-significant		
Iron		Oxidative stress	Blood	39	Significant		
Iron		Oxidative stress	MD			3	Non-significant
non-transferrin bound Iron	NTBI	Oxidative stress	CSF	115	Significant		
Lactoferrin	LF	Oxidative stress	CSF	115	Non-significant		
Reactive oxygen species	ROS	Oxidative stress	Blood	89	Significant		
Redox-active iron	REDOX-Fe	Oxidative stress	CSF	115	Non-significant		
Total antioxidant capacity	TAC	Oxidative stress	Blood	5	Non-significant		
Total antioxidant capacity	TAC	Oxidative stress	CSF	5	Significant		
Transferrin	TF	Oxidative stress	CSF	115, 199	Inconclusive		
Zinc		Oxidative stress	Blood	29	Significant		
<b>Vascular tone</b>							
20-Hydroxyecosatetraenoic acid	20-HETE	Vascular tone	CSF	99	Significant		
6-keto-prostaglandin F <sub>1a</sub>	6-keto-PGF <sub>1a</sub>	Vascular tone	CSF	209	Non-significant		
Asymmetric dimethyl-arginine	ADMA	Vascular tone	CSF	139	Non-significant		
Big endothelin-1	Big ET-1	Vascular tone	Blood	192	Significant	192, 198	Non-significant
Big endothelin-1	Big ET-1	Vascular tone	CSF			198	Significant
Calcitonin gene-related peptide	CGRP	Vascular tone	CSF	208	Non-significant		
Calcium	Ca	Vascular tone	Blood	161	Non-significant		
Carbon dioxide partial pressure	pCO <sub>2</sub>	Vascular tone	Blood			6, 147	Inconclusive
Dihydroxyecosatetraenoic acid	DHET	Vascular tone	CSF	98	Non-significant		

Endothelin-1	ET1	Vascular tone	Blood	180, 192	Significant	180, 192, 197	Non-significant
Endothelin-1	ET1	Vascular tone	CSF	139, 180, 194	Inconclusive	180	Non-significant
Endothelin-1	ET1	Vascular tone	MD	180	Non-significant	180	Non-significant
Endothelin-1/big endothelin-1 ratio	ET1/big-ET1	Vascular tone	Blood	192	Non-significant	192	Non-significant
Endothelin-1/nitric oxide ratio	ET1/NO	Vascular tone	Blood	123	Non-significant		
Endothelin-converting enzyme 1 activity	ECE-1	Vascular tone	Blood	192	Non-significant	192	Non-significant
Epoxyeicosatrienoic acid	EET	Vascular tone	CSF	98	Non-significant		
Leukotriene C4	LTC4	Vascular tone	CSF	209	Significant		
Magnesium	Mg	Vascular tone	Blood	165, 188, 190	Inconclusive		
Neuropeptide Y	NPY	Vascular tone	Blood	93	Significant		
Neuropeptide Y	NPY	Vascular tone	CSF	208	Non-significant		
Parathyroid hormone	PTH	Vascular tone	Blood	161	Non-significant		
Prostaglandin D2	PGD2	Vascular tone	CSF	209	Significant		
Thromboxane B2	TxB2	Vascular tone	CSF	209	Non-significant		
Adrenomedulin	ADM	Vascular tone	Blood	110	Significant		
<i>Bioactive</i> Adrenomedulin	Bio-ADM	Vascular tone	Blood	4	Significant		
<i>Bioactive</i> Adrenomedulin	Bio-ADM	Vascular tone	CSF	4	Significant		
<b>Volemic status</b>							
Albumin		Volemic status	Blood	28, 35, 51, 96	Inconclusive		
Aldosterone		Volemic status	Blood	204	Non-significant		
Antidiuretic hormone	ADH	Volemic status	Blood	204	Non-significant		
Atrial natriuretic peptide	ANP	Volemic status	Blood	204	Non-significant		
Atrial natriuretic peptide	ANP	Volemic status	CSF	208	Non-significant		
Brain natriuretic peptide	BNP	Volemic status	Blood	186	Significant	133, 148	Significant
Copeptin	CP	Volemic status	Blood	146	Significant		
Renin		Volemic status	Blood	204	Non-significant		
Sodium		Volemic status	Blood	1, 28, 35, 116, 129, 137, 167, 191, 204	Inconclusive	71, 116, 145, 147	Significant
Sodium		Volemic status	Urine	121, 167, 204	Inconclusive		
<b>Other</b>							
<i>Soluble</i> Endoglin	sEng	Angiogenesis	Blood			82, 143	Significant
<i>Soluble</i> Endoglin	sEng	Angiogenesis	CSF			82	Non-significant
<i>Soluble</i> Fms-like tyrosine kinase 1	sFlt-1	Angiogenesis	Blood			82	Non-significant
<i>Soluble</i> Fms-like tyrosine kinase 1	sFlt-1	Angiogenesis	CSF			82	Non-significant
Thrombospondin-1	TSP-1	Angiogenesis	Blood	101, 131	Inconclusive	101	Non-significant
Hematocrit	Ht	Blood viscosity	Blood	35, 138, 201	Non-significant	147	Non-significant
Cardiac troponin 1	cTnl	Cardiac	Blood			133	Non-significant
Creatine kinase-myoglobin binding	CK-MB	Cardiac	Blood	200	Non-significant		
<i>Heart-Fatty acid-binding protein</i>	H-FABP	Cardiac	CSF	130	Non-significant	130	Non-significant

Glucose		Cerebral metabolism	MD	187	Non-significant	21	Significant
Lactate		Cerebral metabolism	MD	114, 187	Significant		
Lactate/pyruvate ratio	LPR	Cerebral metabolism	MD	114, 187	Inconclusive	21, 26	Significant
Estradiol	E1	Gender	Blood	102	Significant		
Estrone	E1	Gender	Blood	102	Significant		
Fibroblast growth factor 2	FGF-2	Growth factors	Blood	47	Non-significant		
Platelet derived growth factor AA	PDGF-AA	Growth factors	Blood	47	Non-significant		
Platelet derived growth factor AB/BB	PDGF-ABBB	Growth factors	Blood	47	Significant		
Transforming growth factor a	TGF-a	Growth factors	Blood	47	Non-significant		
Transforming growth factor B1	TGF-B1	Growth factors	Blood			143	Non-significant
Vascular endothelial growth factor	VEGF	Growth factors	CSF	27	Non-significant		
Apolipoprotein A1	ApoA	Lipid metabolism	Blood	131	Significant		
Total cholesterol	TC	Lipid metabolism	Blood	51	Significant	32, 134	Non-significant
Fatty acid-binding protein 4	FABP4	Lipid metabolism	Blood	13	Significant		
High-density lipoprotein	HDL	Lipid metabolism	Blood			32	Significant
Low-density lipoprotein	LDL	Lipid metabolism	Blood			32	Non-significant
Triglycerides	TG	Lipid metabolism	Blood	94	Non-significant	32, 94	Non-significant
Base excess	BE	Metabolism	Blood			147	Non-significant
Hydrogencarbonate	HCO <sub>3</sub> <sup>-</sup>	Metabolism	Blood			147	Non-significant
Lactate		Metabolism	Blood			95, 147	Non-significant
pH	pH	Metabolism	Blood			147	Non-significant
Glycated Hemoglobin	HbA1C	Metabolism	Blood	7, 77	Inconclusive	77	Non-significant
Hemoglobin	Hb	Oxygen-carrying capacity	Blood	20, 28, 35, 56, 69, 106, 152, 159	Significant	61, 147, 152	Inconclusive
Hemoglobin	Hb	Oxygen-carrying capacity	CSF	115	Non-significant		
Oxygen partial pressure	pO <sub>2</sub>	Oxygen-carrying capacity	Blood			147	Non-significant
Red blood cells	RBC	Oxygen-carrying capacity	Blood	151	Non-significant	134	Non-significant
Red cell distribution width	RCDW	Oxygen-carrying capacity	Blood	28	Non-significant		
Cortisol		Stress response	Blood	123, 125, 156	Significant		
Free Cortisol		Stress response	Blood	123	Significant		
Cortisol-binding globulin	CBG	Stress response	Blood	123	Non-significant		
Glucose	BG	Stress response	Blood	28, 35, 69, 77, 94, 119, 160, 184	Inconclusive	65, 77, 94, 95, 118, 134, 147, 184	Inconclusive
Glucose/potassium ratio		Stress response	Blood			65	Significant
Potassium		Stress response	Blood	35	Non-significant	65, 147	Inconclusive
Adrenocorticotrophic hormone	ACTH	Stress response	Blood	123	Non-significant		
Leptin		Nutrition	Blood	9	Non-significant		
Leptin		Nutrition	CSF	9	Non-significant		
circulatory Dipeptidyl peptidase 3	cDPP3	Other	Blood	10	Significant		

Hyaluronidase	Hyal	Other	Blood	164	Non-significant		
Hypocretin-1		Other	CSF	182	Significant		
Pituitary adenylate cyclase-activating polypeptide	PACAP	Other	Blood	92	Significant		
Pituitary polypeptide 7B2	7B2	Other	CSF	208	Non-significant		
Total protein		Other	CSF	182	Significant		
Urea/creatinine ratio	UCR	Other	Blood	2	Significant	2	Significant

# Biomarker-specific figures of biomarkers of clinical DCI which have been studied by more than one study

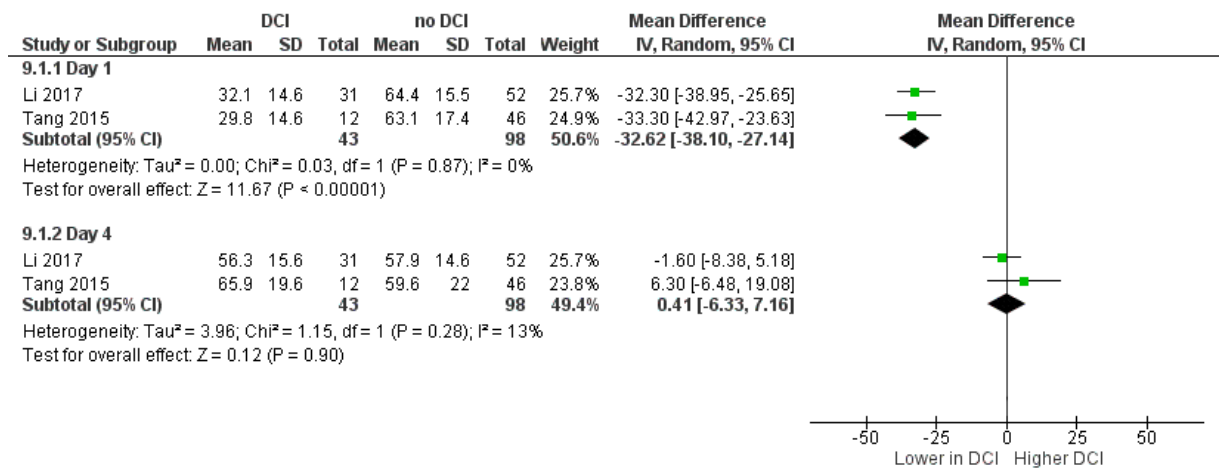
**Figure S1: ADAM Metallopeptidase with Thrombospondin Type 1 Motif 13 (blood)**

Study	DCI	no DCI	Comparison
Li, 2017 <sup>87,†</sup>	31	52	Mean ± SD day 1, 4, 10
Tang, 2015 <sup>107, †</sup>	12	46	Mean ± SD day 1, 4, 9
Vergouwen, 2009 <sup>158</sup>	11	20	Trend in time during day 2, 4, 7, 10, 14, 17

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↓ <sup>†</sup>	ns <sup>†</sup>	ns	
↓ <sup>†</sup>	ns <sup>†</sup>	ns	
sign. bigger decrease			

† meta-analysis possible of 2 of 2 studies with measurements on day 1 and 4, see below

**Figure S1.1:**



**Figure S2: Albumin (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	DCI in hypoalbuminemia & no hypoalbuminemia in pre-operative levels (threshold: unknown)
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Qi, 2019 <sup>51</sup>	53	199	Mean (SD) <24h
Behrouz, 2016 <sup>96</sup>	54	88	% DCI in hypoalbuminemia & normal albumin at admission (threshold: 3.4)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑DCI in hypo-			
ns			
ns*			
ns			

**Figure S3: C-C Motif Chemokine Ligand 5 (blood)**

Study	DCI	no DCI	Comparison
Chaudhry, 2020 <sup>15</sup>	28	52	Median (IQR) on day 1 and 7
Ahn, 2019 <sup>47</sup>	14	46	Median (IQR) <24h, 24-48h, 3-5 and 6-8 days

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns/↑*	↑*/ns		

**Figure S4: soluble CD40 ligand (blood)**

Study	DCI	no DCI	Comparison
Ahn, 2019 <sup>47</sup>	14	46	Median (IQR) <24h, 24-48h, 3-5 and 6-8 days
Chen, 2015 <sup>100</sup>	36	84	Levels at admission

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
significant			

**Figure S5: Cortisol (blood)**

Study	DCI	no DCI	Comparison
July, 2013 <sup>123</sup>	29	15	Mean (SD) pre-operative and post-operative day 2, 4, 7, 10
Lanterna, 2013 <sup>125</sup>	12	14	% DCI in hypo- & normocortisolism between day 1-15 (threshold: 10) Mean on day 5
Vergouwen, 2010 <sup>156</sup>	11	20	Trend in time during day 2, 4, 7, 10, 14, 17

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑*			
ns			
	↑		
↑			

**Figure S6: C-reactive protein (blood)**

Study	DCI	no DCI	Comparison
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Kanamaru, 2019 <sup>43</sup>	16	93	Mean (SEM) on day 1-3, 4-6, 7-9, 10-12 after onset
Chamling, 2017 <sup>76</sup>	16	65	Mean <24h Maximum during day 0, 1, 4, 7, 10, 14
Frontera, 2017 <sup>75</sup>	?	?	Levels at 0-24, 24-48 & 48-72h
Nyquist, 2013 <sup>131</sup>	14	14	Antibody array by ELISA on day (0-3)
Jeon, 2012 <sup>136</sup>	26	65	Mean (SD) levels pre-operative and post-operative day (1-2) & (3-7)
Juvela, 2012 <sup>132</sup>	no DCI: 132 RIND <sup>1</sup> : 17 FIND <sup>1</sup> : 29		Mean (SD) at admission, after surgery (mean day 2), at discharge (mean day 10)
Zhu, 2011 <sup>146</sup>	131	172	Mean at admission
Beeftink, 2011 <sup>141</sup>	10	57	% DCI in high & low mean levels of day 0-12 (threshold: median) % DCI in high & low mean levels of day 0-21 (threshold: median)
Kasius, 2010 <sup>154</sup>	11	40	Levels <72h Difference "before → after DCI onset" vs. paired samples in non-DCI
Rothoerl, 2006 <sup>175</sup>	37	51	Mean on day 1-8

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns*			
ns	ns	ns	
↑*			
maximum: ns*			
ns			
↑			
↑*/ns	↑		
ns		ns	
ns*			
↑DCI in high CRP*			
ns*			
ns*			
ns difference during DCI onset			
ns	ns/↑		

<sup>1</sup> RIND: reversible ischemic neurological deficit, FIND: fixed ischemic neurological deficit

**Figure S7: high-sensitivity C-reactive protein (blood)**

Study	DCI	no DCI	Comparison
Rasmussen, 2019 <sup>46</sup>	24	65	Mean (95% C.I.) on day 3 and 8. Ratio day 8/3
Srinivasan, 2016 <sup>90</sup>	94	152	Mean (SD) levels at admission

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns			
ns			

**Figure S8: D-dimer (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	Pre-operative levels
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Fukuda, 2017 <sup>86</sup>	24	163	Median (IQR) at admission
Zhu, 2011 <sup>146</sup>	131	172	Mean (SD) at admission
Tseng, 2007 <sup>166</sup>	14	66	Mean (SD) day 6 decrease
Juvela, 2006 <sup>173</sup>	37	99	Median (IQR) after surgery & at discharge
Fuji, 1997 <sup>201</sup>	18	99	Mean (SD) on day 0, 3, 6, 14, 30

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			
ns			
ns*			
sign. bigger decrease			
ns		↑	
ns/↑	ns		↑/ns

**Figure S9: Endothelin-1 (blood)**

Study	DCI	no DCI	Comparison
Kastner, 2005 <sup>180</sup>	7	13	Medians day 1-8
Juvela, 2002 <sup>192</sup>	13	37	Mean (SD) at admission, after surgery (mean day 2) and at discharge (mean day 10)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
↑*		↑*	



**Figure S10: Endothelin-1 (CSF)**

Study	DCI	no DCI	Comparison
Jung, 2012 <sup>139</sup>	4	20	Mean (SD) on day 0-14
Kastner, 2005 <sup>180</sup>	7	13	Medians on day 1-8
Mascia, 2001 <sup>194</sup>	4	16	Levels on day 1, 4, 7

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns	ns
ns	ns		
ns	ns/↑		

**Figure S11: Endothelin-1 SNP rs1800541**

Study	DCI	no DCI	Comparison
Griessenauer 2018 <sup>57</sup>	34	115	T/G alleles
Gallek, 2013 <sup>120</sup>	113	122	genotype TT/GT/GG

Results DCI per allele			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			

**Figure S12: Endothelin-1 SNP rs2070699**

Study	DCI	no DCI	Comparison
Griessenauer 2018 <sup>57</sup>	34	115	G/T alleles
Gallek, 2013 <sup>120</sup>	113	122	GG/TT/GT alleles

Results DCI per allele			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			

**Figure S13: E-selectin (blood)**

Study	DCI	no DCI	Comparison
Chamling, 2017 <sup>76</sup>	16	65	Maximum during day 0, 1, 4, 7, 10, 14
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24h
			Difference "before → after DCI" vs. paired samples in non-DCI
Frijns, 2006 <sup>176</sup>	12	55	DCI in high & low levels <72h (threshold 41.5)
Nissen, 2001 <sup>193</sup>	13	23	Mean (SE) of means during hospitalization

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
maximum: ns*			
ns			
sign. bigger increase during DCI onset			
ns*			
ns			

**Figure S14: F2-isoprostane (urine)**

Study	DCI	no DCI	Comparison
Wisniewski, 2020 <sup>22</sup>	19	19	Mean on day 3
			Mean and peak of day 2-6
Wisniewski, 2017 <sup>83</sup>	9	11	Mean on day 3
			Mean and peak of day 2-6

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑			
ns			
↑			
ns			

Cohorts were independent

**Figure S15: Fibrinogen (blood)**

Study	DCI	no DCI	Comparison
Zhang, 2021 <sup>8</sup>	84	355	Median (IQR) at admission
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Tseng, 2007 <sup>166</sup>	14	66	Mean increase from admission to day 3
Fuji, 1997 <sup>201</sup>	18	99	Mean (SD) on day 0, 3, 6, 14, 30

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			
sign. bigger increase			
ns/↑	↑		ns

**Figure S16: Ficolin-1 (blood)**

Study	DCI	no DCI	Comparison
Matzen, 2020 <sup>34</sup>	13	20	Mean (SD) per day, day 0-9 after SAH
			Trend in time from day 0 – 9 after SAH
Zanier, 2014 <sup>113</sup>	14	25	Median (IQR) of early (day 1-3, median: 2) and late (day 4-14, median: 7) sample

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns	
ns*			
ns	ns		

**Figure S17: Ficolin-2 (blood)**

Study	DCI	no DCI	Comparison
Matzen, 2020 <sup>34</sup>	13	20	Mean (SD) per day, day 0-9 after SAH
			Trend in time from day 0 – 9 after SAH
Zanier, 2014 <sup>113</sup>	14	25	Median (IQR) of early (day 1-3, median: 2) and late (day 4-14, median: 7) sample

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns	
ns*			
ns	ns		

**Figure S18: Ficolin-3 (blood)**

Study	DCI	no DCI	Comparison
Matzen, 2020 <sup>34</sup>	13	20	Mean (SD) per day, day 0-9 after SAH
			Trend in time from day 0 – 9 after SAH
Zanier, 2014 <sup>113</sup>	14	25	Median (IQR) of early (day 1-3, median: 2) and late (day 4-14, median: 7) sample

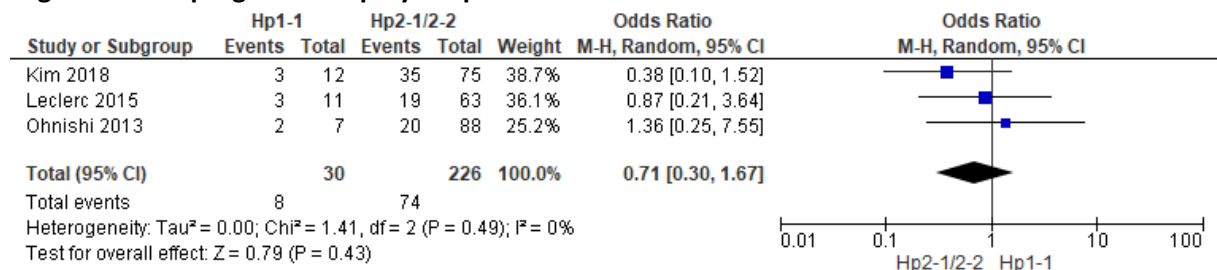
Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns/↑	↑/ns	ns	
ns*			
ns	↓		

**Figure S19: Glucose (blood)**

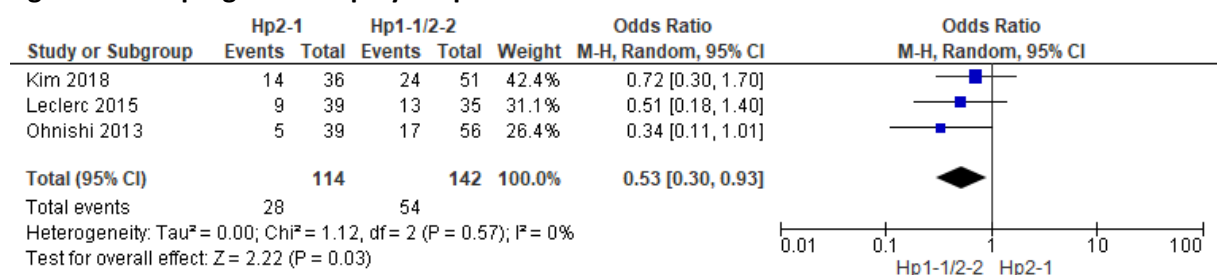
Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	% DCI in high & low pre-operative levels (threshold: 6.1)
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Beseoglu, 2017 <sup>77</sup>	36	51	Relative risk for DCI in hyperglycemia vs. no hyperglycemia <72h (threshold: 7.9)
Tao, 2017 <sup>69</sup>	47	200	Median (IQR) <24h
Mijiti, 2016 <sup>94</sup>	99	244	% DCI in high & low levels at admission (threshold: 7.0)
Bian, 2013 <sup>119</sup>	34	194	% DCI in patients with increasing and decreasing glucose between admission and day 14
Kruyt, 2008 <sup>160</sup>	86	169	% DCI in high & low glucose <48h (threshold: median)
			% DCI in high & low mean fasting glucose in the first week (threshold: median)
Juvela, 2005 <sup>184</sup>	No DCI: 128 RIND*: 17 FIND*: 29		Mean (SD) at admission & after surgery (mean day 1.9)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			
ns			
↑*			
ns*			
ns			
ns*			
↑*			
ns			

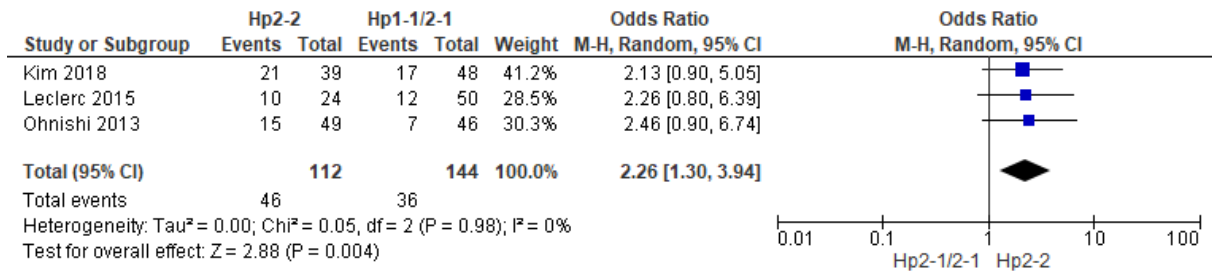
**Figure S20: Haptoglobin 1-1 polymorphism**



**Figure S21: Haptoglobin 2-1 polymorphism**



**Figure S22: Haptoglobin 2-2 polymorphism**



**Figure S23: Hematocrit (blood)**

Study	DCI	no DCI	Comparison
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Watanabe, 2012 <sup>138</sup>	6	34	Mean (SEM) on 2 days pre-DCI, 1 day pre-DCI and day of DCI-onset <sup>1</sup>
Fuji, 1997 <sup>201</sup>	18	99	Mean (SD) on day 0, 3, 6, 14, 30

<sup>1</sup> day of DCI-onset is used for both the DCI and no DCI group, calculated by the mean day of DCI occurrence

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			
ns	ns		ns

**Figure S24: Hemoglobin (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	Pre-operative levels
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Park, 2020 <sup>20</sup>	18	34	Mean (SD), timing of measurement unknown
Ayling, 2018 <sup>56</sup>	72	303	% DCI in high or low levels on day 1-3 and day 5-9. Threshold: 10 g/dL.
Tao, 2017 <sup>69</sup>	47	200	Median (IQR) <24h
Sun, 2015 <sup>106</sup>	78	140	% DCI in mean (post-operative to day 21) levels categorized into ≤11, 11-12, 12-13, >13
Naidech, 2010 <sup>152</sup>	10	34	% DCI in goal level ≥11.5 and ≥10, day 0-13
Oddo, 2009 <sup>159</sup>	8	12	Mean (SD) of day (1-7)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			
ns			
↑ DCI in low	↑ DCI in low		
ns			
Hb-group 11-12: significantly lower risk on DCI compared to other Hb groups			
ns			
↓			

**Figure S25: glycated Hemoglobin (blood)**

Study	DCI	no DCI	Comparison
Park, 2021 <sup>7</sup>	47	463	% DCI in high & low levels at admission (threshold: 6.0%)
Beseoglu, 2017 <sup>77</sup>	36	51	% DCI in high & low levels <72h (threshold: 39 mmol/mol)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑			
ns			

**Figure S26: Intercellular Adhesion Molecule 1 (blood)**

Study	DCI	no DCI	Comparison
Rasmussen, 2019 <sup>46</sup>	24	65	Mean (95% C.I.) on day 3 and 8. Ratio day 8/3
Chamling, 2017 <sup>76</sup>	16	65	Levels <24h Delta (maximum – baseline) value of day 0-14
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24h Difference “before → after DCI” vs. paired samples in non-DCI
Frijns, 2006 <sup>176</sup>	12	55	DCI in high & low levels <72h (threshold 376.5)
Rothoerl, 2006 <sup>174</sup>	5	10	Mean level on day 1-9 Maximum level during day 1-9
Nissen, 2001 <sup>193</sup>	13	23	Mean (SE) of means during hospitalization

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns			
ns			
delta: ↓*			
↓			
ns difference during DCI onset			
ns*			
ns	ns	ns	
maximum: ns			
ns			

**Figure S27: Interferon gamma (blood)**

Study	DCI	no DCI	Comparison
Rasmussen, 2019 <sup>46</sup>	24	65	Mean (95% C.I.) on day 3 and 8. Ratio day 8/3
Ahn, 2019 <sup>47</sup>	14	46	Median (IQR) <24h, 24-48h, 3-5 and 6-8 days

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns			
ns	ns*		

**Figure S28: Interleukin-1B (blood)**

Study	DCI	no DCI	Comparison
Ahn, 2019 <sup>47</sup>	14	46	Median (IQR) <24h, 24-48h, 3-5 and 6-8 days
Hirashima, 1997 <sup>202</sup>	9	12	Mean (SD) on day 0-4, 5-9, 10-14

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns	ns	ns	

**Figure S29: Interleukin-6 (blood)**

Study	DCI	no DCI	Comparison
Rasmussen, 2019 <sup>46</sup>	24	65	Mean (95% C.I.) on day 3 and 8. Ratio day 8/3
Ahn, 2019 <sup>47</sup>	14	46	Median (IQR) <24h, 24-48h, 3-5 and 6-8 days
Chamling, 2017 <sup>76</sup>	16	65	Levels <24h Maximum and delta (maximum – baseline) value of day 0-14
Chaudhry, 2017 <sup>80</sup>	28	52	Mean (SE) on day 1, 3, 5, 7, 9, 11, 13
Tang, 2015 <sup>107</sup>	12	46	Mean (SD) on day 1, 4, 9
Hirashima, 1997 <sup>202</sup>	9	12	Mean (SD) on day (0-4), (5-9), (10-14)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns			
ns	ns		
↓*			
maximum: ns, delta: ns			
ns/↑*	↑*	↑*	
↑	ns	ns	
ns	ns	ns	

**Figure S30: Interleukin-6 (CSF)**

Study	DCI	no DCI	Comparison
Roa, 2020 <sup>27</sup>	5	8	Mean (SEM) on day (0–1), (2–4), (5–9) and >10
Wu, 2016 <sup>97</sup>	27	30	Mean (SD) on day 2
Zanier, 2011 <sup>142</sup>	14	16	Levels during ICU-admission
Hendryk, 2004 <sup>189</sup>	8	7	Mean (SD) at day (0-3), (4-7), (8-15)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns	ns
↑			
ns			
↑	ns	ns	

**Figure S31: Interleukin-8 (blood)**

Study	DCI	no DCI	Comparison
Rasmussen, 2019 <sup>46</sup>	24	65	Mean (95% C.I.) on day 3 and 8. Ratio day 8/3
Ahn, 2019 <sup>47</sup>	14	46	Median (IQR) <24h, 24-48h, 3-5 and 6-8 days

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns			
ns	ns		

**Figure S32: Lactate (microdialysate)**

Study	DCI	no DCI	Comparison
Radolf, 2014 <sup>114</sup>	8	10	Median on day 1-11 Variance on day 8-11
Sarrafzadeh, '04 <sup>187</sup>	10	3	Mean (SD) of 3 day medians

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	↑/ns	ns	
↑*			
ns			

**Figure S33: Lactate/pyruvate ratio (microdialysate)**

Study	DCI	no DCI	Comparison
Radolf, 2014 <sup>114</sup>	8	10	Median on day 1-11
Sarrafzadeh, '04 <sup>187</sup>	10	3	Mean (SD) of 3 day medians

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns	
↑			

**Figure S34: Leukocytes (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	Pre-operative levels
Qi, 2019 <sup>51</sup>	53	199	Mean (SD) <24h
Wu, 2019 <sup>52</sup>	43	79	Mean (SD) <72h
			DCI in high vs low levels (threshold: 11.5*10 <sup>9</sup> /L)
Tao, 2017 <sup>69</sup>	47	200	Mean (SD) <24h
Chamling, 2017 <sup>76</sup>	16	65	Levels <24h
			Delta (maximum – baseline) value of day 0-14
Mijiti, 2016 <sup>94</sup>	99	244	% DCI in high & low levels at admission (threshold: 11.0x10 <sup>9</sup> )
Srinivasan, 2016 <sup>90</sup>	94	152	Mean (SD) at admission
Jeon, 2012 <sup>136</sup>	26	65	Mean (SD) levels on pre-operative and post-operative day 0 & (1-2)
Beefink, 2011 <sup>141</sup>	10	57	DCI in high & low mean levels mean levels of day 0-12 and 0-21 (threshold: median)
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24h
Kasius, 2010 <sup>154</sup>	11	40	Levels <72h
			Difference "before → after DCI onset" vs. paired samples in non-DCI
Oh, 2007 <sup>168</sup>	46	112	% with leukocytosis (WBC>11.000) in DCI vs. no DCI on admission & POD 1,3,5,7 % with severe leukocytosis (WBC>15.000) in DCI & no DCI on POD 7
Rothoerl, 2006 <sup>175</sup>	37	51	Mean on day 1-8
Nissen, 2001 <sup>193</sup>	13	23	Mean (SE) of means during hospitalization
Fuji, 1997 <sup>20</sup>	18	99	Mean (SD) on day 0, 3, 6, 14, 30

POD: post-operative day

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑			
ns*			
ns*			
↑*			
ns			
↑*			
delta: ns*			
↑DCI in high*			
↑			
↑/ns			
ns*			
↑			
ns*			
sign. bigger increase during DCI onset			
↑*/ns	ns		
	ns		
↑/ns	↑/ns		
ns			
ns	ns		ns

**Figure S35: L-selectin (blood)**

Study	DCI	no DCI	Comparison
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24h Difference "before → after DCI" vs. difference in paired samples in non-DCI
Nissen, 2001 <sup>193</sup>	13	23	Mean (SE) of means during hospitalization

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑			
sign. bigger increase during DCI onset			
↓			

**Figure S36: Lymphocytes (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	Pre-operative levels
Qi, 2019 <sup>51,†</sup>	53	199	Mean (SD) <24h
Wu, 2019 <sup>52</sup>	43	79	Mean (SD) <72h
			DCI in high vs low levels (threshold: 1.5*10 <sup>9</sup> /L)
Tao, 2017 <sup>69,†</sup>	47	200	Mean (SD) <24h
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24h

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns*			
ns*			
ns*			
↓*			
↑			

**Figure S37: Magnesium (blood)**

Study	DCI	no DCI	Comparison
Dorhout Mees, 2007 <sup>165</sup>	27	128	Mean (SD) of day 4 to 20/discharge Mean (SD) "latest before DCI"-levels % DCI per quartile of "latest before DCI"-levels in mmol/L: 1) ≥1.10≤1.28, 2) ≥1.28 ≤1.40, 3) >1.40 ≤1.62, 4) >1.62.
Collignon, 2004 <sup>188</sup>	55	72	Admission levels Mean, minimum and maximum during day (0-4), (4-14), (>14) % DCI per hypo- & normo- and hypermagnesemia (threshold: 1.7 and 2.1 mg/dL) during hospitalization
Van den Bergh, 2003 <sup>190</sup>	20	69	% DCI per hypo- & normomagnesemia <48h and between day 2-12 (threshold: 0.70 mmol/L)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
latest before DCI: ns			
↑DCI in Q1*			
ns			
mean, minimum and maximum: ns			
ns			
ns*	↑DCI in hypo*		

**Figure S38: Mannose-binding lectin (blood)**

Study	DCI	no DCI	Comparison
Matzen, 2020 <sup>34</sup>	13	20	Mean (SD) per day, day 0-9 after SAH
			Trend in time from day 0 – 9 after SAH
Zanier, 2014 <sup>113</sup>	14	25	Median (IQR) of early (day 1-3, median: 2) and late (day 4-14, median: 7) sample

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns	
ns*			
ns	ns		

**Figure S39: Mean platelet volume (blood)**

Study	DCI	no DCI	Comparison
Zhang, 2021 <sup>8</sup>	84	355	Median (IQR) at admission
Chen, 2020 <sup>30</sup>	58	139	Median (IQR) on day (1-3), (3-5), (5-7), (7-9)
Rzeplinski, 2020 <sup>23</sup>	8	29	Levels at admission

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
↑	↑*	↑	
ns			

**Figure S40: Mean platelet volume/platelet count ratio (blood)**

Study	DCI	no DCI	Comparison
Chen, 2020 <sup>30</sup>	58	139	Median (IQR) on day (1-3), (3-5), (5-7), (7-9)
Rzeplinski, 2020 <sup>23</sup>	8	29	Levels at admission
Ray, 2018 <sup>66</sup>	38	131	Admission levels
			Mean increase from admission to day 3

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑	ns*/↑	↑	
ns			
ns			
↑			

**Figure S41: Neutrophils (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	Pre-operative levels
Wu, 2019 <sup>52</sup>	43	79	Mean (SD) <72h
			DCI in high vs low levels (threshold: $7.5 \times 10^9/L$ )
Tao, 2017 <sup>69</sup>	47	200	Mean (SD) <24h
Frontera, 2017 <sup>75</sup>	?	?	Levels at 0-24, 24-48 & 48-72h
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24h

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
↑*			
ns*			
ns			
ns			
↑			

**Figure S42: Neutrophil/lymphocyte ratio (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	% DCI in high & low pre-operative ratio (threshold: 5.9)
Al-mufti, 2019 <sup>38</sup>	202	836	% DCI in high & low admission ratio (threshold: 5.9)
Wu, 2019 <sup>52</sup>	43	79	Mean (SD) <72h
Tao, 2017 <sup>69</sup>	47	200	Mean (SD) <24h

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑DCI in high			
↑DCI in high *			
↑*			
↑*			

**Figure S43: endothelial Nitric oxide synthase SNP rs2070744 T-786C**

Study	DCI	no DCI	Comparison
Hendrix, 2017 <sup>74</sup>	34	115	-
Khurana, 2004 <sup>185</sup>	no VS#: 7 as-VS: 10 S-VS: 11		% no/asymptomatic/symptomatic vasospasm per CC/CT/TT

Results			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
CT and C allele significantly associated with higher rates of symptomatic vasospasm†			

#: no VS: no vasospasm, as-VS: asymptomatic vasospasm, S-VS: symptomatic vasospasm (defined as DCI)

**Figure S44: Platelet count (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	Pre-operative levels
Chen, 2020 <sup>30</sup>	58	139	Median (IQR) on day (1-3), (3-5), (5-7), (7-9)
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Rzeplinski, 2020 <sup>23</sup>	8	29	Levels at admission
Tao, 2017 <sup>69</sup>	47	200	Mean (SD) <24h
Aggarwal, 2013 <sup>128</sup>	39	35	DCI in patients with vs. without absolute thrombocytopenia (<150.000) on day 1 & 7
			DCI in patients with vs. without relative thrombo-cytopenia (<80% of baseline) on day 5, 7, 9
Jeon, 2012 <sup>136</sup>	26	65	Mean (SD) levels pre-operative and post-operative day 0 & (1-2)
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24h
Kasius, 2010 <sup>154</sup>	11	36	Levels <72h
			Difference "before → after DCI onset" vs. paired samples in non-DCI
Schebesch, 2007 <sup>169</sup>	37	55	Mean on day 1-10
Nissen, 2001 <sup>193</sup>	13	23	Mean (SE) of means during hospitalization
Fuji, 1997 <sup>201</sup>	18	99	Mean (SD) on day 0, 3, 6, 14, 30

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns	ns*/↑	↑	
ns			
ns			
ns			
↑ DCI in thrombo-cytopenia day 1, 7			
	↑ DCI in thrombo-cytopenia day 5, 7, 9		
ns			
ns			
ns*			
ns difference during DCI onset			
ns	ns	ns	
↑			
ns	ns		ns

**Figure S45: Prothrombin time (blood)**

Study	DCI	no DCI	Comparison
Liu, 2020 <sup>28</sup>	224	663	Pre-operative levels
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Fuji, 1997 <sup>201</sup>	18	99	Mean (SD) on day 0, 3, 6, 14, 30

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			
ns	ns		ns

**Figure S46: P-selectin (blood)**

Study	DCI	no DCI	Comparison
Tang, 2015 <sup>107</sup>	12	46	Mean (SD) on day 1, 4, 9
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24h
			Difference "before → after DCI" vs. paired samples in non-DCI
Frijns, 2006 <sup>176,#</sup>	9	58	DCI in high & low levels <72h (threshold: 92.5)
Frijns, 2006 <sup>177,#</sup>	12	55	Difference "before → after DCI" vs. paired samples in non-DCI
Nissen, 2001 <sup>193</sup>	13	23	Mean (SE) of means during hospitalization

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	↑	ns	
↓			
↑ increase during DCI onset			
ns*			
↑ increase during DCI onset			
↑			

# two studies with overlapping patient cohorts, though different comparison between DCI and no DCI

**Figure S47: s-100B (blood)**

Study	DCI	no DCI	Comparison
Bergstrom, 2014 <sup>112</sup>	15	32	Mean (SD) on day (0-2), (3-5), (6-8), (9-11), (12-15)
Oertel, 2006 <sup>178</sup>	26	25	Mean (SD) of day 1, 2, 3

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns/↑	↑
↓*			

**Figure S48: Sodium (urine)**

Study	DCI	no DCI	Comparison
Nakagawa, 2013 <sup>121</sup>	13	90	% DCI in groups with/without excess sodium excretion on day 2, 3, 7, 14
Igarashi, 2007 <sup>167</sup>	12	55	Mean sodium excretion from 3 days before to 3 days after DCI
			Mean sodium <u>balance</u> (sodium intake – sodium excretion) from 3 days before to 3 days after DCI
Okuchi, 1996 <sup>204</sup>	4	13	Mean (SE) sodium <u>balance</u> on POD 1-10
			Mean negative peak and accumulative sodium <u>balance</u> of day POD 0-10

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		ns
DCI-3 to DCI-1: ns DCI to DCI+3: ↑			
DCI-3,-2, +0-2: ns DCI-1,DCI+3: ↓			
ns	↓/ns	ns	
↓			

POD = postoperative day

**Figure S49: Sodium (blood)**

Study	DCI	no DCI	Comparison
Cohen, 2021 <sup>1</sup>	142	214	Mean sodium during ICU admission
			Mean sodium variability during ICU admission
Liu, 2020 <sup>28</sup>	224	663	Pre-operative hyponatremia vs no hyponatremia
Wang, 2020 <sup>35</sup>	19	39	Levels <48h
Beseoglu, 2014 <sup>116</sup>	total: 274		Levels every 8h on day 0-5
Maimaitili, 2013 <sup>129,†</sup>	19	30	% DCI per normo- & hyponatremia on day 1-3 (threshold: 135)
Vrsajkov, 2012 <sup>137,†</sup>	39	43	% DCI per normo- & hyponatremia during hospitalization (threshold: 135)
Igarashi, 2007 <sup>167</sup>	12	55	Difference during 3 days before DCI → day of DCI
Qureshi, 2002 <sup>191</sup>	54	244	% DCI in patients with and without hypo- (<135) and hypernatremia (>145) between admission and day 9
Okuchi, 1996 <sup>204</sup>	4	13	Mean (range) of day 1-10
			Mean (SD) per day on day 1-10

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns*			
ns*			
ns			
ns			
ns			
↑ DCI in hyponatremia †			
↑ DCI in hyponatremia**			
ns difference around DCI onset			
hypo: ns, hyper: ns			
↓			
ns	↓/ns	ns/↓	

**Figure S50: Tau protein (CSF)**

Study	DCI	no DCI	Comparison
Halawa, 2018 <sup>63</sup>	7	12	Mean on day 4, 10
			Ratio day 10 : day 4
Zanier, 2013 <sup>130</sup>	11	24	Acute (<48h) and delayed (>48h) peak

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
	ns	ns	
	ratio: ↑		
ns			

**Figure S51: Thrombin-antithrombin III complex (blood)**

Study	DCI	no DCI	Comparison
Fuji, 1997 <sup>201</sup>	18	99	Mean (SD) on day 0, 3, 6, 14, 30
Hirashima, 1997 <sup>202</sup>	9	12	Levels at day (5-9)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	↑		ns
	↑		

**Figure S52: Thrombospondin-1 (blood)**

Study	DCI	no DCI	Comparison
Shen, 2015 <sup>101</sup>	33	85	Levels at admission
Nyquist, 2013 <sup>131</sup>	14	14	Levels on day (0-3)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
sign.			
ns			

**Figure S53: Tumor necrosis factor a (blood)**

Study	DCI	no DCI	Comparison
Rasmussen, 2019 <sup>46</sup>	24	65	Mean (95% C.I.) on day 3 and 8.
			Ratio day 8/3
Ahn, 2019 <sup>47</sup>	14	46	Median (IQR) <24, 24-48 hours, 3-5 and 6-8 days
Beefink, 2011 <sup>141</sup>	10	57	DCI in high & low mean levels mean levels of day 0-12 and 0-21 (threshold: median)
Hirashima, 1997 <sup>202</sup>	9	12	Levels at day (0-4), (5-9), (10-14)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns			
ns	ns		
ns			
ns	ns	ns	

**Figure S54: Tumor necrosis factor a (CSF)**

Study	DCI	no DCI	Comparison
Roa, 2020 <sup>27</sup>	5	8	Mean (SEM) on day (0-1), (2-4), (5-9) and >10
Wu, 2016 <sup>97</sup>	27	30	Mean (SD) on day 2
Zanier, 2011 <sup>142</sup>	14	16	Levels during ICU-admission

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns	ns
↑			
ns			

**Figure S55: Transferrin (CSF)**

Study	DCI	no DCI	Comparison
Gomes, 2014 <sup>115</sup>	7	5	Mean (SE) on day 1, 3, 5
Takenaka, 2000 <sup>199</sup>	7	13	Mean (SD)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
↑ (unknown when measured)			



**Figure S56: Vascular cell adhesion protein 1 (blood)**

Study	DCI	no DCI	Comparison
Rasmussen, 2019 <sup>46</sup>	24	65	Mean (95% C.I.) on day 3 and 8. Ratio day 8/3
Wang, 2011 <sup>151</sup>	7	14	Median (IQR) <24 hours Difference "before → after DCI" vs. paired samples in non-DCI
Rothoerl, 2006 <sup>174</sup>	5	10	Mean level on day 1-9 Maximum level during day 1-9
Nissen, 2001 <sup>193</sup>	13	23	Mean (SE) of means during hospitalization

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns			
↑			
ns difference during DCI onset			
ns	ns	ns	
ns			
ns			

**Figure S57: von Willebrand Factor (blood)**

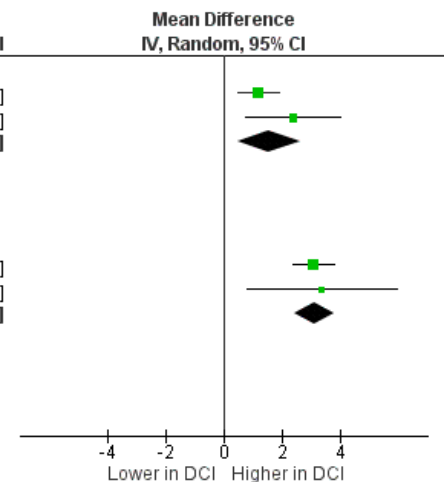
Study	DCI	no DCI	Comparison
Li, 2017 <sup>87,†</sup>	31	52	Mean (SD) on day 1, 4, 10 (U/mL)
Tang, 2015 <sup>107, †</sup>	12	58	Mean (SD) on day 1, 4, 9 (U/mL)
Frijns, 2006 <sup>176</sup>	12	55	DCI in high & low levels <72h (threshold 93.0)
Hirashima, 1997 <sup>202</sup>	9	12	Mean (SD) on day (5-9)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑	↑	ns	
↑	↑	ns	
ns*			
	↑		

† meta-analysis possible of 2 of 2 studies with measurements on day 1 and 4, see below.

**Figure S57.1:**

Study or Subgroup	DCI			no DCI			Weight	Mean Difference IV, Random, 95% CI
	Mean	SD	Total	Mean	SD	Total		
<b>10.1.1 Day 1</b>								
Li 2017	3.4	1.7	31	2.2	1.5	52	31.9%	1.20 [0.48, 1.92]
Tang 2015	4.8	2.7	12	2.4	2.4	58	21.9%	2.40 [0.75, 4.05]
<b>Subtotal (95% CI)</b>			<b>43</b>			<b>110</b>	<b>53.8%</b>	<b>1.56 [0.48, 2.64]</b>
Heterogeneity: Tau <sup>2</sup> = 0.30; Chi <sup>2</sup> = 1.71, df = 1 (P = 0.19); I <sup>2</sup> = 41%								
Test for overall effect: Z = 2.84 (P = 0.005)								
<b>10.1.2 Day 4</b>								
Li 2017	5.3	1.5	31	2.2	1.7	52	32.1%	3.10 [2.40, 3.80]
Tang 2015	5.1	4.5	12	1.7	1.1	58	14.1%	3.40 [0.84, 5.96]
<b>Subtotal (95% CI)</b>			<b>43</b>			<b>110</b>	<b>46.2%</b>	<b>3.12 [2.44, 3.80]</b>
Heterogeneity: Tau <sup>2</sup> = 0.00; Chi <sup>2</sup> = 0.05, df = 1 (P = 0.82); I <sup>2</sup> = 0%								
Test for overall effect: Z = 9.04 (P < 0.00001)								



**Figure S58: von Willebrand Factor propeptide (blood)**

Study	DCI	no DCI	Comparison
Vergouwen, 2009 <sup>158</sup>	11	20	Trend in time during day 2, 4, 7, 10, 14, 17
Frijns, 2006 <sup>176</sup>	12	55	DCI in high & low levels <72h (threshold 7.70)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑ increase first few days			
ns*			

## Biomarkers of radiological DCI which have been studied by more than one study

**Figure S59: Big endothelin-1 (blood)**

Study	DCI	no DCI	Comparison
Juvela, 2002 <sup>192</sup>	?	?	Mean (SD) at admission, after surgery and at discharge
Gruber, 2000 <sup>198</sup>	17	27	Mean (SD) of day 0-14

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns		ns	
ns			

**Figure S60: Brain natriuretic peptide (blood)**

Study	DCI	no DCI	Comparison
Papanikolaou 2012 <sup>133</sup>	12	24	Mean (SE) before repair surgery
Taub, 2011 <sup>148</sup>	46	73	Quartile analysis of levels after admission, mean day 5.5 ± 3 (top quartile vs other)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑*			
↑*			

**Figure S61: Carbon dioxide partial pressure (blood)**

Study	DCI	no DCI	Comparison
Darkwah, 2021 <sup>6</sup>	153	480	Mean daily highest and lowest values of day 0-13
			% DCI in optimal pCO <sub>2</sub> (30-38) vs. non-optimal pCO <sub>2</sub> (<30/>38) during day 0-13
Nakae, 2011 <sup>147</sup>	28	114	Mean (SD), unknown when measured

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
lowest: ns, highest: ns			
↑ DCI in non-optimal*			
ns (unknown when measured)			

**Figure S62: total Cholesterol (blood)**

Study	DCI	no DCI	Comparison
Li, 2020 <sup>32</sup>	37	38	% DCI in high & low levels at admission (threshold: 160)
Yang, 2012 <sup>134</sup>	10	10	Mean (SD) <48h

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns*			
ns			

**Figure S63: C-reactive protein (blood)**

Study	DCI	no DCI	Comparison
Guresir, 2020 <sup>37</sup>	22	209	% DCI in high & low levels at admission (threshold: 5)
Hurth, 2020 <sup>31</sup>	27	111	Median (IQR) at admission, after treatment, on day 4, 9, 14 after SAH and at discharge from ICU
Matano, 2018 <sup>65</sup>	41	292	Mean (SD) at admission
Hostettler, 2018 <sup>55</sup>	186	168	DCI in high & low CRP on day 1 (threshold: 23)
Juvela, 2012 <sup>132</sup>	no ischemia: 69 early ischemia: 59 DCI: 50		Mean(SD) no/early/late ischemic lesion at admission, after surgery (mean day 2), at discharge (mean day 10)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑DCI in high			
ns	ns	↑	↑
ns			
↑DCI in high			
ns		ns	

**Figure S64: D-dimer (blood)**

Study	DCI	no DCI	Comparison
Hurth, 2020 <sup>31</sup>	27	111	Median (IQR) at admission, after treatment, day 4, 9 and 14 after SAH and at discharge from ICU
Fukuda, 2017 <sup>86</sup>	12	175	Median (IQR) at admission
Juvela, 2006 <sup>173</sup>	no ischemia: 69 other ischemia: 59 DCI: 37		Median (IQR) of DCI / other ischemia /no ischemia at surgery & at discharge

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns	ns	↑
ns*			
ns		ns	

**Figure S65: soluble Endoglin (blood)**

Study	DCI	no DCI	Comparison
Griessenauer, 2017 <sup>82</sup>	9	18	Mean on day 1, 6
Dietman, 2011 <sup>143</sup>	7	13	Mean(SE) on day 1-7, 9, 11, 13, 15

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
↓	↓	↓	↓

**Figure S66: Endothelin-1 (blood)**

Study	DCI	no DCI	Comparison
Kastner, 2005 <sup>180</sup>	7	13	Medians day 1-8
Juvela, 2002 <sup>192</sup>	9	37	Mean (SD) at admission, after surgery and at discharge
Juvela, 2000 <sup>197</sup>	20	35	Mean (SE) day (1-5), (6-14), (15-28)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns	ns		
ns		ns	
ns	ns		ns

**Figure S67: Glucose (blood)**

Study	DCI	no DCI	Comparison
Matano, 2018 <sup>65,†</sup>	41	292	Mean (SD) at admission
Beseoglu, 2017 <sup>77</sup>	14	73	Relative risk for DCI in high vs low glucose <72h (threshold: 7.8 mmol/L)
Mijiti, 2016 <sup>94</sup>	87	256	% DCI per dichotomized glucose at admission (threshold: 7.0 mmol/L)
van Donkelaar, 2016 <sup>95</sup>	84	201	Median (IQR) maximum level <24h
De Rooij, 2013 <sup>118</sup>	110	261	Hyperglycemia at admission
Yang, 2012 <sup>134,†</sup>	10	10	Mean (SD) <48h
Nakae, 2011 <sup>147</sup>	28	114	Mean (SD), timing of measurement unknown
Juvela, 2005 <sup>184</sup>	No ischemia: 55 Other ischemia: 54 DCI: 46		Mean (SD) at admission and after surgery (mean day 1.9)

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑			
ns			
ns*			
↑*			
ns*			
ns			
ns (timing unknown)			
ns*/ns			

**Figure S68: Hemoglobin (blood)**

Study	DCI	no DCI	Comparison
Cai, 2018 <sup>61</sup>	17	43	% DCI in high vs. low "lowest level" (threshold: 10g/dL), timing of measurement unknown
Nakae, 2011 <sup>147</sup>	28	114	Mean (SD), timing of measurement unknown
Naidech, 2010 <sup>152</sup>	20	24	% DCI in goal level ≥11.5 and ≥10, day 0-13

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑ DCI in low (timing unknown)			
ns (timing unknown)			
ns			

**Figure S69: Lactate (blood)**

Study	DCI	no DCI	Comparison
Van Donkelaar, 2016 <sup>95</sup>	84	201	Median (IQR) maximum level <24h
Nakae, 2011 <sup>147</sup>	28	114	Mean (SD), timing of measurement unknown

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns*			
ns (timing unknown)			

**Figure S70: Lactate/pyruvate ratio (microdialysate)**

Study	DCI	no DCI	Comparison
Kofler, 2020 <sup>21</sup>	16	35	Levels on day (0-3), (4-7), >7
			Occurrence of metabolic distress (levels >40) on day (0-3), (4-7), >7
Torne, 2020 <sup>26</sup>	5	16	Hours of lactate pyruvate ratio >40, day 0-13

Results DCI compared to no DCI			
Day 0-3	Day 4-8	Day 9-13	Day ≥14
		↑	
		↑	
		↑	

**Figure S71: Leukocytes (blood)**

Study	DCI	no DCI	Comparison
Guresir, 2020 <sup>37</sup>	22	209	% DCI in high vs. low levels at admission (threshold: 12.1)
Hurth, 2020 <sup>31</sup>	27	111	Median (IQR) at admission, after treatment, day 4, 9 and 14 after SAH and at discharge from ICU
Matano, 2018 <sup>65,†</sup>	41	292	Mean (SD) at admission
Mijiti 2016 <sup>94</sup>	87	256	% DCI per dichotomized levels at admission (threshold: $11.0 \times 10^9$ )
Srinivasan 2016 <sup>90+</sup>	62	184	Mean (SD) at admission
Yang 2012 <sup>134+</sup>	10	10	Mean (SD) <48h
Nakae, 2011 <sup>147</sup>	28	114	Mean (SD), timing of measurement unknown

**Results DCI compared to no DCI**

Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns	ns	ns	ns
ns			
↑*			
ns*			
ns			
ns (timing unknown)			

**Figure S72: Neutrophil/lymphocyte ratio (blood)**

Study	DCI	no DCI	Comparison
Giede-Jeppe, 2019 <sup>40</sup>	112	217	% DCI in high vs low levels at admission (threshold: 7.05)
Al-Mufti, 2019 <sup>38</sup>	170	862	% DCI in high vs low levels at admission (threshold: 5.9)

**Results****Results DCI compared to no DCI**

Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns*			
↑ DCI in high			

**Figure S73: Platelet count (blood)**

Study	DCI	no DCI	Comparison
Yang, 2012 <sup>134</sup>	10	10	Mean(SD) <48h
Hirashima, 1994 <sup>206</sup>	9	23	Minimum during at least 11 days

**Results DCI compared to no DCI**

Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
minimum: ↓			

**Figure S74: Potassium (blood)**

Study	DCI	no DCI	Comparison
Matano, 2018 <sup>65</sup>	41	292	Mean (SD) at admission
Nakae, 2011 <sup>147</sup>	28	114	Mean (SD), timing of measurement unknown

**Results DCI compared to no DCI**

Day 0-3	Day 4-8	Day 9-13	Day ≥14
↓			
ns (timing unknown)			

**Figure S75: Procalcitonin (blood)**

Study	DCI	no DCI	Comparison
Guresir, 2020 <sup>37</sup>	22	209	% DCI in high & low levels at admission (threshold: 0.5)
Hurth, 2020 <sup>31</sup>	27	111	Median (IQR) at admission, after treatment, day 4, 9, 14 after SAH and at discharge from ICU

**Results DCI compared to no DCI**

Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns	ns	ns	ns

**Figure S76: Sodium (blood)**

Study	DCI	no DCI	Comparison
Hendrix, 2017 <sup>71</sup>	31	115	DCI in patients with and without hyponatremia <72h of admission (threshold: unknown)
Beseoglu, 2014 <sup>116</sup>	total: 274		Correlation of DCI with hypernatremia (>145) at 56h after ictus
Zheng, 2011 <sup>145</sup>	94	289	% DCI in patients with and without hyponatremia during day 0-9 (threshold: 135)
Nakae, 2011 <sup>147</sup>	28	114	Mean (SD), timing of measurement unknown

**Results DCI compared to no DCI**

Day 0-3	Day 4-8	Day 9-13	Day ≥14
↑ DCI in hypo*			
ns			
↑ DCI in hypo			
ns (timing unknown)			

**Figure S77: Triglycerides (blood)**

Study	DCI	no DCI	Comparison
Li, 2020 <sup>32</sup>	37	38	DCI in low vs. high levels at admission (threshold: 100 mg/dL)
Mjiti, 2016 <sup>94</sup>	87	256	% DCI in high vs. low levels at admission (threshold: 2.26 mmol/L)

**Results DCI compared to no DCI**

Day 0-3	Day 4-8	Day 9-13	Day ≥14
ns			
ns			

## References

1. Cohen J, Delaney A, Anstey J, et al. Dysnatremia and 6-Month Functional Outcomes in Critically Ill Patients With Aneurysmal Subarachnoid Hemorrhage: A Prospective Cohort Study. *Crit care explor.* 2021;3(6):e0445.
2. Albanna W, Weiss M, Veldeman M, et al. Urea-Creatinine Ratio (UCR) After Aneurysmal Subarachnoid Hemorrhage: Association of Protein Catabolism with Complication Rate and Outcome. *World Neurosurg.* 2021;18:18.
3. Helbok R, Rass V, Kofler M, et al. Intracerebral Iron Accumulation may be Associated with Secondary Brain Injury in Patients with Poor Grade Subarachnoid Hemorrhage. *Neurocrit Care.* 2021;9:09.
4. Veldeman M, Dogan R, Weiss M, et al. Levels of bioactive adrenomedullin in plasma and cerebrospinal fluid in relation to delayed cerebral ischemia in patients after aneurysmal subarachnoid hemorrhage: A prospective observational study. *J Neurol Sci.* 2021;427:117533.
5. Krenzlin H, Wesp D, Schmitt J, et al. Decreased Superoxide Dismutase Concentrations (SOD) in Plasma and CSF and Increased Circulating Total Antioxidant Capacity (TAC) Are Associated with Unfavorable Neurological Outcome after Aneurysmal Subarachnoid Hemorrhage. *J Clin Med.* 2021;10(6):12.
6. Darkwah Oppong M, Wrede KH, Muller D, et al. PaCO<sub>2</sub>-management in the neuro-critical care of patients with subarachnoid hemorrhage. *Sci Rep.* 2021;11(1):19191.
7. Park CW, Yi HJ, Lee DH, Sung JH. Association between HbA1C (Glycated Hemoglobin) and Clinical Outcomes in Patients with Subarachnoid Hemorrhage After Neuro-intervention. *Curr Neurovasc Res.* 2021;18(1):93-101.
8. Zhang X, Liu Y, Zhang S, Wang C, Zou C, Li A. Neutrophil-to-Albumin Ratio as a Biomarker of Delayed Cerebral Ischemia After Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg.* 2021;147:e453-e8.
9. Veldeman M, Weiss M, Simon TP, Hoellig A, Clusmann H, Albanna W. Body mass index and leptin levels in serum and cerebrospinal fluid in relation to delayed cerebral ischemia and outcome after aneurysmal subarachnoid hemorrhage. *Neurosurg Rev.* 2021;17:17.
10. Neumaier F, Stoppe C, Veldeman M, et al. Circulatory dipeptidyl peptidase 3 (cDPP3) is a potential biomarker for early detection of secondary brain injury after aneurysmal subarachnoid hemorrhage. *J Neurol Sci.* 2021;422:117333.
11. Heinsberg LW, Arockiaraj AI, Crago EA, et al. Genetic Variability and Trajectories of DNA Methylation May Support a Role for HAMP in Patient Outcomes After Aneurysmal Subarachnoid Hemorrhage. *Neurocrit Care.* 2020;32(2):550-63.
12. Heinsberg LW, Alex, er SA, et al. Genetic Variability in the Iron Homeostasis Pathway and Patient Outcomes After Aneurysmal Subarachnoid Hemorrhage. *Neurocrit Care.* 3:03.
13. Luo YG, Han B, Sun TW, Liu X, Liu J, Zhang J. The association between serum adipocyte fatty acid-binding protein and 3-month disability outcome after aneurysmal subarachnoid hemorrhage. *J Neuroinflammation.* 2020;17(1):66.
14. Veldeman M, Lepore D, Hollig A, et al. Procalcitonin in the context of delayed cerebral ischemia after aneurysmal subarachnoid hemorrhage. *J Neurosurg.* 2020:1-9.
15. Chaudhry SR, Kinfe TM, Lamprecht A, et al. Elevated level of cerebrospinal fluid and systemic chemokine CCL5 is a predictive biomarker of clinical outcome after aneurysmal subarachnoid hemorrhage (aSAH). *Cytokine.* 2020;133:155142.
16. Chaudhry SR, Kahlert UD, Kinfe TM, et al. Elevated Systemic IL-10 Levels Indicate Immunodepression Leading to Nosocomial Infections after Aneurysmal Subarachnoid Hemorrhage (SAH) in Patients. *Int J Mol Sci.* 2020;21(5):25.
17. Kim BJ, Kim Y, Youn DH, et al. Genome-wide blood DNA methylation analysis in patients with delayed cerebral ischemia after subarachnoid hemorrhage. *Sci Rep.* 2020;10(1):11419.

18. Bache S, Rasmussen R, Wolcott Z, et al. Elevated miR-9 in Cerebrospinal Fluid Is Associated with Poor Functional Outcome After Subarachnoid Hemorrhage. *Transl Stroke Res.* 2020;4:04.
19. Wang HB, Wu QJ, Zhao SJ, et al. Early High Cerebrospinal Fluid Glutamate: A Potential Predictor for Delayed Cerebral Ischemia after Aneurysmal Subarachnoid Hemorrhage. *ACS Omega.* 2020;5(25):15385-9.
20. Park JJ, Kim C, Jeon JP. Monitoring of Delayed Cerebral Ischemia in Patients with Subarachnoid Hemorrhage via Near-Infrared Spectroscopy. *J Clin Med.* 2020;9(5):24.
21. Kofler M, Gaasch M, Rass V, et al. The Importance of Probe Location for the Interpretation of Cerebral Microdialysis Data in Subarachnoid Hemorrhage Patients. *Neurocrit Care.* 2020;32(1):135-44.
22. Wisniewski K, Popeda M, Tomasiak B, et al. The Role of Urine F2-ISOPROSTANE CONcentration in Delayed Cerebral Ischemia after Aneurysmal Subarachnoid Haemorrhage-A Poor Prognostic Factor. *Diagnostics.* 2020;11(1):22.
23. Rzeplinski R, Kostyra K, Skadorwa T, Slugocki M, Kostkiewicz B. Acute platelet response to aneurysmal subarachnoid hemorrhage depends on severity and distribution of bleeding: an observational cohort study. *Neurosurg Rev.* 2020;25:25.
24. Youn DH, Kim BJ, Kim Y, Jeon JP. Extracellular Mitochondrial Dysfunction in Cerebrospinal Fluid of Patients with Delayed Cerebral Ischemia after Aneurysmal Subarachnoid Hemorrhage. *Neurocrit Care.* 2:02.
25. Manner A, Thomas D, Wagner M, et al. Sphingosine 1-phosphate levels in cerebrospinal fluid after subarachnoid hemorrhage. *Neurol Res Pract.* 2020;2:49.
26. Torne R, Culebras D, Sanchez-Etayo G, et al. Double hemispheric Microdialysis study in poor-grade SAH patients. *Sci Rep.* 2020;10(1):7466.
27. Roa JA, Sarkar D, Zanaty M, et al. Preliminary results in the analysis of the immune response after aneurysmal subarachnoid hemorrhage. *Sci Rep.* 10(1):11809.
28. Liu H, Xu Q, Li A. Nomogram for predicting delayed cerebral ischemia after aneurysmal subarachnoid hemorrhage in the Chinese population. *J Stroke Cerebrovasc Dis.* 2020;29(9).
29. Arleth T, Olsen MH, Orre M, et al. Hypozincaemia is associated with severity of aneurysmal subarachnoid haemorrhage: a retrospective cohort study. *Acta Neurochir (Wien).* 162(6):1417-24.
30. Chen L, Zhang Q. Dynamic Change in Mean Platelet Volume and Delayed Cerebral Ischemia After Aneurysmal Subarachnoid Hemorrhage. *Front Neurol.* 2020;11:571735.
31. Hurth H, Birkenhauer U, Steiner J, Schlak D, Hennesdorf F, Ebner FH. Delayed Cerebral Ischemia in Patients with Aneurysmal Subarachnoid Hemorrhage - Serum D-dimer and C-reactive Protein as Early Markers. *J Stroke Cerebrovasc Dis.* 2020;29(3):104558.
32. Li B, McIntyre M, hi C, et al. Low total cholesterol and high density lipoprotein are independent predictors of poor outcomes following aneurysmal subarachnoid hemorrhage: A preliminary report. *Clin Neurol Neurosurg.* 2020;197:106062.
33. Xie B, Lin Y, Wu X, Yu L, Zheng S, Kang D. Reduced Admission Serum Fibrinogen Levels Predict 6-Month Mortality of Poor-Grade Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg.* 136:e24-e32.
34. Matzen JS, Krogh CL, Forman JL, Garred P, Moller K, Bache S. Lectin complement pathway initiators after subarachnoid hemorrhage - an observational study. *J Neuroinflammation.* 2020;17(1):338.
35. Wang C, Kou Y, Han Y, Li X. Early Serum Calprotectin (S100A8/A9) Predicts Delayed Cerebral Ischemia and Outcomes after Aneurysmal Subarachnoid Hemorrhage. *J Stroke Cerebrovasc Dis.* 2020;29(5):104770.
36. Jayamanoharan S, Mangum JE, Stylli S, Ziogas J, Adamides AA. Association between elevated cerebrospinal fluid D-dimer levels and delayed cerebral ischaemia after aneurysmal subarachnoid haemorrhage. *J Clin Neurosci.* 76:177-82.
37. Guresir E, Coch C, Fimmers R, et al. Initial inflammatory response is an independent predictor of unfavorable outcome in patients with good-grade aneurysmal subarachnoid hemorrhage. *J Crit Care.* 60:45-9.

38. Al-Mufti F, Amuluru K, Damodara N, et al. Admission neutrophil-lymphocyte ratio predicts delayed cerebral ischemia following aneurysmal subarachnoid hemorrhage. *J Neurointerv Surg.* 2019;11(11):1135-40.
39. Zhang YB, Zheng SF, Shang-Guan HC, Kang DZ, Chen GR, Yao PS. Lower Iron Levels Predict Acute Hydrocephalus Following Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg.* 2019;126:e907-e13.
40. Giede-Jeppe A, Reichl J, Sprugel MI, et al. Neutrophil-to-lymphocyte ratio as an independent predictor for unfavorable functional outcome in aneurysmal subarachnoid hemorrhage. *J Neurosurg.* 2019;132(2):400-7.
41. Bevers MB, Wolcott Z, Bache S, et al. Soluble ST2 links inflammation to outcome after subarachnoid hemorrhage. *Annals of Neurology.* 86(3):384-94.
42. Zhu Y, Jiang H, Li Y, et al. Serum Alkaline Phosphatase Level is Associated with Angiographic Vasospasm, Delayed Cerebral Ischemia-Caused Clinical Deterioration, and Functional Outcome After Aneurysmal Subarachnoid Hemorrhage. *Neurocrit Care.* 2019;31(3):466-75.
43. Kanamaru H, Kawakita F, Nakano F, et al. Plasma Periostin and Delayed Cerebral Ischemia After Aneurysmal Subarachnoid Hemorrhage. *Neurotherapeutics.* 16(2):480-90.
44. Chen JL, Yuan DH, Yang SJ, Gu C, Zhou HS, Shao GF. Serum netrin-1 serves as a prognostic biomarker of aneurysmal subarachnoid hemorrhage. *Clin Chim Acta.* 2019;495:294-300.
45. Tanioka S, Ishida F, Nakano F, et al. Machine Learning Analysis of Matricellular Proteins and Clinical Variables for Early Prediction of Delayed Cerebral Ischemia After Aneurysmal Subarachnoid Hemorrhage. *Mol Neurobiol.* 56(10):7128-35.
46. Rasmussen R, Bache S, Stavngaard T, Moller K. Plasma Levels of IL-6, IL-8, IL-10, ICAM-1, VCAM-1, IFN $\gamma$ , and TNF $\alpha$  are not Associated with Delayed Cerebral Ischemia, Cerebral Vasospasm, or Clinical Outcome in Patients with Subarachnoid Hemorrhage. *World Neurosurg.* 2019;128:e1131-e6.
47. Ahn SH, Savarraj JPJ, Parsha K, et al. Inflammation in delayed ischemia and functional outcomes after subarachnoid hemorrhage. *J Neuroinflammation.* 16(1):213.
48. Hviid CVB, Lauridsen SV, Gyldenholm T, Sunde N, Parkner T, Hvas A-M. Plasma neurofilament light chain is associated with poor functional outcome and mortality rate after spontaneous subarachnoid hemorrhage. *Transl Stroke Res.* 2019:1-7.
49. Kim BJ, Kim Y, Hong EP, et al. Correlation Between Altered DNA Methylation of Intergenic Regions of ITPR3 and Development of Delayed Cerebral Ischemia in Patients with Subarachnoid Hemorrhage. *World Neurosurg.* 2019;130:e449-e56.
50. Kwan K, Arapi O, Wagner KE, et al. Cerebrospinal fluid macrophage migration inhibitory factor: a potential predictor of cerebral vasospasm and clinical outcome after aneurysmal subarachnoid hemorrhage. *J Neurosurg.* 1-6.
51. Qi H, Yang X, Hao C, et al. Clinical Value of Controlling Nutritional Status Score in Patients with Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg.* 2019;126:e1352-e8.
52. Wu Y, He Q, Wei Y, et al. The association of neutrophil-to-lymphocyte ratio and delayed cerebral ischemia in patients with aneurysmal subarachnoid hemorrhage: possible involvement of cerebral blood perfusion. *Neuropsychiatr Dis Treat.* 2019;15:1001.
53. Lin M, Griessenauer CJ, Starke RM, et al. Haplotype analysis of SERPINE1 gene: Risk for aneurysmal subarachnoid hemorrhage and clinical outcomes. *Mol Genet Genomic Med.* 2019;7(8):e737.
54. Aida Y, Kamide T, Ishii H, et al. Soluble receptor for advanced glycation end products as a biomarker of symptomatic vasospasm in subarachnoid hemorrhage. *J Neurosurg.* 2019;134(1):122-30.
55. Hostettler IC, Muroi C, Richter JK, et al. Decision tree analysis in subarachnoid hemorrhage: prediction of outcome parameters during the course of aneurysmal subarachnoid hemorrhage using decision tree analysis. *J Neurosurg.* 2018:1-12.

56. Ayling OGS, Ibrahim GM, Alotaibi NM, Gooderham PA, Macdonald RL. Anemia After Aneurysmal Subarachnoid Hemorrhage Is Associated With Poor Outcome and Death. *Stroke*. 2018;49(8):1859-65.
57. Griessenauer CJ, Starke RM, Foreman PM, et al. Associations between endothelin polymorphisms and aneurysmal subarachnoid hemorrhage, clinical vasospasm, delayed cerebral ischemia, and functional outcome. *J Neurosurg*. 2018;128(5):1311-7.
58. Hendrix P, Foreman PM, Harrigan MR, et al. Association of cystathionine beta-synthase polymorphisms and aneurysmal subarachnoid hemorrhage. *J Neurosurg*. 2018;128(6):1771-7.
59. Nakatsuka Y, Shiba M, Nishikawa H, et al. Acute-Phase Plasma Osteopontin as an Independent Predictor for Poor Outcome After Aneurysmal Subarachnoid Hemorrhage. *Mol Neurobiol*. 2018;55(8):6841-9.
60. Yang DB, Dong XQ, Du Q, et al. Clinical relevance of cleaved RAGE plasma levels as a biomarker of disease severity and functional outcome in aneurysmal subarachnoid hemorrhage. *Clin Chim Acta*. 2018;486:335-40.
61. Cai J, Fang W, Chen F, et al. Cerebral perfusion pressure threshold to prevent delayed cerebral ischemia after aneurysmal subarachnoid hemorrhage. *J Clin Neurosci*. 2018;54:29-32.
62. Thomas AJ, Ogilvy CS, Griessenauer CJ, Hanafy KA. Macrophage CD163 expression in cerebrospinal fluid: association with subarachnoid hemorrhage outcome. *J Neurosurg*. 2018:1-7.
63. Halawa I, Vlachogiannis P, Amandusson Å, et al. Seizures, CSF neurofilament light and tau in patients with subarachnoid haemorrhage. *Acta Neurol Scand*. 2018;137(2):199-203.
64. Kim BJ, Kim Y, Kim SE, Jeon JP. Study of Correlation Between Hp alpha1 Expression of Haptoglobin 2-1 and Clinical Course in Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg*. 2018;117:e221-e7.
65. Matano F, Fujiki Y, Mizunari T, et al. Serum Glucose and Potassium Ratio as Risk Factors for Cerebral Vasospasm after Aneurysmal Subarachnoid Hemorrhage. *J Stroke Cerebrovasc Dis*. 2018;28(7):1951-7.
66. Ray B, Tinsley L, Ford L, Thompson DM, Sidorov EV, Bohnstedt BN. Trends of Platelet Volume Index Predicts Delayed Cerebral Ischemia After Subarachnoid Hemorrhage. *World Neurosurg*. 2018;111:e624-e31.
67. Suzuki H, Nakatsuka Y, Yasuda R, et al. Dose-Dependent Inhibitory Effects of Cilostazol on Delayed Cerebral Infarction After Aneurysmal Subarachnoid Hemorrhage. *Transl Stroke Res*. 2018:23.
68. Chaudhry SR, Guresir A, Stoffel-Wagner B, et al. Systemic High-Mobility Group Box-1: A Novel Predictive Biomarker for Cerebral Vasospasm in Aneurysmal Subarachnoid Hemorrhage. *Crit Care Med*. 2018;46(11):e1023-e8.
69. Tao C, Wang J, Hu X, Ma J, Li H, You C. Clinical Value of Neutrophil to Lymphocyte and Platelet to Lymphocyte Ratio After Aneurysmal Subarachnoid Hemorrhage. *Neurocrit Care*. 2017;26(3):393-401.
70. Griessenauer CJ, Tubbs RS, Foreman PM, et al. Associations of renin-angiotensin system genetic polymorphisms and clinical course after aneurysmal subarachnoid hemorrhage. *J Neurosurg*. 2017;126(5):1585-97.
71. Hendrix P, Foreman PM, Harrigan MR, et al. Association of Plasminogen Activator Inhibitor 1 (SERPINE1) Polymorphisms and Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg*. 2017;105:672-7.
72. Hendrix P, Foreman PM, Harrigan MR, et al. Impact of High-Mobility Group Box 1 Polymorphism on Delayed Cerebral Ischemia After Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg*. 2017;101:325-30.
73. Hendrix P, Foreman PM, Harrigan MR, et al. Ryanodine Receptor 1 Polymorphism Is Not Associated with Aneurysmal Subarachnoid Hemorrhage or its Clinical Sequelae. *World Neurosurg*. 2017;100:190-4.
74. Hendrix P, Foreman PM, Harrigan MR, et al. Endothelial Nitric Oxide Synthase Polymorphism Is Associated with Delayed Cerebral Ischemia Following Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg*. 2017;101:514-9.



75. Frontera JA, Provencio JJ, Sehba FA, et al. The Role of Platelet Activation and Inflammation in Early Brain Injury Following Subarachnoid Hemorrhage. *Neurocrit Care*. 2017;26(1):48-57.
76. Chamling B, Gross S, Stoffel-Wagner B, et al. Early Diagnosis of Delayed Cerebral Ischemia: Possible Relevance for Inflammatory Biomarkers in Routine Clinical Practice? *World Neurosurg*. 2017;104:2-5.
77. Beseoglu K, Steiger HJ. Elevated glycated hemoglobin level and hyperglycemia after aneurysmal subarachnoid hemorrhage. *Clin Neurol Neurosurg*. 2017;163:128-32.
78. Hollig A, Stoffel-Wagner B, Clusmann H, Veldeman M, Schubert GA, Coburn M. Time courses of inflammatory markers after aneurysmal subarachnoid hemorrhage and their possible relevance for future studies. *Front Neurol*. 2017;8 (DEC) (no pagination)(694):694.
79. Chaudhry SR, Guresir E, Vatter H, et al. Aneurysmal subarachnoid hemorrhage lead to systemic upregulation of IL-23/IL-17 inflammatory axis. *Cytokine*. 2017;97:96-103.
80. Chaudhry SR, Stoffel-Wagner B, Kiefe TM, et al. Elevated systemic IL-6 levels in patients with aneurysmal subarachnoid hemorrhage is an unspecific marker for post-SAH complications. *Int J Mol Sci*. 2017;18 (12) (no pagination)(2580):01.
81. Bache S, Rasmussen R, Rossing M, Laigaard FP, Nielsen FC, Moller K. MicroRNA Changes in Cerebrospinal Fluid after Subarachnoid Hemorrhage. *Stroke*. 2017;48(9):2391-8.
82. Griessenauer CJ, Chua MH, Hanafy KA, et al. Soluble Fms-Like Tyrosine Kinase 1 (sFlt-1) and Risk of Cerebral Vasospasm After Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg*. 2017;108:84-9.
83. Wisniewski K, Bienkowski M, Tomasik B, et al. Urinary  $\text{F}_{2\text{-isoprostane}}$  Concentration as a Poor Prognostic Factor After Subarachnoid Hemorrhage. *World Neurosurg*. 2017;107:185-93.
84. Spitzer D, Spitzer NJ, Deininger M, et al. Activation of Cytotoxic Natural Killer Cells After Aneurysmal Subarachnoid Hemorrhage. *World Neurosurg*. 2017;101:666-76.e1.
85. Xu T, Wang W, Zhai L, et al. Serum Gamma-glutamyl Transferase Levels Predict Functional Outcomes after Aneurysmal Subarachnoid Hemorrhage. *Biomed Environ Sci*. 2017;30(3):170-6.
86. Fukuda H, Lo B, Handa A, Yamamoto Y, Kurosaki Y, Yamagata S. Plasma D-dimer may predict poor functional outcomes through systemic complications after aneurysmal subarachnoid hemorrhage. *J Neurosurg*. 2017;127(2):284-90.
87. Li WH, Hui CJ, Ju H. Expression and significance of vWF, GMP-140 and ADAMTS13 in patients with aneurysmal subarachnoid hemorrhage. *European review for medical and pharmacological sciences*. 2017;21(19):4350-6.
88. Lu G, Wong MS, Xiong MZQ, et al. Circulating MicroRNAs in Delayed Cerebral Infarction After Aneurysmal Subarachnoid Hemorrhage. *J Am Heart Assoc*. 2017;6(4):25.
89. Ewelina G, Krzysztof S, Marek M, Krzysztof K. Blood free Radicals Concentration Determined by Electron Paramagnetic Resonance Spectroscopy and Delayed Cerebral Ischemia Occurrence in Patients with Aneurysmal Subarachnoid Hemorrhage. *Cell Biochem Biophys*. 2017;75(3-4):351-8.
90. Srinivasan A, Aggarwal A, Gaudihalli S, et al. Impact of Early Leukocytosis and Elevated High-Sensitivity C-Reactive Protein on Delayed Cerebral Ischemia and Neurologic Outcome after Subarachnoid Hemorrhage. *World Neurosurg*. 2016;90:91-5.
91. Ding YS, Sun B, Jiang JX, Zhang Q, Lu J, Gao GZ. Increased serum concentrations of signal peptide-Cub-Egf domain-containing protein-1 in patients with aneurysmal subarachnoid hemorrhage. *Clin Chim Acta*. 2016;459:117-22.
92. Jiang L, Wang WH, Dong XQ, et al. The change of plasma pituitary adenylate cyclase-activating polypeptide levels after aneurysmal subarachnoid hemorrhage. *Acta Neurol Scand*. 2016;134(2):131-9.
93. Rasmussen R, Stavngaard T, Jessing IR, et al. High plasma levels of neuropeptide y correlate with good clinical outcome but are not correlated to cerebral blood flow or vasospasm after subarachnoid hemorrhage. *J Neurosurg Anesthesiol*. 2016;28(1):65-70.

94. Mijiti M, Mijiti P, Axier A, et al. Incidence and predictors of angiographic vasospasm, symptomatic vasospasm and cerebral infarction in chinese patients with aneurysmal subarachnoid hemorrhage. *PLoS one*. 2016;11 (12) (no pagination)(e0168657):e0168657.
95. Donkelaar CEv, Bakker NA, Veeger NJGM, et al. Prediction of outcome after subarachnoid hemorrhage: timing of clinical assessment. *J Neurosurg*. 2017;126(1):52.
96. Behrouz R, Godoy DA, Topel CH, et al. Early Hypoalbuminemia is an Independent Predictor of Mortality in Aneurysmal Subarachnoid Hemorrhage. *Neurocrit Care*. 2016;25(2):230-6.
97. Wu W, Guan Y, Zhao G, et al. Elevated IL-6 and TNF-alpha Levels in Cerebrospinal Fluid of Subarachnoid Hemorrhage Patients. *Mol Neurobiol*. 2016;53(5):3277-85.
98. Donnelly MK, Conley YP, Crago EA, et al. Genetic markers in the EET metabolic pathway are associated with outcomes in patients with aneurysmal subarachnoid hemorrhage. *J Cereb Blood Flow Metab*. 2015;35(2):267-76.
99. Donnelly MK, Crago EA, Conley YP, et al. 20-HETE is associated with unfavorable outcomes in subarachnoid hemorrhage patients. *J Cereb Blood Flow Metab*. 2015;35(9):1515-22.
100. Chen X-D, Sun J, Lu C, et al. The prognostic value of plasma soluble CD40 ligand levels following aneurysmal subarachnoid hemorrhage. *Thromb Res*. 2015;136(1):24-9.
101. Shen YF, Wang WH, Yu WH, et al. The prognostic value of plasma thrombospondin-1 concentrations after aneurysmal subarachnoid hemorrhage. *Clin Chim Acta*. 2015;448:155-60.
102. Crago EA, Sherwood PR, Bender C, Balzer J, Ren D, Poloyac SM. Plasma Estrogen Levels Are Associated With Severity of Injury and Outcomes After Aneurysmal Subarachnoid Hemorrhage. *Biol Res Nurs*. 2015;17(5):558-66.
103. Helbok R, Schiefecker AJ, Beer R, et al. Early brain injury after aneurysmal subarachnoid hemorrhage: A multimodal neuromonitoring study. *Crit Care*. 2015;19 (1) (no pagination)(75):75.
104. Kofler M, Schiefecker A, Ferger B, et al. Cerebral Taurine Levels are Associated with Brain Edema and Delayed Cerebral Infarction in Patients with Aneurysmal Subarachnoid Hemorrhage. *Neurocrit Care*. 2015;23(3):321-9.
105. Leclerc JL, Blackburn S, Neal D, et al. Haptoglobin phenotype predicts the development of focal and global cerebral vasospasm and may influence outcomes after aneurysmal subarachnoid hemorrhage. *Proc Natl Acad Sci USA*. 2015;112(4):1155-60.
106. Sun J, Tan G, Xing W, He Z. Optimal hemoglobin concentration in patients with aneurysmal subarachnoid hemorrhage after surgical treatment to prevent symptomatic cerebral vasospasm. *Neuroreport*. 2015;26(5):263-6.
107. Tang Q-F, Lu S-Q, Zhao Y-M, Qian J-X. The changes of von willebrand factor/a disintegrin-like and metalloprotease with thrombospondin type I repeats-13 balance in aneurysmal subarachnoid hemorrhage. *Int J Clin Exp*. 2015;8(1):1342.
108. Suzuki H, Kanamaru K, Shiba M, et al. Tenascin-C is a possible mediator between initial brain injury and vasospasm-related and -unrelated delayed cerebral ischemia after aneurysmal subarachnoid hemorrhage. *Acta Neurochir Suppl*. 2015;120:117-21.
109. Helbok R, Schiefecker A, Delazer M, et al. Cerebral tau is elevated after aneurysmal subarachnoid haemorrhage and associated with brain metabolic distress and poor functional and cognitive long-term outcome. *J Neurol Neurosurg Psychiatry*. 2015;86(1):79-86.
110. Cai JY, Chen XD, Ba HJ, et al. Identification of plasma adrenomedullin as a possible prognostic biomarker for aneurysmal subarachnoid hemorrhage. *Peptides*. 2014;59:9-13.
111. Martini RP, Ward J, Siler DA, et al. Genetic variation in soluble epoxide hydrolase: Association with outcome after aneurysmal subarachnoid hemorrhage. *J Neurosurg*. 2014;121(6):1359-66.
112. Bergstrom A, Staalso JM, Romner B, Olsen NV. Impaired endothelial function after aneurysmal subarachnoid haemorrhage correlates with arginine:asymmetric dimethylarginine ratio. *Br J Anaesth*. 2014;112(2):311-8.
113. Zanier ER, Zangari R, Munthe-Fog L, et al. Ficolin-3-mediated lectin complement pathway activation in patients with subarachnoid hemorrhage. *Neurology*. 2014;82(2):126-34.
114. Radolf S, Smoll N, Drenckhahn C, Dreier JP, Vajkoczy P, Sarrafzadeh AS. Cerebral Lactate Correlates with Early Onset Pneumonia after Aneurysmal SAH. *Transl Stroke Res*. 2014;5(2):278-85.

115. Gomes JA, Selim M, Cotleur A, et al. Brain Iron Metabolism and Brain Injury Following Subarachnoid Hemorrhage: iCeFISH-Pilot (CSF Iron in SAH). *Neurocrit Care*. 2014;21(2):285-93.
116. Besegluglu K, Etminan N, Steiger HJ, Hanggi D. The relation of early hypernatremia with clinical outcome in patients suffering from aneurysmal subarachnoid hemorrhage. *Clin Neurol Neurosurg*. 2014;123:164-8.
117. Uekusa H, Miyazaki C, Kondo K, et al. Hydroperoxide in internal jugular venous blood reflects occurrence of subarachnoid hemorrhage-induced delayed cerebral vasospasm. *J Stroke Cerebrovasc Dis*. 2014;23(9):2217-24.
118. de Rooij NK, Greving JP, Rinkel GJ, Frijns CJ. Early prediction of delayed cerebral ischemia after subarachnoid hemorrhage: development and validation of a practical risk chart. *Stroke*. 2013;44(5):1288-94.
119. Bian L, Liu L, Wang C, et al. Hyperglycemia within day 14 of aneurysmal subarachnoid hemorrhage predicts 1-year mortality. *Clin Neurol Neurosurg*. 2013;115(7):959-64.
120. Gallek M, Alexander S, Crago E, et al. Endothelin-1 and endothelin receptor gene variants and their association with negative outcomes following aneurysmal subarachnoid hemorrhage. *Biol Res Nurs*. 2013;15(4):390-7.
121. Nakagawa I, Hironaka Y, Nishimura F, et al. Early inhibition of natriuresis suppresses symptomatic cerebral vasospasm in patients with aneurysmal subarachnoid hemorrhage. *Cerebrovasc Dis*. 2013;35(2):131-7.
122. Ohnishi H, Iihara K, Kaku Y, et al. Haptoglobin phenotype predicts cerebral vasospasm and clinical deterioration after aneurysmal subarachnoid hemorrhage. *J Stroke Cerebrovasc Dis*. 2013;22(4):520-6.
123. July J, Yunus Y, Sungono V, As'ad S, Suhadi B, Islam AA. Cortisol dynamics and endothelin-1/nitric oxide ratio are associated with clinical vasospasm. *Med J Indones*. 2013;22(3):161-6.
124. Barges-Coll J, Perez-Neri I, Avendano J, Mendez-Rosito D, Gomez-Amador JL, Rios C. Plasma taurine as a predictor of poor outcome in patients with mild neurological deficits after aneurysmal subarachnoid hemorrhage. *J Neurosurg*. 2013;119(4):1021-7.
125. Lanterna LA, Spreafico V, Gritti P, et al. Hypocortisolism in noncomatose patients during the acute phase of subarachnoid hemorrhage. *J Stroke Cerebrovasc Dis*. 2013;22(7):e189-e96.
126. Fischer M, Dietmann A, Beer R, et al. Differential Regulation of Matrix-Metalloproteinases and Their Tissue Inhibitors in Patients with Aneurysmal Subarachnoid Hemorrhage. *PloS one*. 2013;8(3) (no pagination):e59952:e59952.
127. Kim H, Crago E, Kim M, et al. Cerebral vasospasm after sub-arachnoid hemorrhage as a clinical predictor and phenotype for genetic association study. *International Journal of Stroke*. 2013;8(8):620-5.
128. Aggarwal A, Salunke P, Singh H, et al. Vasospasm following aneurysmal subarachnoid hemorrhage: Thrombocytopenia a marker. *J Neurosci Rural Pract*. 2013;4(3):257-61.
129. Maimaitili A, Maimaitili M, Rexidan A, et al. Pituitary hormone level changes and hyponatremia in aneurysmal subarachnoid hemorrhage. *Exp Ther Med*. 2013;5(6):1657-62.
130. Zanier ER, Zoerle T, Fiorini M, et al. Heart-fatty acid-binding and tau proteins relate to brain injury severity and long-term outcome in subarachnoid haemorrhage patients. *Br J Anaesth*. 2013;111(3):424-32.
131. Nyquist PA, Wang H, Suffredini AF. Protein biomarkers in patients with subarachnoid hemorrhage, vasospasm, and delayed ischemic neurological deficits. *Cerebral Vasospasm: Neurovascular Events After Subarachnoid Hemorrhage*. 2013;Acta Neurochirurgica, Supplementum. 115:23-5.
132. Juvela S, Kuhmonen J, Siironen J. C-reactive protein as predictor for poor outcome after aneurysmal subarachnoid haemorrhage. *Acta Neurochir (Wien)*. 2012;154(3):397-404.
133. Papanikolaou J, Makris D, Karakitsos D, et al. Cardiac and central vascular functional alterations in the acute phase of aneurysmal subarachnoid hemorrhage. *Crit Care Med*. 2012;40(1):223-32.

134. Yang TM, Lin YJ, Tsai NW, et al. The prognostic value of serial leukocyte adhesion molecules in post-aneurysmal subarachnoid hemorrhage. *Clin Chim Acta*. 2012;413(3-4):411-6.
135. Zhu XD, Chen JS, Zhou F, Liu QC, Chen G, Zhang JM. Relationship between plasma high mobility group box-1 protein levels and clinical outcomes of aneurysmal subarachnoid hemorrhage. *J Neuroinflammation*. 2012;9 (no pagination)(194):194.
136. Jeon YT, Lee JH, Lee H, et al. The postoperative C-reactive protein level can be a useful prognostic factor for poor outcome and symptomatic vasospasm in patients with aneurysmal subarachnoid hemorrhage. *J Neurosurg Anesthesiol*. 2012;24(4):317-24.
137. Vrsajkov V, Javanovic G, Stanisavljevic S, Uvelin A, Vrsajkov JP. Clinical and predictive significance of hyponatremia after aneurysmal subarachnoid hemorrhage. *Balkan Medical Journal*. 2012;29(3):243-6.
138. Watanabe A, Tagami T, Yokobori S, et al. Global end-diastolic volume is associated with the occurrence of delayed cerebral ischemia and pulmonary edema after subarachnoid hemorrhage. *Shock*. 2012;38(5):480-5.
139. Jung CS, Lange B, Zimmermann M, Seifert V. The CSF concentration of ADMA, but not of ET-1, is correlated with the occurrence and severity of cerebral vasospasm after subarachnoid hemorrhage. *Neurosci Lett*. 2012;524(1):20-4.
140. Sarrafzadeh A, Copin JC, Bengualid DJ, et al. Matrix metalloproteinase-9 concentration in the cerebral extracellular fluid of patients during the acute phase of aneurysmal subarachnoid hemorrhage. *Neurol Res*. 2012;34(5):455-61.
141. Beeftink MM, Ruigrok YM, Rinkel GJ, van den Bergh WM. Relation of serum TNF-alpha and TNF-alpha genotype with delayed cerebral ischemia and outcome in subarachnoid hemorrhage. *Neurocrit Care*. 2011;15(3):405-9.
142. Zanier ER, Brandi G, Peri G, et al. Cerebrospinal fluid pentraxin 3 early after subarachnoid hemorrhage is associated with vasospasm. *Intensive Care Medicine*. 2011;37(2):302-9.
143. Dietmann A, Lackner P, Fischer M, et al. Soluble Endoglin and Transforming Growth Factor-beta<sup>1</sup> and the Development of Vasospasm after Spontaneous Subarachnoid Hemorrhage: A Pilot Study. *Cerebrovasc Dis*. 2011;33(1):16-22.
144. Fischer M, Broessner G, Dietmann A, et al. Angiopoietin-1 is associated with cerebral vasospasm and delayed cerebral ischemia in subarachnoid hemorrhage. *BMC Neurol*. 2011;11 (no pagination)(59):59.
145. Zheng B, Qiu Y, Jin H, et al. A predictive value of hyponatremia for poor outcome and cerebral infarction in high-grade aneurysmal subarachnoid haemorrhage patients. *J Neurol Neurosurg Psychiatry*. 2011;82(2):213-7.
146. Zhu XD, Chen JS, Zhou F, Liu QC, Chen G, Zhang JM. Detection of copeptin in peripheral blood of patients with aneurysmal subarachnoid hemorrhage. *Crit Care*. 2011;15 (6) (no pagination)(R288):R288.
147. Nakae R, Yokota H, Yoshida D, Teramoto A. Transcranial Doppler ultrasonography for diagnosis of cerebral vasospasm after aneurysmal subarachnoid hemorrhage: mean blood flow velocity ratio of the ipsilateral and contralateral middle cerebral arteries. *Neurosurgery*. 2011;69(4):876-83; discussion 83.
148. Taub PR, Fields JD, Wu AH, et al. Elevated BNP is associated with vasospasm-independent cerebral infarction following aneurysmal subarachnoid hemorrhage. *Neurocrit Care*. 2011;15(1):13-8.
149. Zanier ER, Refai D, Zipfel GJ, et al. Neurofilament light chain levels in ventricular cerebrospinal fluid after acute aneurysmal subarachnoid haemorrhage. *J Neurol Neurosurg Psychiatry*. 2011;82(2):157-9.
150. Hirashima Y, Doshi M, Hayashi N, et al. Plasma platelet activating factor-acetylhydrolase activity and the levels of free forms of bio-marker of lipid peroxidation in cerebrospinal fluid of patients with aneurysmal subarachnoid hemorrhage. *Neurosurgery*. 2011;19:602-9.
151. Wang HC, Lin WC, Yang TM, et al. The association between symptomatic delayed cerebral infarction and serum adhesion molecules in aneurysmal subarachnoid hemorrhage. *Neurosurgery*. 2011;68(6):1611-7.

152. Naidech AM, Shaibani A, Garg RK, et al. Prospective, randomized trial of higher goal hemoglobin after subarachnoid hemorrhage. *Neurocrit Care*. 2010;13(3):313-20.
153. Lackner P, Dietmann A, Beer R, et al. Cellular microparticles as a marker for cerebral vasospasm in spontaneous subarachnoid hemorrhage. *Stroke*. 2010;41(10):2353-7.
154. Kasius KM, Frijns CJ, Algra A, Rinkel GJ. Association of platelet and leukocyte counts with delayed cerebral ischemia in aneurysmal subarachnoid hemorrhage. *Cerebrovascular Diseases*. 2010;29(6):576-83.
155. Ruigrok YM, Slooter AJ, Rinkel GJ, Wijmenga C, Rosendaal FR. Genes influencing coagulation and the risk of aneurysmal subarachnoid hemorrhage, and subsequent complications of secondary cerebral ischemia and rebleeding. *Acta Neurochir (Wien)*. 2010;152(2):257-62.
156. Vergouwen MD, van Geloven N, de Haan RJ, Kruijff ND, Vermeulen M, Roos YB. Increased cortisol levels are associated with delayed cerebral ischemia after aneurysmal subarachnoid hemorrhage. *Neurocrit Care*. 2010;12(3):342-5.
157. Juvela S, Siironen J, Lappalainen J. Apolipoprotein E genotype and outcome after aneurysmal subarachnoid hemorrhage: Clinical article. *J Neurosurg*. 2009;110(5):989-95.
158. Vergouwen MD, Bakhtiari K, van Geloven N, Vermeulen M, Roos YB, Meijers JC. Reduced ADAMTS13 activity in delayed cerebral ischemia after aneurysmal subarachnoid hemorrhage. *J Cereb Blood Flow Metab*. 2009;29(10):1734-41.
159. Oddo M, Milby A, Chen I, et al. Hemoglobin concentration and cerebral metabolism in patients with aneurysmal subarachnoid hemorrhage. *Stroke*. 2009;40(4):1275-81.
160. Kruijff ND, Roos YW, Dorhout Mees SM, et al. High mean fasting glucose levels independently predict poor outcome and delayed cerebral ischaemia after aneurysmal subarachnoid haemorrhage. *J Neurol Neurosurg Psychiatry*. 2008;79(12):1382-5.
161. van den Bergh WM, van de Water JM, Hoff RG, Algra A, Rinkel GJ. Calcium homeostasis during magnesium treatment in aneurysmal subarachnoid hemorrhage. *Neurocrit Care*. 2008;8(3):413-7.
162. Starke RM, Kim GH, Komotar RJ, et al. Endothelial nitric oxide synthase gene single-nucleotide polymorphism predicts cerebral vasospasm after aneurysmal subarachnoid hemorrhage. *J Cereb Blood Flow Metab*. 2008;28(6):1204-11.
163. Lewis SB, Wolper RA, Miraliam L, Yang C, Shaw G. Detection of phosphorylated NF-H in the cerebrospinal fluid and blood of aneurysmal subarachnoid hemorrhage patients. *J Cereb Blood Flow Metab*. 2008;28(6):1261-71.
164. Isman F, Kucur M, Tanriverdi T, et al. Serum hyaluronidase levels in patients with aneurysmal subarachnoid haemorrhage. *Singapore Med J*. 2008;49(5):405-9.
165. Dorhout Mees SM, van den Bergh WM, Algra A, Rinkel GJ. Achieved serum magnesium concentrations and occurrence of delayed cerebral ischaemia and poor outcome in aneurysmal subarachnoid haemorrhage. *J Neurol Neurosurg Psychiatry*. 2007;78(7):729-31.
166. Tseng MY, Hutchinson PJ, Turner CL, et al. Biological effects of acute pravastatin treatment in patients after aneurysmal subarachnoid hemorrhage: A double-blind, placebo-controlled trial. *J Neurosurg*. 2007;107(6):1092-100.
167. Igarashi T, Moro N, Katayama Y, Mori T, Kojima J, Kawamata T. Prediction of symptomatic cerebral vasospasm in patients with aneurysmal subarachnoid hemorrhage: Relationship to cerebral salt wasting syndrome. *Neurol Res*. 2007;29(8):835-41.
168. Oh SY, Kwon JT, Hong HJ, Kim JB, Suk JS. Relationship between leukocytosis and vasospasms following aneurysmal subarachnoid hemorrhage. *J Korean Neurosurg Soc*. 2007;41(3):153-6.
169. Schebesch KM, Woertgen C, Brawanski A, Rothoerl RD. A study of possible correlation between subarachnoid haemorrhage related vasospasm and the post-bleed blood platelet count chart in a Caucasian population. *Acta Neurochir (Wien)*. 2007;149(4):387-91.
170. Schoch B, Regel JP, Wichert M, Gasser T, Volbracht L, Stolke D. Analysis of intrathecal interleukin-6 as a potential predictive factor for vasospasm in subarachnoid hemorrhage. *Neurosurgery*. 2007;60(5):828-35.

171. Siironen J, Juvela S, Kanarek K, Vilkki J, Hernesniemi J, Lappalainen J. The Met allele of the BDNF Val66Met polymorphism predicts poor outcome among survivors of aneurysmal subarachnoid hemorrhage. *Stroke*. 2007;38(10):2858-60.
172. Lewis SB, Velat GJ, Miralia L, et al. Alpha-II spectrin breakdown products in aneurysmal subarachnoid hemorrhage: a novel biomarker of proteolytic injury. *J Neurosurg*. 2007;107(4):792-6.
173. Juvela S, Siironen J. D-dimer as an independent predictor for poor outcome after aneurysmal subarachnoid hemorrhage. *Stroke*. 2006;37(6):1451-6.
174. Rothoerl RD, Schebesch KM, Kubitzka M, Woertgen C, Brawanski A, Pina AL. ICAM-1 and VCAM-1 expression following aneurysmal subarachnoid hemorrhage and their possible role in the pathophysiology of subsequent ischemic deficits. *Cerebrovasc Dis*. 2006;22(2-3):143-9.
175. Rothoerl RD, Axmann C, Pina AL, Woertgen C, Brawanski A. Possible role of the C-reactive protein and white blood cell count in the pathogenesis of cerebral vasospasm following aneurysmal subarachnoid hemorrhage. *J Neurosurg Anesthesiol*. 2006;18(1):68-72.
176. Frijns CJ, Fijnheer R, Algra A, van Mourik JA, van Gijn J, Rinkel GJ. Early circulating levels of endothelial cell activation markers in aneurysmal subarachnoid haemorrhage: associations with cerebral ischaemic events and outcome. *J Neurol Neurosurg Psychiatry*. 2006;77(1):77-83.
177. Frijns CJ, Kasius KM, Algra A, Fijnheer R, Rinkel GJ. Endothelial cell activation markers and delayed cerebral ischaemia in patients with subarachnoid haemorrhage. *J Neurol Neurosurg Psychiatry*. 2006;77(7):863-7.
178. Oertel M, Schumacher U, McArthur DL, Kastner S, Boker DK. S-100B and NSE: markers of initial impact of subarachnoid haemorrhage and their relation to vasospasm and outcome. *J Clin Neurosci*. 2006;13(8):834-40.
179. Lanterna LA, Rigoldi M, Tredici G, et al. APOE influences vasospasm and cognition of noncomatose patients with subarachnoid hemorrhage. *Neurology*. 2005;64(7):1238-44.
180. Kastner S, Oertel MF, Scharbrodt W, Krause M, Boker DK, Deinsberger W. Endothelin-1 in plasma, cisternal CSF and microdialysate following aneurysmal SAH. *Acta Neurochir (Wien)*. 2005;147(12):1271-9.
181. Asaeda M, Sakamoto M, Kurosaki M, et al. A non-enzymatic derived arachidonyl peroxide, 8-iso-prostaglandin F  $\alpha_2$ , in cerebrospinal fluid of patients with aneurysmal subarachnoid hemorrhage participates in the pathogenesis of delayed cerebral vasospasm. *Neurosci Lett*. 2005;373(3):222-5.
182. Dohi K, Ripley B, Fujiki N, et al. CSF hypocretin-1/orexin-A concentrations in patients with subarachnoid hemorrhage (SAH). *Peptides*. 2005;26(11):2339-43.
183. Tanriverdi T, Sanus GZ, Ulu MO, et al. Serum and cerebrospinal fluid concentrations of E-selectin in patients with aneurysmal subarachnoid hemorrhage. *Braz J Med Biol*. 2005;38(11):1703-10.
184. Juvela S, Siironen J, Kuhmonen J. Hyperglycemia, excess weight, and history of hypertension as risk factors for poor outcome and cerebral infarction after aneurysmal subarachnoid hemorrhage. *J Neurosurg*. 2005;102(6):998-1003.
185. Khurana VG, Sohni YR, Mangrum WI, et al. Endothelial nitric oxide synthase gene polymorphisms predict susceptibility to aneurysmal subarachnoid hemorrhage and cerebral vasospasm. *J Cereb Blood Flow Metab*. 2004;24(3):291-7.
186. McGirt MJ, Blessing R, Nimjee SM, et al. Correlation of serum brain natriuretic peptide with hyponatremia and delayed ischemic neurological deficits after subarachnoid hemorrhage. *Neurosurgery*. 2004;54(6):1369-73; discussion 73-4.
187. Sarrafzadeh AS, Haux D, Ludemann L, et al. Cerebral ischemia in aneurysmal subarachnoid hemorrhage: a correlative microdialysis-PET study. *Stroke*. 2004;35(3):638-43.
188. Collignon FP, Friedman JA, Piegras DG, et al. Serum magnesium levels as related to symptomatic vasospasm and outcome following aneurysmal subarachnoid hemorrhage. *Neurocrit Care*. 2004;1(4):441-8.
189. Hendryk S, Jarzab B, Josko J. Increase of the IL-1 beta and IL-6 levels in CSF in patients with vasospasm following aneurysmal SAH. *Neuroendocrinol Lett*. 2004;25(1-2):141-7.

190. van den Bergh WM, Algra A, van der Sprenkel JW, Tulleken CA, Rinkel GJ. Hypomagnesemia after aneurysmal subarachnoid hemorrhage. *Neurosurgery*. 2003;52(2):276-81; discussion 81-2.
191. Qureshi AI, Suri MF, Sung GY, et al. Prognostic significance of hypernatremia and hyponatremia among patients with aneurysmal subarachnoid hemorrhage. *Neurosurgery*. 2002;50(4):749-55; discussion 55-6.
192. Juvela S. Plasma endothelin and big endothelin concentrations and serum endothelin-converting enzyme activity following aneurysmal subarachnoid hemorrhage. *J Neurosurg*. 2002;97(6):1287-93.
193. Nissen JJ, Mantle D, Gregson B, Mendelow AD. Serum concentration of adhesion molecules in patients with delayed ischaemic neurological deficit after aneurysmal subarachnoid haemorrhage: The immunoglobulin and selectin superfamilies. *J Neurol Neurosurg Psychiatry*. 2001;71(3):329-33.
194. Mascia L, Fedorko L, Stewart DJ, et al. Temporal relationship between endothelin-1 concentrations and cerebral vasospasm in patients with aneurysmal subarachnoid hemorrhage. *Stroke*. 2001;32(5):1185-9.
195. Hirashima Y, Endo S, Nakamura S, Kurimoto M, Takaku A. Cerebrospinal fluid membrane-bound tissue factor and myelin basic protein in the course of vasospasm after subarachnoid hemorrhage. *Neurol Res*. 2001;23(7):715-20.
196. Nam DH, Kim JS, Hong SC, et al. Expression of interleukin-1beta in lipopolysaccharide stimulated monocytes derived from patients with aneurysmal subarachnoid hemorrhage is correlated with cerebral vasospasm. *Neurosci Lett*. 2001;312(1):41-4.
197. Juvela S. Plasma endothelin concentrations after aneurysmal subarachnoid hemorrhage. *J Neurosurg*. 2000;92(3):390-400.
198. Gruber A, Roessler K, Georgopoulos A, Missbichler A, Bonelli R, Richling B. Evaluation of big endothelin-1 concentrations in serum and ventricular cerebrospinal fluid after early surgical compared with nonsurgical management of ruptured intracranial aneurysms. *Neurosurg Focus*. 2000;8(5):e6.
199. Takenaka KV, Sakai N, Murase S, Kuroda T, Okumura A, Sawada M. Elevated transferrin concentration in cerebral spinal fluid after subarachnoid hemorrhage. *Neurol Res*. 2000;22(8):797-801.
200. Mayer SA, Lin J, Homma S, et al. Myocardial injury and left ventricular performance after subarachnoid hemorrhage. *Stroke*. 1999;30(4):780-6.
201. Fujii Y, Takeuchi S, Sasaki O, Minakawa T, Koike T, Tanaka R. Serial changes of hemostasis in aneurysmal subarachnoid hemorrhage with special reference to delayed ischemic neurological deficits. *Journal of neurosurgery*. 1997;86(4):594-602.
202. Hirashima Y, Nakamura S, Endo S, Kuwayama N, Naruse Y, Takaku A. Elevation of Platelet Activating Factor, Inflammatory Cytokines, and Coagulation Factors in the Internal Jugular Vein of Patients with Subarachnoid Hemorrhage. *Neurochem Res*. 1997;22(10):1249-55.
203. Hirashima Y, Nakamura S, Suzuki M, et al. Cerebrospinal fluid tissue factor and thrombin-antithrombin III complex as indicators of tissue injury after subarachnoid hemorrhage. *Stroke*. 1997;28(9):1666-70.
204. Okuchi K, Fujioka M, Fujikawa A, et al. Rapid natriuresis and preventive hypervolaemia for symptomatic vasospasm after subarachnoid haemorrhage. *Acta Neurochir (Wien)*. 1996;138(8):951-7.
205. Hirashima Y, Kurimoto M, Tsukamoto E, Endo S, Takaku A. Anti-phospholipid antibodies and cerebral vasospasm following subarachnoid haemorrhage. *Acta Neurochir (Wien)*. 1995;135(3-4):191-7.
206. Hirashima Y, Endo S, Kurimoto M, Tsukamoto E, Takaku A. Platelet-activating factor and antiphospholipid antibodies in subarachnoid haemorrhage. *Acta Neurochir (Wien)*. 1994;128(1-4):144-9.
207. Hirashima Y, Endo S, Ohmori T, Kato R, Takaku A. Platelet-activating factor (PAF) concentration and PAF acetylhydrolase activity in cerebrospinal fluid of patients with subarachnoid hemorrhage. *J Neurosurg*. 1994;80(1):31-6.

208. Suzuki Y, Sato S, Suzuki H, et al. Increased neuropeptide Y concentrations in cerebrospinal fluid from patients with aneurysmal subarachnoid hemorrhage. *Stroke*. 1989;20(12):1680-4.
209. Rodriguez y Baena R, Gaetani P, Paoletti P. A study on cisternal CSF levels of arachidonic acid metabolites after aneurysmal subarachnoid hemorrhage. *J Neurol Sci*. 1988;84(2-3):329-35.